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Rheology of mixed alginate-hyaluronan aqueous solutions

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25 **Abstract**

26 The present manuscript addresses the description of binary systems of hyaluronan (HA) and
27 alginate (Alg) in semi-concentrated solution. The two polysaccharides were completely miscible in
28 the entire range of relative weight fraction explored at a total polymer concentration of up to 3 %
29 (w/V). The rheological study encompassed steady flow and mechanical spectra for HA/Alg systems
30 at different weight fractions with hyaluronan at different molecular weights. These extensive
31 analyses allowed us to propose a model for the molecular arrangement in solution that envisages a
32 mutual exclusion between the two polysaccharides even though a clear phase separation does not
33 occur. This result may have profound implications when biomaterials based on the combination of
34 alginate and hyaluronan are proposed in the field of biomedical materials.

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1. Introduction

Alginate is a collective term to describe a family of polysaccharides isolated from brown seaweeds and bacteria [1]. Chemically, alginates are linear copolymers of 1→4 linked β-D-mannuronic acid (M) and its C-5 epimer, α-L-guluronic acid (G), arranged in a blockwise pattern, with homopolymeric regions of M- and G residues, indicated as M-blocks and G-blocks, respectively, interspersed with regions of alternating structure (MG blocks). At around neutral pH, alginate behaves as an anionic polyelectrolyte and as a semi-flexible chain upon reduction of the medium ionic strength. The ability to form stable hydrogels upon treatment with divalent ions, such as calcium ions [2], have resulted in alginates being widely used in industrial applications. In addition, alginates have relevant applications in the field of biomedical materials [3,4] due to their versatility in gel formation which has been exploited for the encapsulation of, for example, the insulin-producing Langerhans islets for the treatment of type I diabetes [5-8]. In fact, the availability of pure and well-characterised alginate samples makes this material an ideal choice for cell microencapsulation.

In addition to cell encapsulation, alginate has been proposed for several other applications in the biomedical field [3]. For example, alginate-based scaffolds containing hydroxyapatite showed structural features similar to those of the trabecular bone [9-12], and in combination with silver nanoparticles, as well as having antimicrobial properties, have shown good osteointegration in *in vivo* models [12].

However, it should be underlined that unmodified alginate shows non-adhesive properties and, as such, it does not favour cell adhesion [13], which is the first step towards cell proliferation and colonisation. For this reason, the addition of bioactive polymers to alginate has been considered as a way to improve its features [14,15].

Hyaluronan (HA), an alternating copolymer composed of β-D-glucuronic acid (GlcA) and β-D-N-acetylglucosamine (GlcNAc), linked in the sequence →4 GlcA1→3 GlcNAc1→ is the simplest

glycosaminoglycan and a major constituent of the extracellular matrix (ECM) as a high molar mass anionic polyelectrolyte component [16]. Apart from its well-known structural role due to the peculiar chain semi-flexibility, which gives the polymer exceptional viscoelastic behaviour, hyaluronan shows important biological features. HA has six known surface receptors [17], among which CD44 and RHAMM are the most well characterised. While both receptors bind HA through a common binding domain, they differ, to some extent, in the cellular functions: CD44 mediates cell attachment, HA uptake and degradation while RHAMM mediates cell locomotion in response to soluble HA [18].

The idea of exploiting mixtures of alginate and hyaluronan to develop novel biomaterials has been pursued by several authors for mesenchymal stem cell encapsulation [19], as a post-surgical tissue adhesion barrier [20] and for the encapsulation of chondrocytes [21]. The combination of alginate and hyaluronan has also been proposed for cartilage engineering by Lindenhayn and co-workers [22]. In this paper, it was noted that hyaluronan was, to some extent and depending on the alginate gelation conditions, excluded from the hydrogel network. In addition, Park *et al.* prepared hyaluronan beads starting from its mixture with alginate and exploiting the ability of the latter to form “reversible” calcium hydrogels [23]. Most of the above mentioned papers focused on the biological properties exerted by the mixed construct, but they overlook the physical-chemical implications incidental to the presence of two anionic polyelectrolytes in solution, which might impact properties such as gel formation kinetics, stability and mechanical performance.

2. Materials and Methods

Alginate (Alg) (LVG, $M_W = 120000$; $F_G = 0.69$; $F_{GG} = 0.59$; $N_{G>1} = 16.3$; $PI = 1.9$) and hyaluronan (HA) with M_W of 1500000 (HA150; $PI = 2.2$) and of 800000 (HA80; $PI = 2.4$) were kindly provided by FMC, Drammen (Norway). Hyaluronan with a M_W of 240000 (HA24) was kindly provided by Sigea S.r.L., Trieste (Italy). Hyaluronan with M_W of 400000 (HA40; $PI = 2.7$) was obtained by acid degradation of HA150 according to a procedure previously reported [24] and the molar mass was evaluated from intrinsic viscosity measurements [25]. In alginate characterization, F_G denotes the fraction of alginate consisting of guluronic acid, F_{GG} indicates the fraction of alginate consisting of guluronic acid in blocks and $N_{G>1}$ denotes the average length of guluronic acid blocks. Binary mixtures of HA and Alg were prepared in the presence of aqueous 0.15 M NaCl at a different weight ratio of HA (ϕ_{HA}) and of alginate ($\phi_{Alg} = 1 - \phi_{HA}$).

