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Title: Crossreactivity between MHC class I-restricted antigens from cancer cells and an enterococcal bacteriophage.

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One sentence summary:

Cytotoxic T lymphocytes that recognize antigens from a prophage of a commensal enterococcus can mediate anticancer immunosurveillance by recognizing cross-reactive tumor-associated
95 antigens.

Abstract:

It has been speculated that the intestinal microbiota induces commensal-specific memory T cells
100 that then cross-react with tumor-associated antigens. Here, we identified MHC class I-binding
epitopes within the tail length tape measure protein (TMP) of a prophage found in the genome of
Enterococcus hirae. Mice bearing *E. hirae* strains harboring this prophage mounted a TMP-
specific H-2K^b restricted CD8⁺ T lymphocyte response upon immunotherapy with
cyclophosphamide or anti-PD1 antibodies. Such TMP-specific T cells also recognized a 78%-
105 identical H-2K^b-binding peptide derived from the proteasome (20S) subunit beta type-4
(PSMB4), allowing them to control mouse tumors expressing this oncogenic driver.
Administration of bacterial strains engineered to express the TMP epitope improved the outcome
of immunotherapy. Tumors bearing PSMB4 knock-in mutations that abolish crossreactivity with
TMP became immunotherapy-resistant. In renal and lung cancer patients, the presence of the
110 enterococcal prophage in stools, as well as the expression of a TMP-cross reactive antigen by
tumors, predicted the long-term benefit of PD-1 blockade. In melanoma patients, we detected T
cell clones recognizing naturally processed cancer antigens that are cross-reactive with microbial
peptides. Altogether, these results support the idea that intestinal microbe-specific T cell
responses contribute to anticancer immunosurveillance.

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Main Text:

120 Unleashing immune responses against tumor-associated antigens through chemotherapy,
radiotherapy, targeted therapies or immune checkpoint inhibitors has become the mainstay of
successful cancer treatments (1, 2). The recent discovery that the gut microbiota determines the
cancer-immune set point, thus influencing the clinical outcome of antineoplastic therapies, has
rekindled the concept that microbes or their products modulate not only intestinal but also
125 systemic immunity (3, 4). Indeed, memory responses by interferon- γ (IFN γ) secreting CD4⁺ and
CD8⁺ T cells specific for *Enterococcus hirae*, *Bacteroides fragilis*, and *Akkermansia muciniphila*
are associated with favorable clinical outcome in cancer patients (5–8), suggesting that microbe-
specific T lymphocytes may contribute to antitumor immune responses. The mechanisms through
which microbes trigger chronic intestinal inflammation and systemic autoimmune disease have
130 not been resolved (9). The theory of molecular mimicry (10–14) posits that T cells elicited by
bacteria or viruses accidentally recognize autoantigens as they ‘escape’ from self-tolerance
inducing mechanisms (such as clonal deletion or inactivation). While MHC class I and class II
binding epitopes encoded by bacterial genomes may be immunogenic (10–14), very few reports
have demonstrated that microbe-specific CD4⁺ or CD8⁺ T lymphocytes attack normal or
135 neoplastic tissues (15–17).

Cyclophosphamide (CTX) induces the translocation of *E. hirae* from the gut lumen to the
mesenteric and splenic immune tissues, thereby eliciting specific CD4⁺ and CD8⁺ T lymphocytes
140 producing interleukin-17 (IL17) and interferon- γ (IFN γ), correlating with therapeutically
effective anticancer immune responses (6, 18). Broad-spectrum antibiotics abolished the
therapeutic efficacy of CTX unless *E. hirae* was supplied by oral gavage (6). When comparing a
panel of distinct *E. hirae* strains (Table S1, Figure S1A) for their capacity to restore the
antibiotic-perturbed anticancer effects of CTX, we found that only a few *E. hirae* isolates (such
145 as 13144 and IGR11) were efficient (Figure 1A-B, Ref. (6)). Given that the therapeutic efficacy
of the combination of CTX and *E. hirae* 13144 is abrogated by the depletion of CD8⁺ T cells or
the neutralization of IFN γ (6), we screened the differential capacity of *E. hirae* strains to elicit
memory T cell responses after priming of the host, measured as the *ex vivo* recall response (IFN γ
secretion) of splenic CD8⁺ T cells against various *E. hirae* strains loaded onto dendritic cells

150 (DC) (Figure S2A). While *E. hirae* 13144 triggered specific CD8⁺ T cell responses (that were not cross-reactive against irrelevant enterococci), *E. hirae* 708 and 13344 (two prototypic inefficient strains) (6) failed to do so (Figure S2A).

To identify relevant T cell epitopes, we aligned the sequences of bacterial genes encoding putative cell wall and secreted proteins for immunogenic (13144) versus non-immunogenic (708 and 13344) *E. hirae* strains, followed by the *in silico* identification of 13144-specific nonapeptides with strong affinity (<50 nM) for the MHC class I H-2K^b protein (Table S2). Subsequently, we recovered splenic CD8⁺ T cells from mice that had been exposed to *E. hirae* 13144 and CTX (Figure 1C), restimulated them *in vitro* with pools of potentially immunogenic nonapeptides from *E. hirae* 13144 to measure IFN γ production (Table S2, Figure S2B) and finally split the most efficient pool (No. 7) into individual peptides (Figure 1D). This approach led to the identification of one dominant epitope (one-letter amino acid [aa] code: TSLARFANI, abbreviation TMP1) in position 187 to 197 of the aa sequence of the phage tail length tape measure protein (TMP, 1506 aa) from a 39.2 kb prophage of *E. hirae* 13144 (Figure 1D, Figure S3, Tables S2-S3). The 39.2kb prophage encodes 65 genes, including one shared between all 18 *E. hirae* genomes and 38 unique to *E. hirae* 13144 (Figure S1B), encoding capsid, portal and tail structures characteristic of *Siphoviridae* phages. Importantly, the TMP1 epitope of the 39.2kb prophage from *E. hirae* 13144 and the prophage fragment contained in *E. hirae* IGR11 showed 100% sequence identity (Figure S3 and S4A). Accordingly, *E. hirae* IGR11 was as efficient as *E. hirae* 13144 in reducing the growth of MCA205 sarcomas treated with CTX (Figure 1A-B). In contrast, the absence of a *bona fide* TMP1 epitope (observed in *E. hirae* 708 and 13344, Figure S1B) and a mutation in position 3 of the TSLARFANI peptide (L \rightarrow F observed in *E. hirae* ATCC9790, Figure S4A) correlated with the lack of anticancer effects of these *E. hirae* strains (Figure 1B and Ref. (6)). ELISpot assays designed to detect peptide-specific IFN γ -producing T cells revealed that mice gavaged with *E. hirae* 13144 or IGR11 mounted a CD8⁺ T cell response against TMP1 (but not against the control peptides TMP2 and TMP3), while mice receiving *E. hirae* strains lacking TMP1 (strains 708, 13344) or a strain possessing a mutated TMP1 (strain ATCC9790) were unable to do so (Figure 1D). We used a fluorescent H-2K^b/TSLARFANI tetrameric complex (and its negative control H-2K^b/SIINFEKL binding to ovalbumine (OVA) specific CD8⁺ T cells) to detect the frequency and distribution of TMP1-specific cytotoxic T

lymphocytes (CTLs) in naive and MCA205 sarcoma bearing C57BL/6 mice. We observed a specific increase in splenic CD8⁺ T cells that recognized the TMP1 peptide (but not the OVA peptide SIINFEKL) at day 7 following treatment with CTX and gavage with *E. hirae* 13144 (Figure 1E), as well as in tumor draining lymph nodes (LN) of tumor bearers at day 14 after
185 treatment with CTX and gavage with *E. hirae* 13144 (Figure S2C-D). Splenic TMP1 (but not OVA)-specific (H-2K^b/TSLARFANI tetramer-positive) CTLs also increased in their frequency after gavage with *E. hirae* IGR11 (but not 13344 nor ATCC9790) (Figure 1E). The H-2K^b/TSLARFANI tetramer-positive CTLs were specifically enriched in the CXCR3⁺CCR9⁺ fraction of CD8⁺ T cells from secondary lymphoid organs (Figure S2C). Even in mice colonized
190 with human fecal materials, CTX administration and oral gavage with *E. hirae* 13144 induced an anticancer effect (Figure S2E) and an expansion of H-2K^b/TSLARFANI tetramer-positive CTL in tumor draining LN at day 7 and in tumor beds at day 17 while vanishing from mesenteric LN (Figure S2F-H). Hence, immunogenic *E. hirae* elicits a H-2K^b restricted CTL response against the TMP-derived peptide TMP1/TSLARFANI.