2.1 Viscometry

Specific viscosity was measured at 25 °C by means of a Schott-Geräte AVS/G automatic measuring apparatus and an Ubbelohde-type capillary viscometer in aqueous 0.15 M NaCl. For the polysaccharides used in the present study, the intrinsic viscosity ($[\eta]$) values were determined by analyzing the polymer concentration dependence of the reduced specific viscosity (η_{sp}/c) and of the logarithm of the reduced relative viscosity ($\ln(\eta_{rel})/c$) by means of the Huggins (eq. 1) and Kraemer (eq. 2) equations, respectively.

$$\frac{\eta_{sp}}{c} = [\eta] + k'[\eta]c \quad (1)$$

$$\frac{\ln(\eta_{rel})}{c} = [\eta] - k''[\eta]c \quad (2)$$

121 where k' and k'' are the Huggins and Kraemer constants, respectively. The specific viscosity of the
122 Alg/HA mixed solutions was measured in aqueous 0.15 M NaCl at different polysaccharide weight
123 fractions maintaining a constant total polymer concentration of 1 g/L.

124

125 2.2 Rheological determination

126 Rheological tests were performed on mixed solutions of alginate and hyaluronan under continuous
127 shear conditions to determine steady viscosity values in the stress range 1 to 100 Pa, as well as
128 under oscillatory shear conditions to determine the extension of the linear viscoelasticity regime
129 (stress sweep tests at 1 Hz in the stress range $1 < \tau < 500$ Pa) and the mechanical spectra (frequency
130 sweep, $\tau = 4$ Pa, within the linear viscoelastic range). The complex viscosity (η^*), the storage (G')
131 and loss (G'') moduli of the binary mixtures and of the hydrogels were recorded in the frequency
132 range 0.1 – 50 Hz. All tests were carried out with the controlled stress rheometer Rheostress Haake
133 RS 150 operating at 25 °C. A cone-plate CP60/1° geometry was used in all cases. A glass bell
134 covering the measuring device was used to improve thermal control and limit evaporation.

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3. Results and Discussion

3.1 Viscosity in dilute and semi-dilute conditions of binary Alg/HA mixtures

The (macroscopic) mutual compatibility of alginate (Alg) and hyaluronan in dilute binary solution was assessed by both transmittance measurements and viscometry. In the presence of aqueous NaCl (0.15 M), the former technique showed no notable variation in the transmittance of the binary system with respect to that displayed by the polysaccharides alone, thereby excluding the formation of macroscopic phase separation (data not reported). The complete macroscopic miscibility of the two polysaccharides was confirmed by the dependence of the specific viscosity of the HA/Alg dilute binary mixtures on their composition (Figure 1).

Figure 1

It can be noticed that the experimental results are in very good agreement with the theoretical prediction that was calculated assuming that no other interaction, apart from the hydrodynamic ones, occurred between the two polysaccharides [26].

The characterisation of HA/Alg binary mixtures was carried out in semi-concentrated solutions by means of a rheological measurements. The flow curves of the systems under analysis were recorded in 0.15 M NaCl aqueous solution while varying the weight fraction (ϕ) of the two polysaccharides and the molecular weight of the hyaluronan samples. In all cases, a pseudo-plastic behavior was detected with a clear Newtonian plateau for low values of the shear stress applied. The experimental curves were fitted with a simplified version of the Cross equation (eq. 3):

$$\eta = \eta_{\dot{\gamma} \rightarrow \infty} + \frac{\eta_0 - \eta_{\dot{\gamma} \rightarrow \infty}}{1 + (k\dot{\gamma})^n} = \frac{\eta_0}{1 + (k\dot{\gamma})^n} \quad (3)$$

159 where $\eta_{\dot{\gamma} \rightarrow \infty}$, the viscosity at infinite shear rate, was set equal to zero. η_0 is the zero-shear viscosity,
 160 corresponding to the limiting Newtonian plateau for $\dot{\gamma} \rightarrow 0$, and k is the characteristic polymer
 161 relaxation time, often indicated with τ [27]. τ is the time associated with large-scale motion in the
 162 structure of the polymer. n is known as the Cross Rate Constant and it can be considered as a
 163 “*pseudoplasticity index*” [28]. The dependence of the zero-shear viscosity on the molecular weight
 164 of hyaluronan for both the polysaccharide alone and for binary HA/Alg mixtures is reported in
 165 Figure 2a, showing a power-law dependence ($\eta_0 \propto MW^\beta$) when the total polysaccharide
 166 concentration was maintained equal to 30 g/L.

168 **Figure 2**

169
 170 In particular, when hyaluronan alone was considered, the power-law dependence showed a scaling
 171 factor of approximately 3.1. This is close to the value (3.4) predicted [29] and reported for different
 172 flexible polysaccharides including hyaluronan [30] and alginate [31] in the semi-dilute regime.
 173 Upon addition of alginate to the system, the scaling factor β showed a non-linear decrease upon
 174 reduction of the fraction of HA in the mixture (Figure 2, inset). This result likely stemmed from the
 175 dilution of hyaluronan due to the addition of the second polysaccharide, *i.e.*, alginate. In fact, the β
 176 exponent decreases for all (saccharidic) polymers at lower concentrations, down to the value of 1 in
 177 very dilute conditions. The effect on overall viscosity of the binary system due to the presence of
 178 alginate was also evident from the zero-shear viscosity which was 36.2 Pa·s for $C_{HA}=15$ g/L and
 179 increased to 84.9 Pa·s in the $\phi_{HA} = 0.5$ mixture (still with $C_{HA}=15$ g/L).

180 In Figure 2b, the dependence of the relaxation time, τ , on the molecular weight of HA is reported
 181 for both hyaluronan alone and for HA/Alg binary mixtures. In the first case, a power-law of
 182 $\tau \propto MW^{3.5}$ was found, which is very close to the theoretically expected relationship (3.4) [32-34].

183 Similarly, in the case of the binary hyaluronan/alginate mixtures, the exponent of the power law
184 was 3.2, which is even closer to some reported experimental values of τ for HA (3.0) [32-34]. In
185 Figure 3a the dependence of the zero-shear viscosity for the binary Alg/HA mixtures on the weight
186 fraction of HA (ϕ_{HA}) is reported for HA at different values of the molecular weight.

187

188

Figure 3

189

190 It can be seen that in all cases, a scaling law holds, at least in the ϕ_{HA} range explored. In Figure 3b
191 and 3c, the dependence of the zero-shear viscosity and of the polymer relaxation time, τ , on the
192 concentration of HA is reported for both a hyaluronan solution alone and a HA/Alg binary mixture
193 with $\phi_{\text{HA}} = 0.50$. For both properties, the addition of alginate brings about an effect that differs from
194 that of the pure dilution with the solvent. This provides an indication of the arrangement between
195 the two components of the mixture at a molecular level. In fact, considering the onset of the shear-
196 thinning behavior as a progressive decrease in the steady-state entanglements density of the
197 hyaluronan chains [33], the higher τ in the presence of alginate seems to point to an effective
198 higher concentration of HA which might arise from a (at least partial) segregation, rather than its
199 homogeneous mixing with alginate.

200

201 3.2 Mechanical spectroscopy

202 The stress sweep curves for the samples were recorded and a type I behavior was found in all cases.
203 A typical curve is reported in Figure 4 together with the best fit of the experimental data according
204 to the Soskey-Winter equation (eq. 4).

205

206

$$G' = G'_0 \frac{1}{1 + (b\gamma)^n} \quad (4)$$

Where γ is the strain, G'_0 is the limiting value of the storage modulus for $\gamma \rightarrow 0$, while b and n are adjustable parameters. The critical strain, γ_c , marking the limit of the linear viscoelastic regime, was arbitrarily assumed to correspond to $G'/G'_0 = 0.95$. The limit of the linear viscoelastic response is reported in table 1.

Figure 4

Table 1

The mechanical response of hyaluronan alone and of its binary mixtures with alginate was recorded as a function of pulsation, ω ($= 2\pi\nu$). Depending on the molecular weight of hyaluronan, a transition from viscous ($G'' > G'$) to elastic ($G' > G''$) behaviour took place in the frequency range explored (Figure 5a, for the sample case of HA150) with a cross-over point.

Figure 5

The experimental data were fitted using a generalised Maxwell model composed of a sequence of elements in parallel (spring and dashpot). The frequency dependence of the viscous (G') and elastic (G'') response is then described with the following equations (eqs. 5 and 6):

$$G' = \sum_{i=1}^n G_i \frac{(\lambda_i \omega)^2}{1 + (\lambda_i \omega)^2} \quad (5)$$

$$G'' = \sum_{i=1}^n G_i \frac{\lambda_i \omega}{1 + (\lambda_i \omega)^2} \quad \text{with} \quad G_i = \eta_i / \lambda_i \quad (6)$$

where n is the number of Maxwell elements considered, G_i , η_i and λ_i represent the spring constant, the dashpot viscosity and the relaxation time of the i th Maxwell element, respectively [35]. The fitting of the experimental data was performed assuming that the relaxation times were not independent of each other but they were scaled by a factor 10. The number of Maxwell elements was selected, based on a statistical procedure, to minimise the product $\chi^2 N_p$, where χ^2 is the sum of the squared errors, while $N_p (= 2 + n)$ indicates the number of fitting parameters (table 2).