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To explore the capacity of TMP1-specific H-2K^b restricted T cells to control the growth of MCA205 cancers, we subcutaneously (s.c.) immunized naive C57BL/6 mice with dendritic cells (DC) loaded with heat-inactivated *E. hirae* 13144 (positive control), the naturally occurring TMP1/TSLARFANI peptide from 13144 and IGR11, its L→F mutant from *E. hirae* ATCC9790
200 ('mut3', Figure 2A, Figure S4A) or other non-immunogenic bacterial peptides (group 1, Figure S2B). In this prophylactic setting, DC pulsed with TMP1 (but not mut3) were as efficient as the whole *E. hirae* extract in reducing tumor growth (Figure 2B-C). Next, we explored whether the TMP1 peptide would be able to confer immunogenicity to the usually inefficient bacterium *Escherichia coli* strain DH5α in the therapeutic setting, in which antibiotic treatment is followed
205 by gavage with different bacterial strains and CTX-based chemotherapy (Figure 1A and Ref. (6)). *E. coli* engineered to express TMP1 (Figure S5) was as efficient as *E. hirae* 13144 in restraining MCA205 tumor growth (Figure S4B, Figure 2D) and eliciting tetramer binding CTL in the spleen (Figure 2E). In contrast, *E. coli* expressing an irrelevant sequence (encoding mouse EGFP protein), mut3 or mutant TMP1 bearing a S→A exchange in the anchor position 2 ('mut2')
210 (Figure 2A) failed to induce such a cancer-protective immune response (Figure 2D-E).

To explore the mechanism by which TMP1 exerts its anticancer activity against MCA205 tumors in C57BL/6 mice, we investigated whether H-2K^b-restricted mouse tumor antigens with high identity to the TMP1 peptide (TSLARFANI) exist. Using the NCBI BLASTP suite, we found that the peptide (GSLARFRNI) belonging to the proteasome subunit beta type-4 (PSMB4) located at amino acid positions 76-84 shared a strong homology (7 out of 9 amino acids with identical amino acids at the MHC Class I anchoring positions 2 and 9) with TMP1 (Figure 3A). We queried for potential neoepitopes of MCA205 but found no significant homology with TMP1, prompting us to focus on the non-mutated PSMB4 peptide. In fact, some mouse tumors (such as MCA205 sarcomas and TC1 lung cancers) overexpress the PSMB4 antigen compared with their normal tissues of origin, while others (such as MC38 colon cancers) failed to do so (Figure 3B). This correlates with the fact that MCA205 and TC1 tumors respond to the treatment with CTX+*E. hirae* 13144, while MC38 cancer does not (Figure S6A-B). PSMB4 is an oncogenic driver involved in proliferation and invasion (19) in a variety of malignancies such as glioblastoma (20), melanoma (21) and breast cancers (22), associated with dismal prognosis (19, 20, 22). CRISPR/Cas9-mediated genomic knock-in of the PSMB4 sequence replacing GSLARFRNI by GALARFRNI (with an S→A exchange in position 2) or GSEARFRNI (with an L→F exchange in position 3 equivalent to mut 3 of TSLARFANI) in MCA205 cells (Figure S7) significantly affected tumor growth kinetics (Figure S6C-D), suggesting that this PSMB4 epitope contributes to the oncogenic activity of PSMB4. While these knock-in mutations did not interfere with the efficacy of CTX treatment alone, they drastically blunted the anticancer effects of *E. hirae* 13144 (Figure 3C-D). We extended these findings to a second tumor model where the anticancer effects of the combination of CTX+*E. hirae* 13144 were additive even in the absence of antibiotic-induced dysbiosis. Introducing a knock-in mutation in position 3 of PSMB4 into TC1 lung cancer cells again compromised the antitumor effects of CTX (Figure 3E). Moreover, in the setting of PD-1 blockade, administration of *E. hirae* 13144 without prior conditioning with antibiotics reduced the growth of parental but not PSMB4-mutated MCA205 cancers (Figure S6E). These results support the idea that the TSLARFANI TMP1 peptide encoded by *E. hirae* 13144 indeed induces T cell responses against the PSMB4-derived GSLARFRNI peptide across different tumor types and therapy modalities.

Reinforcing the notion of molecular mimicry between phage-encoded and cancer antigens, flow cytometric analyses using fluorescent-labelled tetramers H-2K^b/TSLARFANI (from TMP1) and H-2K^b/GSLARFRNI (from PSMB4) identified a subset of double-positive CTLs that infiltrate MCA205 tumors from CTX/*E. hirae* 13144-treated mice (Figure S6F) and that was as frequent as CTLs recognizing the PSMB4 peptide only (Figure 4A). We purified the splenic CD8⁺ T cells using either the TMP1-H-2K^b or PSMB4-H-2K^b specific tetramers and stimulated them with irrelevant (OVA-derived-SIINFEKL) *versus* relevant (TMP-derived TSLARFANI or PSMB4-derived GSLARFRNI) peptides (Figure 4B). CD8⁺ T cells binding H-2K^b-TMP1 tetramers produced IFN γ not only in response to TMP1 (up to 5-fold increase in IFN γ secreting T cells) but also in response to the PSMB4 epitope (2-fold increase, as much as with heat-killed *E. hirae* 13144 processed by DC) (Figure 4C, Figure S6G). Similarly, CD8⁺ T cells binding H-2K^b-PSMB4 tetramers functionally recognized TMP1, albeit less efficiently than the PSMB4 epitope (Figure S6G). We analyzed the T cell receptor (TCR) repertoire of these two tetramer-reactive CD8⁺ T cell subsets. In accordance with the functional data, half of the CD8⁺ T cells labelled with PSMB4-H-2K^b tetramers shared clonotypes with the much wider TCR repertoire of T cells labelled with the TMP1-H-2K^b specific tetramers (Figure 4D, Table S4-S5) (but not with the negative fraction, Figure S6H). In sum, T cells recognizing the TMP1 epitope of immunogenic *E. hirae* can crossreact with a peptide contained in the oncogenic driver PSMB4 and *vice versa*.

Temperate bacteriophages are bacterial viruses that can transfer virulence, antimicrobial resistance genes, and immunogenic sequences to new bacterial hosts (23). The TMP protein, which contains a variable number of tandem repeats with highly conserved tryptophan and phenylalanine residues at fixed positions is encoded by the genome of *Siphoviridae* phages (24, 25). To investigate the capacity of the *E. hirae* 13144 phage to lysogenize other bacterial species *in vivo*, we performed culturomic analyses of the ileal content from C57BL/6 mice subjected to oral gavage with *E. hirae* 13144 and systemic CTX therapy, followed by PCR analyses seeking TMP sequences (Figure S8A-B). We tested 7 to 18 bacterial colonies from each animal and a total of 76 colonies. We only found lysogenic conversion of *E. gallinarum* by the *E. hirae*-temperate phage *in vivo*, as confirmed by sequencing of the phage genome in the second host (Figure 4E, Figure S8B-C). In contrast, none of the 90 colonies (mostly of *E. gallinarum*) isolated from naive mice harbored the TMP sequence (Figure S8A). Similarly, *in vitro* coculture

of TMP⁺ *E. hirae* 13144 together with TMP⁻ *E. gallinarum* spp. at a 1:1 ratio uncovered a significant (~15%) rate of lysogenic conversion (Figure S8D). Examination of a preparation admixing *E. hirae* 13144 and *E. gallinarum* at a 1:10 ratio by means of transmission electron microscopy revealed numerous phages with the typical *Siphoviridae* morphology in the medium, whereas control cultures (bacteria separately) were free of such phages (Figure 4F). Altogether, these results indicate that the TMP1 peptide-encoding *Siphoviridae* phage from *E. hirae* 13144 is a virulent phage.