Table 2

Figure 5b reports the dependence of the frequency corresponding to the cross-over point, $\omega_{\text{cross}} (= 2\pi\nu_{\text{cross}})$, on the molecular weight of hyaluronan for both HA alone and for its binary mixtures with alginate. In both cases, a very similar power-law relation holds for HA alone and for the HA/Alg mixture, namely $\omega_{\text{cross}} \propto MW^{-3.1}$ and $\omega_{\text{cross}} \propto MW^{-3.2}$, respectively. This is in good agreement with the experimental findings reported in the literature for hyaluronan solutions [36]. Despite the increase in ω_{cross} determined by the dilution of hyaluronan with alginate, it appears that the entangled structure was not altered, with respect to the dilution of hyaluronan alone, by the presence of alginate. However, in the case of the HA sample with the highest molecular weight, the mechanical response of the binary mixture was also compared with a solution containing only hyaluronan at the same concentration as in the mixture (Figure 5b, triangle). It can be noted that in this latter case, the ω_{cross} is higher than that of the mixture.

Figure 5c reports the dependence of the modulus-at-cross-over, $G_{\text{cross}} (G_{\text{cross}} \equiv G'_{\text{cross}} = G''_{\text{cross}})$, on the molecular weight of HA. It can be seen that in the case of hyaluronan alone, a power-law

dependence $G_{cross} \propto MW_{HA}^{-0.1}$ holds, which is in good accordance with the experimental results previously reported for hyaluronan, *i.e.* $G_{cross} \propto MW^0$ [36]. In contrast, when alginate is added to HA to form the binary system, G' and G'' show a non-zero (negative) dependence on the molecular weight of hyaluronan - more specifically, $G_{cross} \propto MW^{-1.2}$. It follows that the presence of the second polysaccharide brings about interference on the viscoelastic properties of the binary system whose extent is dependent on the molecular weight of the hyaluronan used in the system. Figure 6 shows the relative effect on η_0 and ω_{cross} due to the dilution of a hyaluronan solution with the solvent and of a HA/Alg binary mixture with $\phi_{HA} = 0.5$.

Figure 6

In relative terms, the variation of ω_{cross} was more marked in the case of the mixture than of the HA alone while basically no effect was noted on η_0 . This behaviour indicates that alginate lacks specific interactions with hyaluronan and contributes to the viscosity in simply additive terms. At variance, alginate markedly affects the formation of the complex network of entanglement which is the basis for the mechanical response of HA containing systems.

4. Conclusion

The biotechnological relevance of polysaccharide mixtures as ECM mimics has gained attention over recent years. However, while mixed systems have been exploited for *in-vitro* experiments, extended physical-chemical analyses are still partially lacking.

The present manuscript deals with the experimental physical-chemical characterisation by rotational rheometry of binary mixtures of alginate and hyaluronan. These analyses allow us to propose a model for the mutual arrangement of the two polysaccharides in solution that envisages a situation

275 where the polyanions tend to segregate instead of mixing freely, but without leading to macroscopic
276 phase separation. These results may impact systems in which a three-dimensional arrangement
277 (hydrogel), for biological purposes where a controlled release of the biologically active component
278 might affect the final outcome. Rheological measurements in semi-dilute solution provide new
279 information on the system at the macroscopic level and have shown that, although the combined
280 presence of the two polysaccharides does not give rise to marked synergistic effects, alginate and
281 hyaluronan tends to exclude each other, thus forming microdomains enriched in the two separate
282 components instead of a true homogeneous one-phase solution. This conclusion has notable
283 implications for systems using hydrogel formations. In fact, the treatment of the binary mixtures
284 with calcium ions, although leading to a three dimensional architecture due to the presence of
285 alginate, could result in an uneven distribution of the two components where hyaluronan might be
286 excluded from the three dimensional structure and may be confined at the borders of the hydrogel
287 [22]. The present manuscript will be followed by another work where the formation and mechanical
288 properties of hydrogels from mixed systems of alginate and hyaluronan, together with their
289 biological properties, will be explored.

290

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Captions to Figures

301

302 **Figure 1.** Dependence of the reduced specific viscosity from mixture composition for HA/Alg
303 systems at total polymer concentration of 1 g/L in aqueous 0.15M NaCl. The dotted line represents
304 the theoretical dependence based on Donati et al [26].

305

306 **Figure 2.** Dependence of the zero-shear viscosity (η_0 , **a**) and relaxation time (k , **b**) on HA
307 molecular weight (MW) for hyaluronan alone (■) and for HA-Alg binary mixtures with $\phi_{HA} = 0.5$
308 (□). In **a**, ▲ represents a hyaluronan solution at a total polymer concentration of 15 g/L and △
309 represents the theoretical value of zero-shear viscosity calculated for a HA-Alg binary mixtures
310 with $\phi_{HA} = 0.5$ according to [26]. Inset in **a**: Dependence of the scaling factor β ($\eta_0 \propto MW^\beta$) on the
311 fraction of HA in the HA/Alg binary mixture. In all cases, total polymer concentration = 30 g/L.