We next explored the possible pathophysiological relevance of these findings. We first screened a total of 3,027 adult and mother-infant metagenomes (26), validated by a second independent metagenomic-assembly based screening of 9,428 metagenomes (27) (28), to assess the breadth of coverage (BOC) of the *E. hirae* genome and its phages (Figure S9A). *E. hirae* was present with 100% confidence (i.e. BOC > 80%) in less than 150 fecal samples from disparate geography, age and datasets. This phage (and its host) could be vertically transmitted from mothers to infants and then colonizes the neonate. There was an increased prevalence of the phage (57%) in fecal microbiomes from children (representing 16% of all metagenomes, Fisher's test p-value <0.00001). Of note, the *E. hirae* 13144 phage was detectable in many samples lacking the presence of the *E. hirae* core genome, suggesting that other bacteria than *E. hirae* can host this phage. All host genomes belonged to the *Enterococcus* genus (except two assigned to *Coprobacillus*), in particular *E. faecalis* (80 genomes), *E. faecium* (23), and *E. hirae* (15), suggesting that phage 13144 (and its homologues from *E. hirae* 708, and 13344) are genus-specific but not species-specific.

Contrasting with metagenomics that has a low sensitivity to detect poor abundance species, culturomics followed by matrix-assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF) provides a technology for detecting rare *E. hirae* colonies in the stool of healthy individuals (29) or cancer patients (8). PCR analyses of each single cultivatable enterococcal colony (up to 5 per species and individual) from 76 cancer patients led to the detection of the TMP sequence encompassing the TMP1 peptide in 34% of the patients, only in *E. faecalis* and *E. hirae* (Figure S9B, Figure S10). Advanced renal and lung cancer patients (cohort described in Ref. (16)) with detectable fecal TMP at diagnosis exhibited prolonged

overall survival after therapy with immune checkpoint inhibitors targeting PD-1 (Figure 4G).
305 Therefore, we screened sixteen TMP-derived nonapeptides predicted to bind the human MHC
class I HLA-A*0201 with high affinity for their ability to prime naive CD8⁺ T cells from six
healthy volunteers *in vitro*. We found 6 out of 16 epitopes capable of triggering significant
peptide-specific IFN γ release that were located in two distinct regions of the TMP protein (504-
708 and 1397-1462, Figure S11A-B, Table S6). Using the NCBI BLASTP suite, we searched the
310 human cancer peptidome (of the TCGA database) for a high degree of homology with these 6
HLA-A*0201 -restricted immunogenic nonapeptides. We found that only the TMP-derived
peptide KLAKFASVV (aa 631-639) shared significant homology (7 out of 9 aa, with identical
residues at the MHC anchoring positions 2 and 9) with a peptide contained in the protein
glycerol-3-phosphate dehydrogenase 1-like (GPD1-L) (Figure S11C). GPD1-L reportedly
315 counteracts the oncogenic HIF1 α -dependent adaptation to hypoxia, and its expression is
associated with favorable prognosis in head and neck squamous cell carcinomas (30–32). The
TCGA transcriptomics database unveiled that high expression of GPD1-L is associated with
improved overall survival in lung adenocarcinoma and kidney cancers (Figure S11D). Moreover,
high expression of GPD1-L mRNA by tumors at diagnosis was associated with improved
320 progression-free survival in three independent cohorts of non-small cell lung cancer (NSCLC)
patients (n=157, Table S7) treated with anti-PD1 Abs (Figure S11E-F). Expression of GPD-1L
failed to correlate with that of PD-L1 in NSCLC (Figure S11G). Of note, mutations in or adjacent
to the 631-639 amino acid sequence of GPD-1L gene could rarely be identified in several types
of neoplasia (Figure S12).

325 We derived an HLA-A*0201-restricted, phage peptide (KLAKFASVV)-specific T cell line from
peripheral blood mononuclear cells of a human volunteer. Clones from this line also recognized
the HLA-A*0201 -restricted, GPD-1L epitope (KLQKFASTV) (Figure S13A-C). Moreover, we
detected CD8⁺ T cells binding HLA-A*0201/KLAKFASVV tetramers exhibiting hallmarks of
330 effector functions after *in vitro* stimulation of PBMC with the KLAKFASVV phage epitope in 3
out of 6 NSCLC patients (Fig. S13D-F). In the reverse attempt searching for molecular mimicry
between well known and naturally processed non-mutated melanoma differentiation antigens
recognized by human T cell clones (such as HLA-A*0201-binding MART-1 or MELOE
epitopes) and gut commensal antigens, we found microbial analogs in the public microbiome data

335 bases (Figure S14, Table S8-Table S9, Figure S15, Table S8-S10). Some of these microbial peptides are recognized by the corresponding TCR (Tables S9- S10) with similar affinities as the parental (tumoral) epitope.

Altogether the present results demonstrate that microbial genomes code for MHC class I-
340 restricted antigens that induce a memory CD8⁺ T cell response, which then crossreacts with cancer antigens. Several lines of evidence plead in favor of this interpretation, as exemplified for the TMP1 epitope found within a phage that infects enterococci. First, naturally occurring ('mut3' in *E. hirae* strain ATCC9790) or artificial mutations ('mut2' or 'mut3' in *E.coli*) introduced into the TMP1 epitope suppressed the tumor-prophylactic and therapeutic potential of
345 bacteria expressing TMP1. Second, transfer of the TMP1-encoding gene into *E. coli* conferred immunogenic capacity to this proteobacterium, which acquired the same antitumor properties as TMP1-expressing *E. hirae*. Third, when cancer cells were genetically modified to remove the TMP1-crossreactive peptide within the PSMB4 protein, they formed tumors that could no longer be controlled upon oral gavage with TMP1-expressing *E. hirae*. Fourth, cancer patients carrying
350 the TMP phage sequence in fecal enterococci spp. or the GPD1-L tumoral antigen homologous to TMP epitopes exhibited a better response to PD-1 blockade, suggesting that this type of microbe-cancer cross-reactivity might be clinically relevant.

Recent reports point to the pathological relevance of autoantigen-crossreactive, microbiota-
355 derived peptides for autoimmune disorders such as myocarditis, lupus and rheumatoid arthritis (34–36). Given the enormous richness of the commensal proteome (37), we expect the existence of other microbial antigens mimicking auto- and tumor antigens. In fact, we extended these findings to naturally processed melanoma-specific antigens that have microbial orthologs recognized by the same TCRs. Global phage numbers have been estimated to reach as high as
360 10³¹ particles with the potential of 10²⁵ phage infections occurring every second (38, 39). Thus, the perspective opens that, within the microbiota, bacteriophages may enrich the therapeutic armamentarium for modulating the intestinal flora and for stimulating systemic anticancer immune responses.