312

313 **Figure 3. a)** Dependence of the zero-shear viscosity, η_0 , on hyaluronan fraction (ϕ_{HA}) for binary
314 mixtures with alginate of HA150 (■), HA80 (□) and HA40 (●). **b)** Dependence of the zero shear
315 viscosity, η_0 , on hyaluronan concentration (C_{HA}) of a HA solution upon addition of alginate at
316 constant total polymer concentration of 30 g/L (■), or upon dilution (□). **c)** Dependence of the
317 polymer-chain relaxation time, k , on hyaluronan concentration (C_{HA}) of a HA solution upon
318 addition of alginate at constant total polymer concentration of 30 g/L (■), or upon dilution (□).

319

320 **Figure 4.** Example of a stress sweep (G') test for a mixture of hyaluronan and alginate (■). Dotted
321 line represents the best fit of the experimental data according to the Soskey-Winter equation (eq. 5).

322

323 **Figure 5. a)** Dependence of the storage (G' , ■) and loss (G'' , □) moduli from the pulsation, ω ($=$
324 $2\pi\nu$), for a HA/Alg binary solution, with hyaluronan weight fraction $\phi_{\text{HA}} = 0.5$ (sample HA150).
325 Solid and dotted lines represent generalised Maxwell model best fitting (eq. 5 and 6). **b)**
326 Dependence of the cross-over pulsation on molecular weight of hyaluronan for HA alone (■) and
327 HA/Alg binary solutions with $\phi_{\text{HA}} = 0.5$ (□). In both cases, total polysaccharide concentration was
328 30 g/L. Parenthesis indicate ω_{cross} values outside the frequency range explored and calculated with
329 eqs. 5 and 6. In **b)**, ▲ represents the pulsation corresponding to cross-over for a HA solution of 15
330 g/L. **c)** Dependence on the molecular weight of HA of the elastic (G') and viscous (G'') modulus at
331 the cross-over point for a hyaluronan solution alone (■) and for a HA/Alg binary solution with ϕ_{HA}
332 $= 0.5$ (□). All solutions were prepared in aqueous 0.15 M NaCl.

333

334 **Figure 6.** Relative variation of zero shear viscosity ($\eta_{0c}/\eta_{0c=30g/L}$) with total polymer concentration
335 ($C_{\text{Tot}} = C_{\text{HA}} + C_{\text{Alg}}$) (right y-scale) for HA (■) and HA/Alg mixture (□, $\phi_{\text{HA}} = 0.5$) upon dilution
336 with solvent (0.15 M NaCl). Relative dependence of the pulsation corresponding to the crossover ($\omega_{\text{cross},c}/\omega_{\text{cross},c=30g/L}$) on total polymer concentration (left y-scale) for HA (●) and for the HA/Alg
337 mixture (○, $\phi_{\text{HA}} = 0.5$).

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Table 1. Composition, critical strain and critical stress (τ_c) for binary HA/Alg mixtures in NaCl 0.15 M. ϕ_{HA} represents the weight fraction of hyaluronan in the mixture.

Sample	ϕ_{HA}	γ_c	τ_c (Pa)
HA150 30 g/L	1	0.44	123.3
HA150 22g/L	1	0.50	72.6
HA150 18g/L	1	0.51	45.5
HA150 15g/L	1	0.99	48.3
HA80 30 g/L	1	0.55	107.3
HA40 30 g/L	1	0.79	38.2
HA150/Alg 30 g/L	0.73	0.7	79.5
HA150/Alg 30 g/L	0.6	0.68	77.1
HA150/Alg 30 g/L	0.5	0.58	46.5
HA150/Alg 30 g/L	0.4	0.62	30.3
HA80/Alg 30 g/L	0.73	0.54	61.4
HA80/Alg 30 g/L	0.6	0.64	33.9
HA80/Alg 30 g/L	0.5	0.91	47.0
HA80/Alg 30 g/L	0.4	0.68	21.1
HA40/Alg 30 g/L	0.73	1.0	26.8
HA40/Alg 30 g/L	0.6	0.86	13.2
HA40/Alg 30 g/L	0.5	0.85	16.8
HA150/Alg 25 g/L	0.5	0.59	29.2
HA150/Alg 20 g/L	0.5	0.65	15.0
HA150/Alg 15 g/L	0.5	0.65	6.8

Table 2. Parameters obtained from the fitting of the experimental datapoints of the mechanical spectra for the hyaluronan/alginate binary mixture (G_i , λ_i , η_i , eqs. 5 and 6).

Sample	ϕ_{HA}	G_i (Pa)	λ_i (s)	η_i (Pa·s)
HA150 30g/L	1	281.2	0.0041	1.2
		288.9	0.0414	11.9
		189.9	0.414	78.6
		62.1	4.14	257.1
HA150 22 g/L	1	202.0	0.0074	1.49
		165.2	0.074	12.2
		77.0	0.739	56.9
		12.7	7.388	93.7
HA150 18 g/L	1	153.2	0.005	0.8
		126.1	0.0504	6.4
		53.1	0.5041	26.8
		9.2	5.0414	46.2
HA150 15 g/L	1	91.8	0.0011	0.1
		94.2	0.0109	1.0
		64.1	0.1093	7.0
		17.3	1.0935	18.9
HA80 30 g/L	1	432.2	0.0045	1.9
		313.3	0.0451	14.1
		110.8	0.4513	49.9
		14.3	4.5513	64.7
HA40 30g/L	1	589.6	0.0006	0.4
		388.4	0.0061	2.4
		85.6	0.0615	5.3
		4.8	0.6147	3.0
HA24 30g/L	1	351.9	0.0001	0.04
		290.8	0.0012	0.3
		54.1	0.012	0.6
		2.8	0.12	0.3

Table A.2 (continued)

Sample	ϕ_{HA}	G_i (Pa)	λ_i (s)	η_i (Pa·s)
HA150/Alg 30 g/L	0.73	281.1	0.004	1.1
		179.9	0.040	7.2
		112.6	0.399	44.9
		32.1	3.986	128.0
HA150/Alg 30 g/L	0.6	325.4	0.0015	0.5
		168.1	0.0153	2.6
		112.0	0.1533	17.2
		43.3	1.5333	66.3
HA150/Alg 30 g/L	0.5	256.4	0.004	1.1
		107.2	0.041	4.4
		53.4	0.412	22.0
		11.2	4.119	46.2
HA150/Alg 30 g/L	0.4	385.5	0.0013	0.5
		96.3	0.0129	1.2
		51.7	0.1292	6.7
		15.4	1.2924	19.8
HA80/Alg 30 g/L	0.73	506.1	0.001	0.5
		295.9	0.009	2.9
		157.3	0.098	15.4
		29.6	0.976	28.9
HA80/Alg 30 g/L	0.6	325.3	0.003	1.0
		124.0	0.031	3.9
		29.6	0.313	9.3
		1.9	3.129	6.0
HA80/Alg 30 g/L	0.5	329.6	0.003	1.1
		120.4	0.032	3.9
		30.5	0.322	9.8
		2.3	3.218	7.4
HA80/Alg 30 g/L	0.4	393.2	0.0006	0.3
		172.8	0.006	1.1
		51.7	0.064	3.3
		6.9	0.644	4.5

Table A.2 (continued)

Sample	ϕ_{HA}	G_i (Pa)	λ_i (s)	η_i (Pa·s)
HA40/Alg 30 g/L	0.73	478.1	0.002	1.0
		111.3	0.021	2.3
		8.8	0.206	1.8
		0.2	2.064	0.4
HA40/Alg 30 g/L	0.6	386.9	0.002	0.9
		54.5	0.023	1.3
		3.2	0.236	0.7
		0.01	2.357	0.03
HA40/Alg 30 g/L	0.5	481.7	0.002	1.2
		66.3	0.024	1.6
		4.1	0.244	1.0
		0.07	2.436	0.2
HA150/Alg 25 g/L	0.5	287.0	0.001	0.4
		95.0	0.012	1.2
		55.7	0.122	6.8
		16.4	1.222	20.1
HA150/Alg 20 g/L	0.5	280.9	0.0008	0.2
		46.3	0.008	0.4
		33.3	0.082	2.7
		8.2	0.825	6.7