365 **References and Notes:**

1. P. Sharma, J. P. Allison, Immune checkpoint targeting in cancer therapy: toward combination strategies with curative potential. *Cell*. **161**, 205–214 (2015).
2. L. Galluzzi, A. Buqué, O. Kepp, L. Zitvogel, G. Kroemer, Immunological Effects of
370 Conventional Chemotherapy and Targeted Anticancer Agents. *Cancer Cell*. **28**, 690–714 (2015).
3. L. Zitvogel, Y. Ma, D. Raoult, G. Kroemer, T. F. Gajewski, The microbiome in cancer immunotherapy: Diagnostic tools and therapeutic strategies. *Science*. **359**, 1366–1370 (2018).
- 375 4. T. Tanoue, S. Morita, D. R. Plichta, A. N. Skelly, W. Suda, Y. Sugiura, S. Narushima, H. Vlamakis, I. Motoo, K. Sugita, A. Shiota, K. Takeshita, K. Yasuma-Mitobe, D. Riethmacher, T. Kaisho, J. M. Norman, D. Mucida, M. Suematsu, T. Yaguchi, V. Bucci, T. Inoue, Y. Kawakami, B. Olle, B. Roberts, M. Hattori, R. J. Xavier, K. Atarashi, K. Honda, A defined commensal consortium elicits CD8 T cells and anti-cancer immunity. *Nature*.
380 **565**, 600–605 (2019).
5. M. Vétizou, J. M. Pitt, R. Daillère, P. Lepage, N. Waldschmitt, C. Flament, S. Rusakiewicz, B. Routy, M. P. Roberti, C. P. M. Duong, V. Poirier-Colame, A. Roux, S. Becharef, S. Formenti, E. Golden, S. Cording, G. Eberl, A. Schlitzer, F. Ginhoux, S. Mani, T. Yamazaki, N. Jacquelot, D. P. Enot, M. Bérard, J. Nigou, P. Opolon, A. Eggermont, P.-L. Woerther, E.
385 Chachaty, N. Chaput, C. Robert, C. Mateus, G. Kroemer, D. Raoult, I. G. Boneca, F. Carbonnel, M. Chamillard, L. Zitvogel, Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. *Science*. **350**, 1079–1084 (2015).
6. R. Daillère, M. Vétizou, N. Waldschmitt, T. Yamazaki, C. Isnard, V. Poirier-Colame, C. P. M. Duong, C. Flament, P. Lepage, M. P. Roberti, B. Routy, N. Jacquelot, L. Apetoh, S. Becharef, S. Rusakiewicz, P. Langella, H. Sokol, G. Kroemer, D. Enot, A. Roux, A. Eggermont, E. Tartour, L. Johannes, P.-L. Woerther, E. Chachaty, J.-C. Soria, E. Golden, S. Formenti, M. Plebanski, M. Madondo, P. Rosenstiel, D. Raoult, V. Cattoir, I. G. Boneca, M. Chamillard, L. Zitvogel, Enterococcus hirae and Barnesiella intestinihominis Facilitate
390 Cyclophosphamide-Induced Therapeutic Immunomodulatory Effects. *Immunity*. **45**, 931–943 (2016).
7. Y. Rong, Z. Dong, Z. Hong, Y. Jin, W. Zhang, B. Zhang, W. Mao, H. Kong, C. Wang, B. Yang, X. Gao, Z. Song, S. E. Green, H. K. Song, H. Wang, Y. Lu, Reactivity toward Bifidobacterium longum and Enterococcus hirae demonstrate robust CD8+ T cell response and better prognosis in HBV-related hepatocellular carcinoma. *Exp. Cell Res*. **358**, 352–359
400 (2017).
8. B. Routy, E. Le Chatelier, L. Derosa, C. P. M. Duong, M. T. Alou, R. Daillère, A. Fluckiger, M. Messaoudene, C. Rauber, M. P. Roberti, M. Fidelle, C. Flament, V. Poirier-Colame, P. Opolon, C. Klein, K. Iribarren, L. Mondragón, N. Jacquelot, B. Qu, G. Ferrere, C. Clémenson, L. Mezquita, J. R. Masip, C. Naltet, S. Brosseau, C. Kaderbhai, C. Richard,

- 405 H. Rizvi, F. Levenez, N. Galleron, B. Quinquis, N. Pons, B. Ryffel, V. Minard-Colin, P. Gonin, J.-C. Soria, E. Deutsch, Y. Loriot, F. Ghiringhelli, G. Zalcman, F. Goldwasser, B. Escudier, M. D. Hellmann, A. Eggermont, D. Raoult, L. Albiges, G. Kroemer, L. Zitvogel, Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science*. **359**, 91–97 (2018).
- 410 9. N. R. Rose, Negative selection, epitope mimicry and autoimmunity. *Curr. Opin. Immunol.* **49**, 51–55 (2017).
10. V. Rubio-Godoy, V. Dutoit, Y. Zhao, R. Simon, P. Guillaume, R. Houghten, P. Romero, J.-C. Cerottini, C. Pinilla, D. Valmori, Positional scanning-synthetic peptide library-based analysis of self- and pathogen-derived peptide cross-reactivity with tumor-reactive Melan-A-specific CTL. *J. Immunol. Baltim. Md 1950*. **169**, 5696–5707 (2002).
- 415 11. L. Vujanovic, M. Mandic, W. C. Olson, J. M. Kirkwood, W. J. Storkus, A mycoplasma peptide elicits heteroclitic CD4+ T cell responses against tumor antigen MAGE-A6. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **13**, 6796–6806 (2007).
12. M. E. Perez-Muñoz, P. Joglekar, Y.-J. Shen, Y.-J. Shen, K. Y. Chang, D. A. Peterson, Identification and Phylogeny of the First T Cell Epitope Identified from a Human Gut Bacteroides Species. *PloS One*. **10**, e0144382 (2015).
- 420 13. Y. Yang, M. B. Torchinsky, M. Gobert, H. Xiong, M. Xu, J. L. Linehan, F. Alonzo, C. Ng, A. Chen, X. Lin, A. Sczesnak, J.-J. Liao, V. J. Torres, M. K. Jenkins, J. J. Lafaille, D. R. Littman, Focused specificity of intestinal TH17 cells towards commensal bacterial antigens. *Nature*. **510**, 152–156 (2014).
- 425 14. J. N. Chai, Y. Peng, S. Rengarajan, B. D. Solomon, T. L. Ai, Z. Shen, J. S. A. Perry, K. A. Knoop, T. Tanoue, S. Narushima, K. Honda, C. O. Elson, R. D. Newberry, T. S. Stappenbeck, A. L. Kau, D. A. Peterson, J. G. Fox, C.-S. Hsieh, Helicobacter species are potent drivers of colonic T cell responses in homeostasis and inflammation. *Sci. Immunol.* **2** (2017), doi:10.1126/sciimmunol.aal5068.
- 430 15. Q. Ji, A. Perchet, J. M. Goverman, Viral infection triggers central nervous system autoimmunity via activation of CD8+ T cells expressing dual TCRs. *Nat. Immunol.* **11**, 628–634 (2010).
16. V. P. Balachandran, M. Łuksza, J. N. Zhao, V. Makarov, J. A. Moral, R. Remark, B. Herbst, G. Askan, U. Bhanot, Y. Senbabaoglu, D. K. Wells, C. I. O. Cary, O. Grbovic-Huezo, M. Attiyeh, B. Medina, J. Zhang, J. Loo, J. Saglimbeni, M. Abu-Akeel, R. Zappasodi, N. Riaz, M. Smoragiewicz, Z. L. Kelley, O. Basturk, Australian Pancreatic Cancer Genome Initiative, Garvan Institute of Medical Research, Prince of Wales Hospital, Royal North Shore Hospital, University of Glasgow, St Vincent’s Hospital, QIMR Berghofer Medical Research Institute, University of Melbourne, Centre for Cancer Research, University of Queensland, Institute for Molecular Bioscience, Bankstown Hospital, Liverpool Hospital, Royal Prince Alfred Hospital, Chris O’Brien Lifehouse, Westmead Hospital, Fremantle Hospital, St John of God Healthcare, Royal Adelaide Hospital, Flinders Medical Centre, Envoi Pathology, Princess Alexandria Hospital, Austin Hospital, Johns Hopkins Medical
- 435
- 440