Figure 1

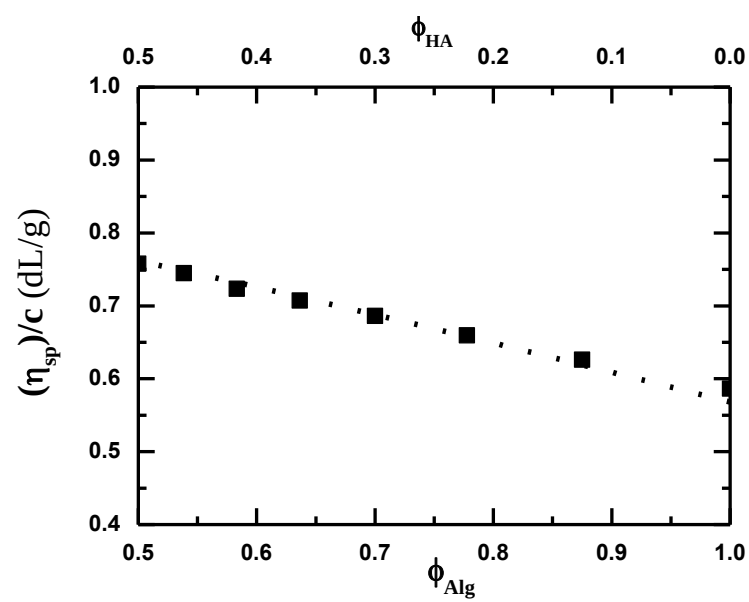


Figure 2

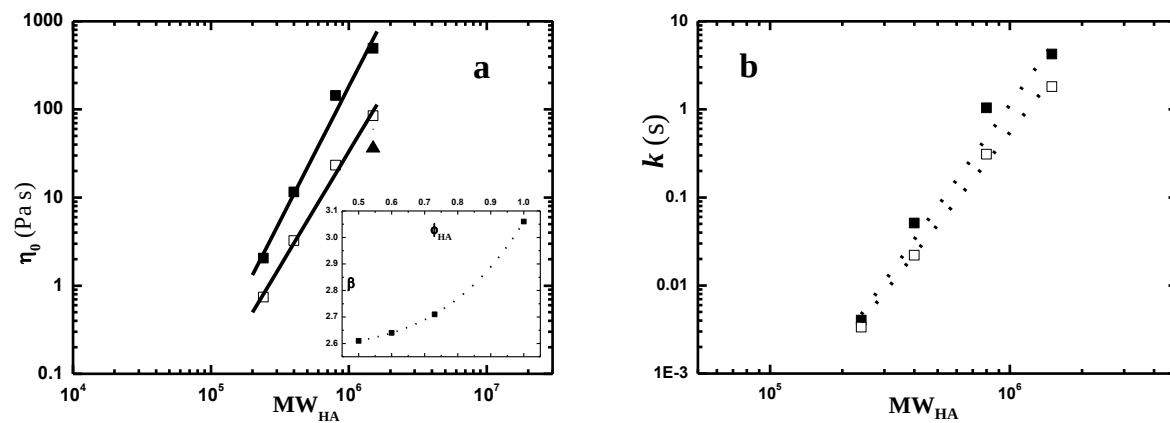


Figure 3

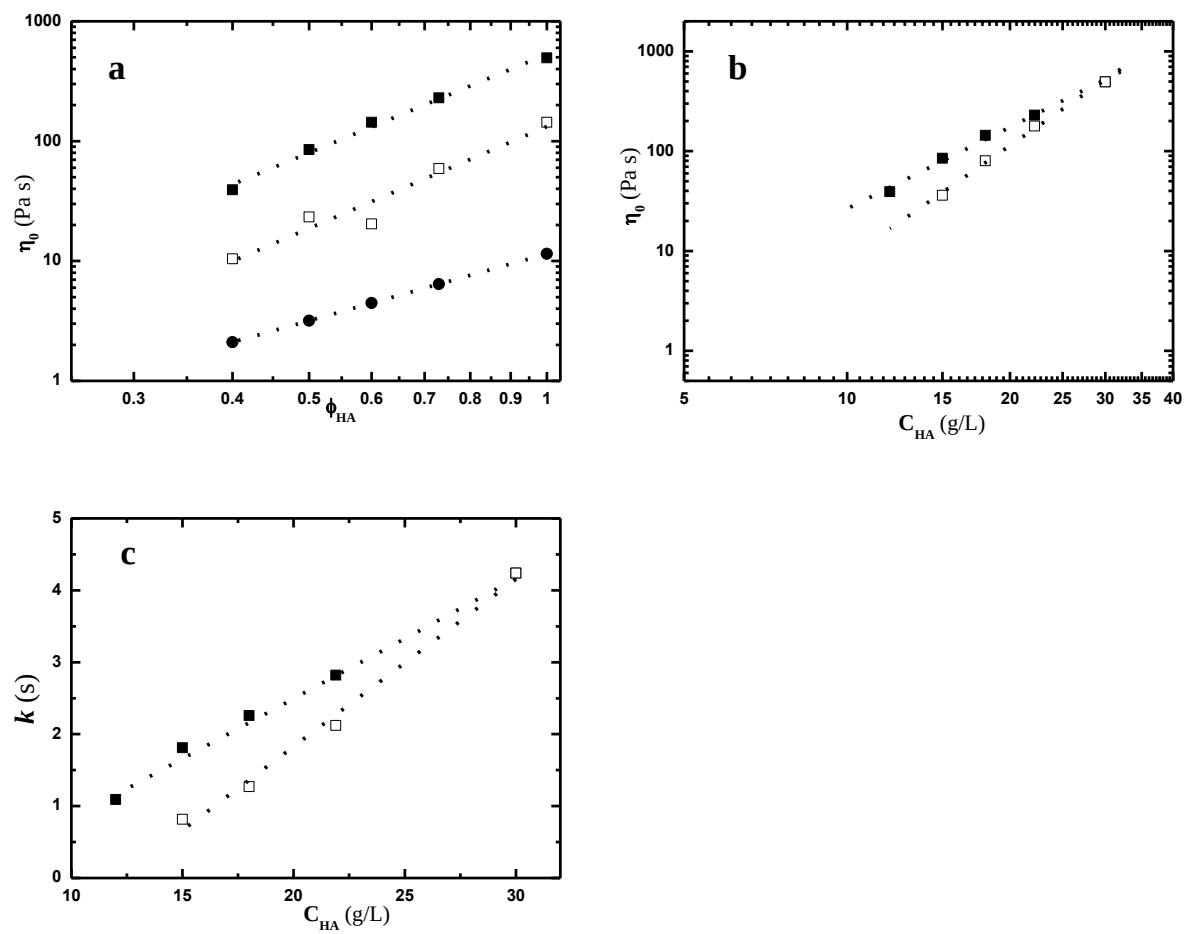