- 445 Institutes, ARC-Net Centre for Applied Research on Cancer, M. Gönen, A. J. Levine, P. J. Allen, D. T. Fearon, M. Merad, S. Gnjatic, C. A. Iacobuzio-Donahue, J. D. Wolchok, R. P. DeMatteo, T. A. Chan, B. D. Greenbaum, T. Merghoub, S. D. Leach, Identification of unique neoantigen qualities in long-term survivors of pancreatic cancer. *Nature*. **551**, 512–516 (2017).
- 450 17. C. P. Bradley, F. Teng, K. M. Felix, T. Sano, D. Naskar, K. E. Block, H. Huang, K. S. Knox, D. R. Littman, H.-J. J. Wu, Segmented Filamentous Bacteria Provoke Lung Autoimmunity by Inducing Gut-Lung Axis Th17 Cells Expressing Dual TCRs. *Cell Host Microbe*. **22**, 697-704.e4 (2017).
- 455 18. S. Viaud, F. Saccheri, G. Mignot, T. Yamazaki, R. Daillère, D. Hannani, D. P. Enot, C. Pfirschke, C. Engblom, M. J. Pittet, A. Schlitzer, F. Ginhoux, L. Apetoh, E. Chachaty, P.-L. Woerther, G. Eberl, M. Bérard, C. Ecobichon, D. Clermont, C. Bizet, V. Gaboriau-Routhiau, N. Cerf-Bensussan, P. Opolon, N. Yessaad, E. Vivier, B. Ryffel, C. O. Elson, J. Doré, G. Kroemer, P. Lepage, I. G. Boneca, F. Ghiringhelli, L. Zitvogel, The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. *Science*. **342**, 971–976 (2013).
- 460 19. G. Y. Lee, P. M. Haverty, L. Li, N. M. Kljavin, R. Bourgon, J. Lee, H. Stern, Z. Modrusan, S. Seshagiri, Z. Zhang, D. Davis, D. Stokoe, J. Settleman, F. J. de Sauvage, R. M. Neve, Comparative oncogenomics identifies PSMB4 and SHMT2 as potential cancer driver genes. *Cancer Res*. **74**, 3114–3126 (2014).
- 465 20. Y.-C. Cheng, W.-C. Tsai, Y.-C. Sung, H.-H. Chang, Y. Chen, Interference with PSMB4 Expression Exerts an Anti-Tumor Effect by Decreasing the Invasion and Proliferation of Human Glioblastoma Cells. *Cell. Physiol. Biochem. Int. J. Exp. Cell. Physiol. Biochem. Pharmacol*. **45**, 819–831 (2018).
- 470 21. X. Zhang, D. Lin, Y. Lin, H. Chen, M. Zou, S. Zhong, X. Yi, S. Han, Proteasome beta-4 subunit contributes to the development of melanoma and is regulated by miR-148b. *Tumour Biol. J. Int. Soc. Oncodevelopmental Biol. Med*. **39**, 1010428317705767 (2017).
22. H. Wang, Z. He, L. Xia, W. Zhang, L. Xu, X. Yue, X. Ru, Y. Xu, PSMB4 overexpression enhances the cell growth and viability of breast cancer cells leading to a poor prognosis. *Oncol. Rep*. **40**, 2343–2352 (2018).
- 475 23. M. G. Weinbauer, Ecology of prokaryotic viruses. *FEMS Microbiol. Rev*. **28**, 127–181 (2004).
24. M. Piuri, G. F. Hatfull, A peptidoglycan hydrolase motif within the mycobacteriophage TM4 tape measure protein promotes efficient infection of stationary phase cells. *Mol. Microbiol*. **62**, 1569–1585 (2006).
- 480 25. M. Belcaid, A. Bergeron, G. Poisson, The evolution of the tape measure protein: units, duplications and losses. *BMC Bioinformatics*. **12 Suppl 9**, S10 (2011).

26. E. Pasolli, L. Schiffer, P. Manghi, A. Renson, V. Obenchain, D. T. Truong, F. Beghini, F. Malik, M. Ramos, J. B. Dowd, C. Huttenhower, M. Morgan, N. Segata, L. Waldron, Accessible, curated metagenomic data through ExperimentHub. *Nat. Methods*. **14**, 1023–1024 (2017).
485
27. P. Ferretti, E. Pasolli, A. Tett, F. Asnicar, V. Gorfer, S. Fedi, F. Armanini, D. T. Truong, S. Manara, M. Zolfo, F. Beghini, R. Bertorelli, V. De Sanctis, I. Bariletti, R. Canto, R. Clementi, M. Cologna, T. Crifò, G. Cusumano, S. Gottardi, C. Innamorati, C. Masè, D. Postai, D. Savoi, S. Duranti, G. A. Lugli, L. Mancabelli, F. Turroni, C. Ferrario, C. Milani, M. Mangifesta, R. Anzalone, A. Viappiani, M. Yassour, H. Vlamakis, R. Xavier, C. M. Collado, O. Koren, S. Tateo, M. Soffiati, A. Pedrotti, M. Ventura, C. Huttenhower, P. Bork, N. Segata, Mother-to-Infant Microbial Transmission from Different Body Sites Shapes the Developing Infant Gut Microbiome. *Cell Host Microbe*. **24**, 133-145.e5 (2018).
490
28. E. Pasolli, F. Asnicar, S. Manara, M. Zolfo, N. Karcher, F. Armanini, F. Beghini, P. Manghi, A. Tett, P. Ghensi, M. C. Collado, B. L. Rice, C. DuLong, X. C. Morgan, C. D. Golden, C. Quince, C. Huttenhower, N. Segata, Extensive Unexplored Human Microbiome Diversity Revealed by Over 150,000 Genomes from Metagenomes Spanning Age, Geography, and Lifestyle. *Cell*. **176**, 649-662.e20 (2019).
495
29. B. Samb-Ba, C. Mazonot, A. Gassama-Sow, G. Dubourg, H. Richet, P. Hugon, J.-C. Lagier, D. Raoult, F. Fenollar, MALDI-TOF identification of the human Gut microbiome in people with and without diarrhea in Senegal. *PLoS One*. **9**, e87419 (2014).
500
30. T. J. Kelly, A. L. Souza, C. B. Clish, P. Puigserver, A hypoxia-induced positive feedback loop promotes hypoxia-inducible factor 1 α stability through miR-210 suppression of glycerol-3-phosphate dehydrogenase 1-like. *Mol. Cell. Biol.* **31**, 2696–2706 (2011).
31. Z. Feng, J. N. Li, L. Wang, Y. F. Pu, Y. Wang, C. B. Guo, The prognostic value of glycerol-3-phosphate dehydrogenase 1-like expression in head and neck squamous cell carcinoma. *Histopathology*. **64**, 348–355 (2014).
505
32. S.-C. Liu, S.-M. Chuang, C.-J. Hsu, C.-H. Tsai, S.-W. Wang, C.-H. Tang, CTGF increases vascular endothelial growth factor-dependent angiogenesis in human synovial fibroblasts by increasing miR-210 expression. *Cell Death Dis.* **5**, e1485 (2014).
510
33. S. Simon, Z. Wu, J. Cruard, V. Vignard, A. Fortun, A. Khammari, B. Dreno, F. Lang, S. J. Rulli, N. Labarriere, TCR Analyses of Two Vast and Shared Melanoma Antigen-Specific T Cell Repertoires: Common and Specific Features. *Front. Immunol.* **9**, 1962 (2018).
34. C. Gil-Cruz, C. Perez-Shibayama, A. De Martin, F. Ronchi, K. van der Borcht, R. Niederer, L. Onder, M. Lütge, M. Novkovic, V. Nindl, G. Ramos, M. Arnoldini, E. M. C. Slack, V. Boivin-Jahns, R. Jahns, M. Wyss, C. Mooser, B. N. Lambrecht, M. T. Maeder, H. Rickli, L. Flatz, U. Eriksson, M. B. Geuking, K. D. McCoy, B. Ludewig, Microbiota-derived peptide mimics drive lethal inflammatory cardiomyopathy. *Science*. **366**, 881–886 (2019).
515
35. T. M. Greiling, C. Dehner, X. Chen, K. Hughes, A. J. Iñiguez, M. Boccitto, D. Z. Ruiz, S. C. Renfroe, S. M. Vieira, W. E. Ruff, S. Sim, C. Kriegel, J. Glanternik, X. Chen, M. Girardi,
520

- P. Degnan, K. H. Costenbader, A. L. Goodman, S. L. Wolin, M. A. Kriegel, Commensal orthologs of the human autoantigen Ro60 as triggers of autoimmunity in lupus. *Sci. Transl. Med.* **10** (2018), doi:10.1126/scitranslmed.aan2306.
- 525 36. M. F. König, L. Abusleme, J. Reinholdt, R. J. Palmer, R. P. Teles, K. Sampson, A. Rosen, P. A. Nigrovic, J. Sokolove, J. T. Giles, N. M. Moutsopoulos, F. Andrade, Aggregatibacter actinomycetemcomitans-induced hypercitrullination links periodontal infection to autoimmunity in rheumatoid arthritis. *Sci. Transl. Med.* **8**, 369ra176 (2016).
- 530 37. J. Li, H. Jia, X. Cai, H. Zhong, Q. Feng, S. Sunagawa, M. Arumugam, J. R. Kultima, E. Prifti, T. Nielsen, A. S. Juncker, C. Manichanh, B. Chen, W. Zhang, F. Levenez, J. Wang, X. Xu, L. Xiao, S. Liang, D. Zhang, Z. Zhang, W. Chen, H. Zhao, J. Y. Al-Aama, S. Edris, H. Yang, J. Wang, T. Hansen, H. B. Nielsen, S. Brunak, K. Kristiansen, F. Guarner, O. Pedersen, J. Doré, S. D. Ehrlich, MetaHIT Consortium, P. Bork, J. Wang, MetaHIT Consortium, An integrated catalog of reference genes in the human gut microbiome. *Nat. Biotechnol.* **32**, 834–841 (2014).
- 535 38. M. L. Pedulla, M. E. Ford, J. M. Houtz, T. Karthikeyan, C. Wadsworth, J. A. Lewis, D. Jacobs-Sera, J. Falbo, J. Gross, N. R. Pannunzio, W. Brucker, V. Kumar, J. Kandasamy, L. Keenan, S. Bardarov, J. Kriakov, J. G. Lawrence, W. R. Jacobs, R. W. Hendrix, G. F. Hatfull, Origins of highly mosaic mycobacteriophage genomes. *Cell.* **113**, 171–182 (2003).
- 540 39. K. E. Wommack, R. R. Colwell, Virioplankton: viruses in aquatic ecosystems. *Microbiol. Mol. Biol. Rev. MMBR.* **64**, 69–114 (2000).

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Competing interests statement: RD, DR, LZ and GK are cofounders of everImmune, a biotech company devoted to the use of commensal microbes for the treatment of cancers. RD is a full-
570 time employee of everImmune. RD and LZ hold patents on immunogenic phage sequences.

Supplementary Materials:

Materials and Methods

Figures S1-S15

Tables S1-S10

575 Reference (1-17)

Statistical report

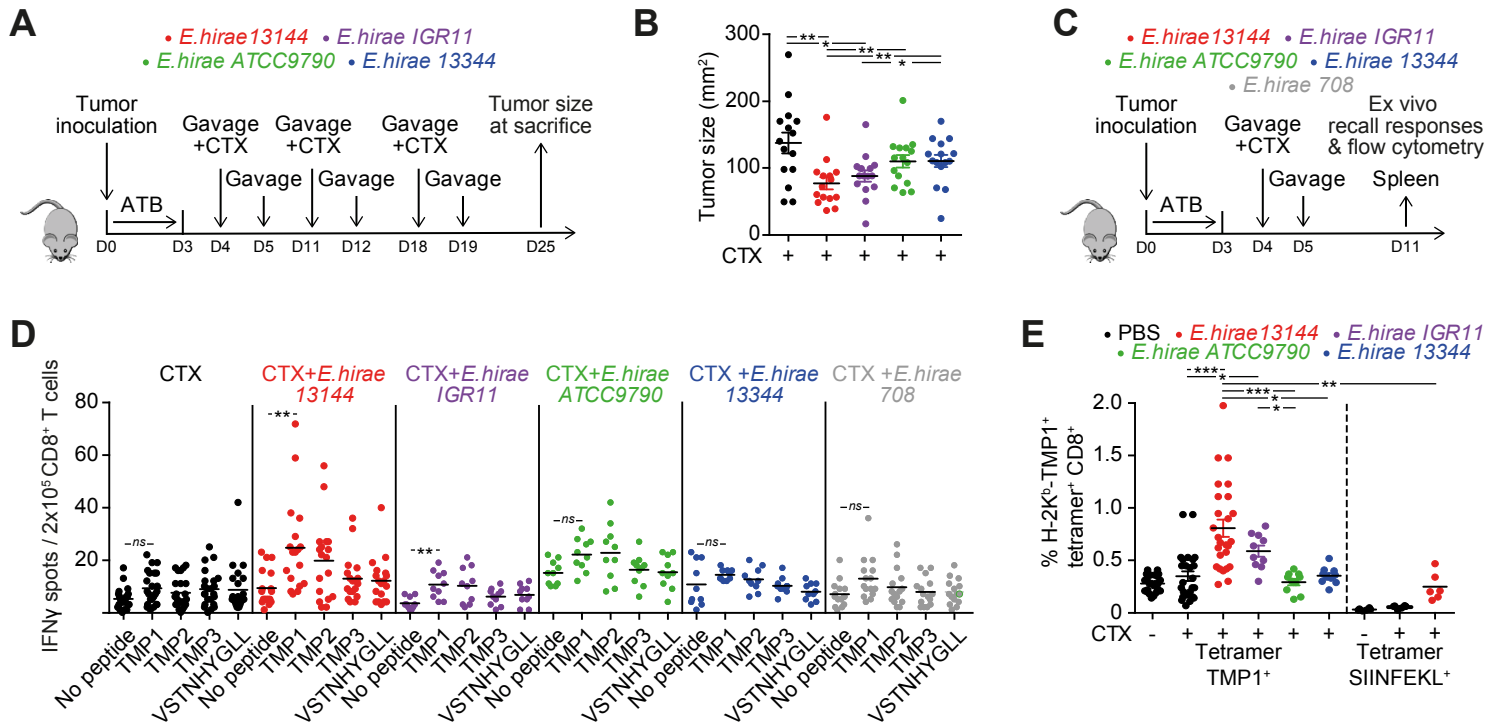


Figure 1

Legends to Figures

Figure 1. Phage Tail Length Tape Measure Protein as the unique antigenic sequence in *E. hirae* 13144.