Figure 4

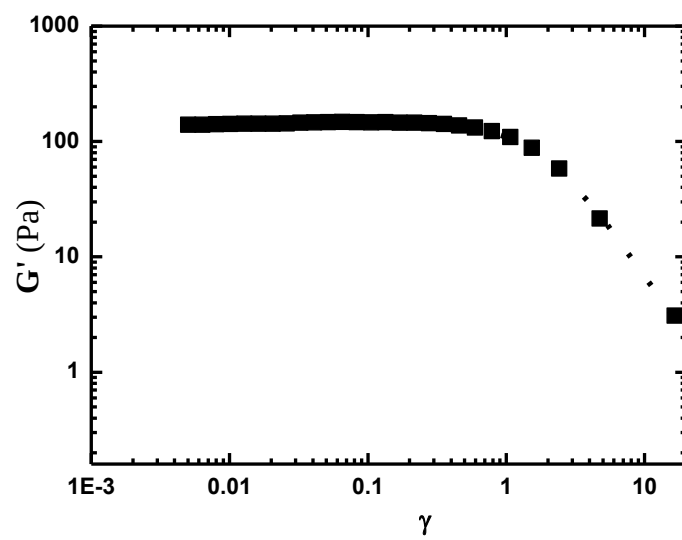


Figure 5

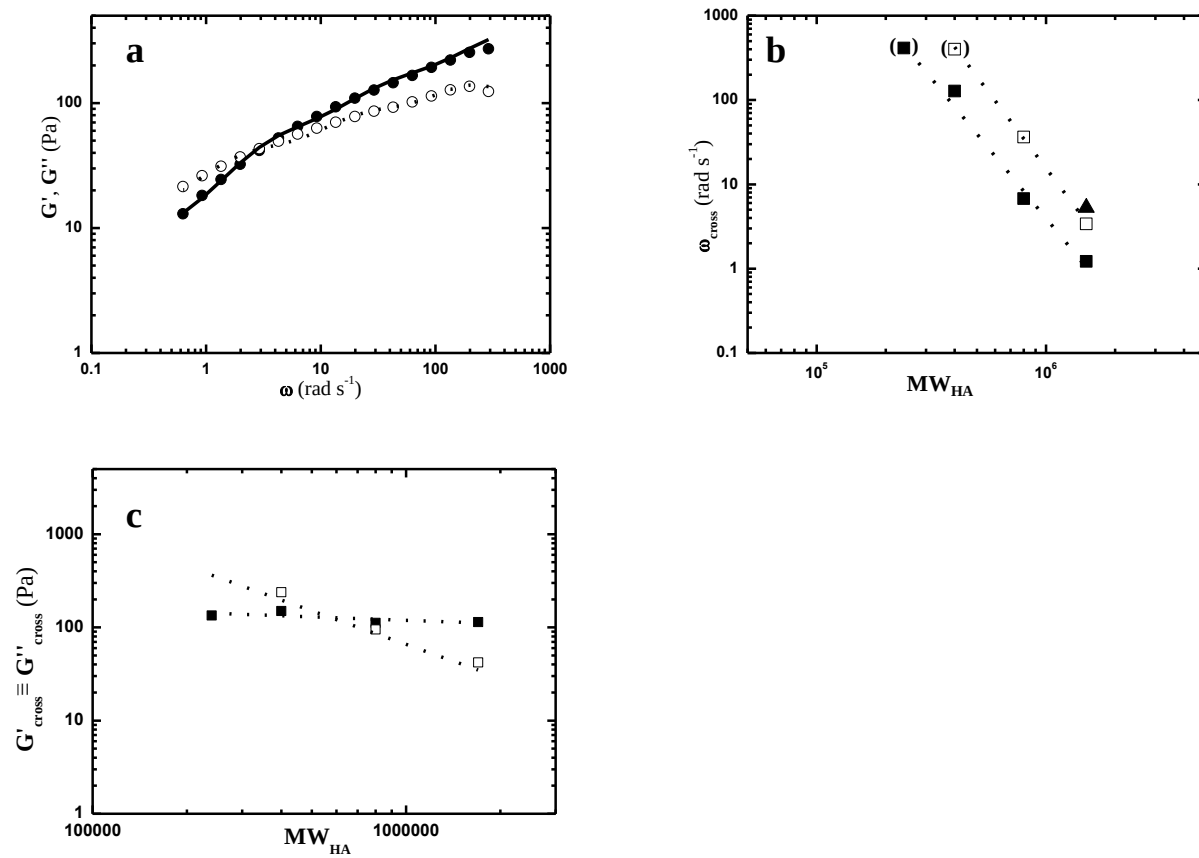


Figure 6