A, B. C57BL/6 mice bearing MCA205 sarcomas were conditioned with broad spectrum antibiotics (streptomycin, colistin, ampicillin, vancomycin) for 3 days before performing oral gavages with *E. hirae* strain 13144 and i.p. injections of cyclophosphamide (CTX), as indicated (A), and tumor size was recorded for each mouse at sacrifice on day 25 (B). C-E. Naïve C57BL/6 mice were conditioned with antibiotics, gavaged with distinct *E. hirae* strains and treated with CTX (C). Day 11 purified CD8⁺ T splenocytes were restimulated *ex vivo* in a recall assay with bone marrow-derived dendritic cells loaded with the indicated peptides (Table S2, group 7) to quantify IFN γ -secreting CD8⁺T cells (D). H-2K^b/TMP1 (TSLARFANI) or H-2K^b/SIINFEKL tetramer binding CD8⁺ splenocytes were detected by cytofluorometry at day 11 (E). Also refer to Figure S2. Each graph assembles results from 2-3 independent experiments containing groups of 5-6 mice. ANOVA statistical analyses (Kruskal-wallis test): * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. Refer to the statistical report.

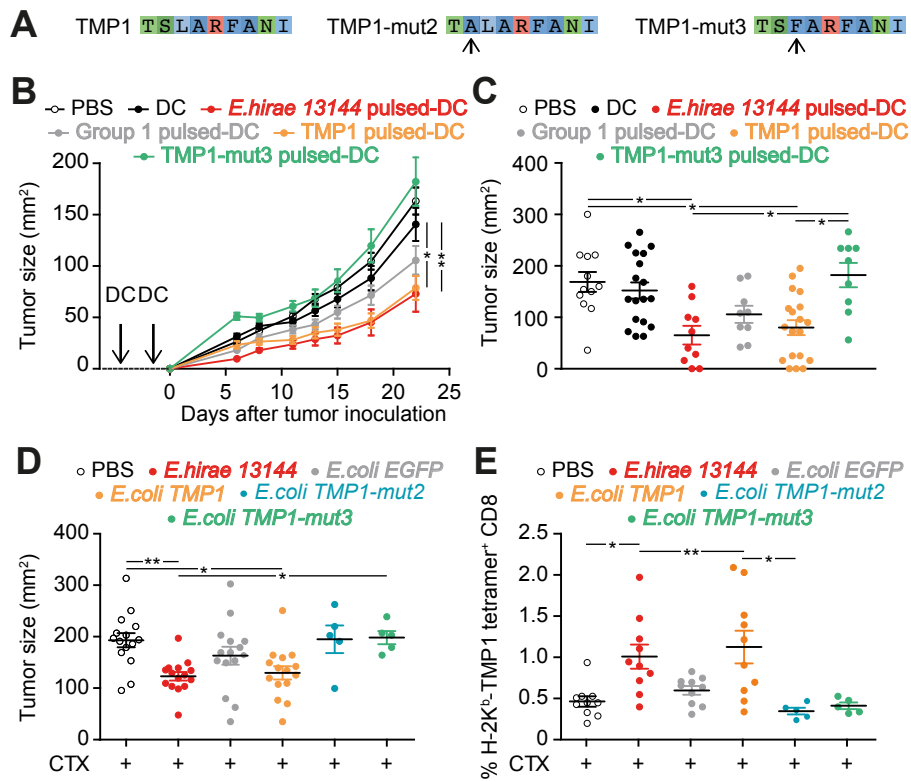


Figure 2

Figure 2. Prophylactic and therapeutic immunization using Phage Tail Length Tape Measure Protein (TMP) against sarcomas.

A. Sequence of the immunogenic epitope TMP1 (TSLARFANI) with the artificial and naturally occurring mutations in positions 2 and 3, respectively. B-C. *Prophylactic vaccinations*. TLR3 ligand-exposed dendritic cell (DC) were pulsed with peptides or heat-inactivated bacteria and then s.c. inoculated twice into mice. One month later, MCA205 sarcomas were implanted in the opposite flank, followed by monitoring of tumor size (means±SEM in B, individual results in C). D-E. *Therapeutic settings*. MCA205 tumor bearing mice were treated with cyclophosphamide (CTX) and gavaged with *E. hirae* 13144 or *E. coli* (like in Fig. 1A) that were genetically modified to express the indicated peptides or enhanced green fluorescent protein (EGFP) as a negative control. Tumor growth at sacrifice (D) and the frequency of H-2K^b/TMP1 tetramer binding splenic CD8⁺ T cells (E) were monitored. Results are shown for 12-18 animals, gathered from 2-3 independent experiments. ANOVA statistical analyses (Kruskal-wallis test): * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. Refer to the statistical report.

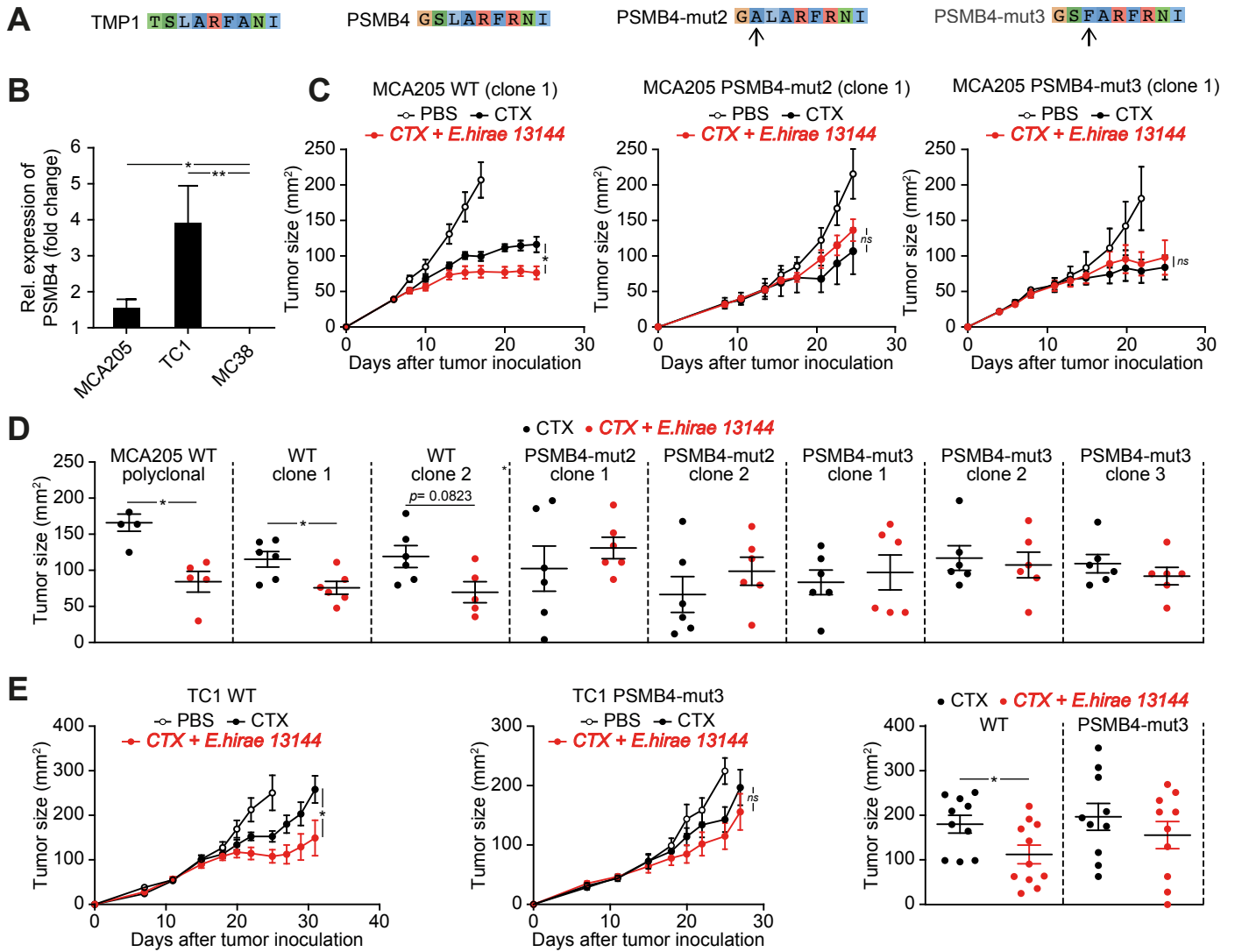


Figure 3

Figure 3. Molecular mimicry between enterophage TMP and the oncogenic driver PSMB4 in two mouse cancers.

A. Sequence alignment of the enterophage TMP1 peptide and a PSMB4 epitope with its two experimental mutants. B. Relative expression of PSMB4 mRNA in MCA205 sarcoma, TC1 lung cancer and MC38 colon carcinomas as compared to their healthy tissue of origin (mean ratio \pm SEM, n=3). C-D. Therapeutic response of wild type *versus* knock-in mutants of MCA205 to cyclophosphamide (CTX) alone or in combination with immunogenic *E. hirae* strain 13144 (setting as in Fig. 1A). Results are shown as tumor growth kinetics (means \pm SEM) for selected MCA205 clones (C) or as individual results (one dot corresponds to one mouse) on day 25 (D). E. Therapeutic response of wild type *versus* mutated TC1 lung cancers to CTX alone or in combination with *E. hirae* 13144 (setting as in Fig. 1A, but without antibiotic preconditioning) reflected by tumor growth kinetics and individual tumor sizes at sacrifice. Results are shown as means \pm SEM. Mann Whitney test or ANOVA statistical analyses (Kruskal-wallis test): * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. Refer to the statistical report.

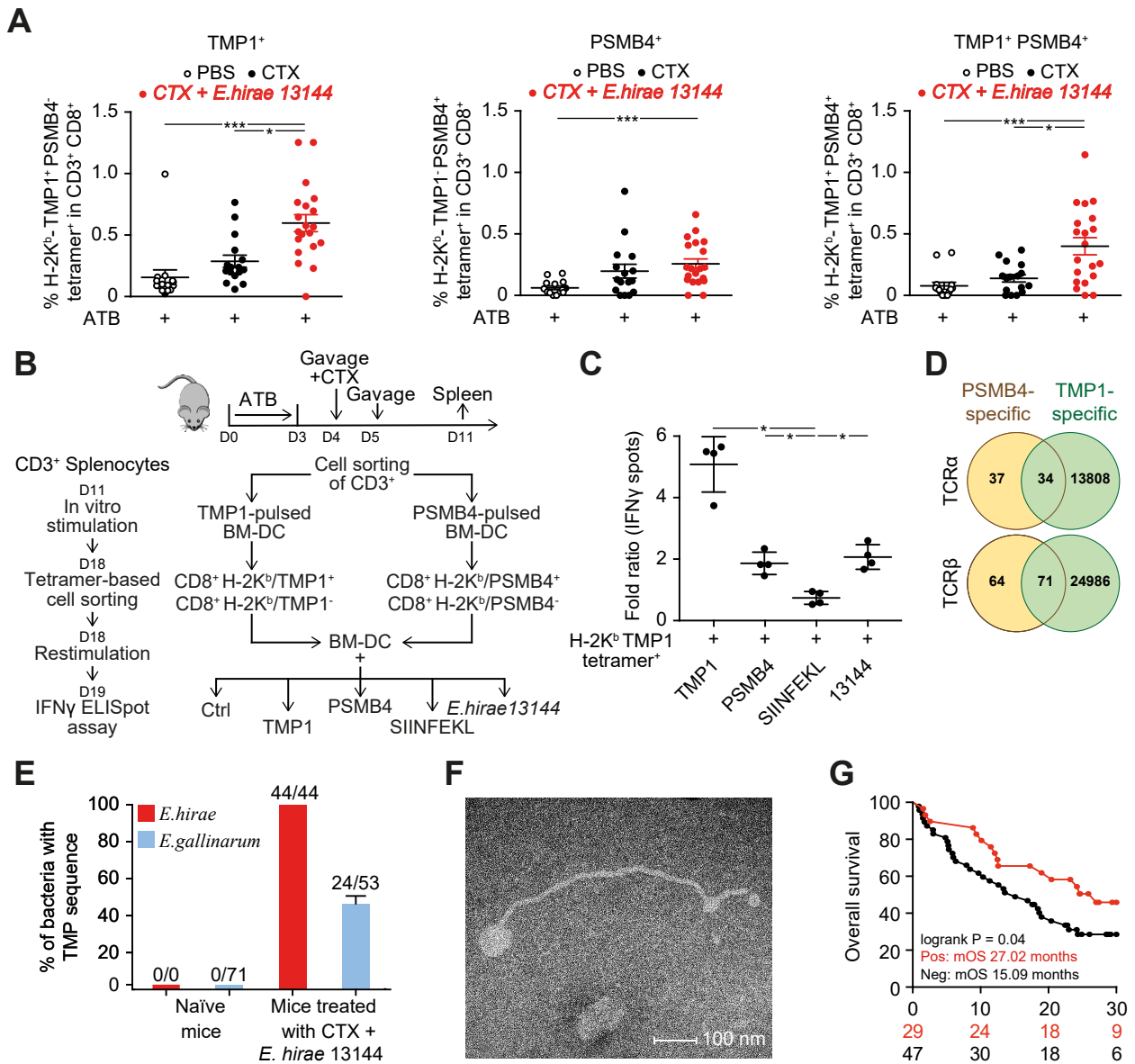


Figure 4

Figure 4. TMP crossreacts with the PSMB4 cancer epitope and affects human anticancer immune responses.

A. Flow cytometry analysis of CD8⁺ tumor-infiltrating lymphocytes (from tumors treated as in Fig. 1A) after co-staining with two different tetramers (H-2K^b/TMP1 and H-2K^b/PSMB4, sequences in Fig. 3A). Each dot depicts one tumor. The graphs assemble the results of 3 independent experiments with 5 mice/group. B,C. Purified CD3⁺ T splenocytes from animals treated with CTX and *E. hirae* 13144 were restimulated *ex vivo* with bone marrow-derived dendritic cells (DC) loaded with TMP1 or PSMB4 peptide. One week after *ex vivo* restimulation, peptide-specific CD8⁺ T cells were purified after staining with the corresponding tetramer to measure IFN γ secretion in response to DC loaded with peptides (TMP1, PSMB4, SIINFEKL as negative control) or heat-inactivated *E. hirae* 13144. These results were performed in parallel on the tetramer-binding *versus* non-binding fraction and were normalized to the PBS controls (Ctrl). Each dot represents one culture. Mann Whitney test or ANOVA statistical analyses (Kruskal-wallis test): * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. D. Venn diagram of TCR α and β chains from tetramer positive CD8⁺ T cells specific for PSMB4 (yellow) or TMP1 (green). E. Lysogenic conversion of *E. gallinarum* by the *E. hirae* siphoviridae phage *in vivo*. Ileal content was obtained from naïve mice or from mice receiving *E. hirae* together with cyclophosphamide (CTX), followed by cultivation and isolation of bacterial colonies, MALDI-TOF identification and PCR-based detection of TMP. Results are from 5 mice/group. F. Transmission electron microscopy of the phage produced by *E. hirae* 13144. G. Kaplan Meier survival plots of 76 patients with non-small cell lung cancer or renal cell cancer subjected to PD-1-targeting immunotherapy, stratified according to the presence or absence of TMP in at least 5 *E. faecalis* or *E. hirae* colonies/patient. Univariate Log-rank (Mantel-Cox) analysis. Refer to the statistical report.



Crossreactivity between MHC class I-restricted antigens from cancer cells and an enterococcal bacteriophage.

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Methods:

Cell culture, reagents and tumor cell lines. MC38, TC1, MCA205 (WT or PSMB4-mutated) tumor cell lines or clones were cultured at 37°C with 5% CO₂ in RPMI 1640 medium containing 10% fetal calf serum (FCS), 2 mM L-glutamine, 100 UI/mL penicillin/streptomycin, 1 mM sodium pyruvate and MEM non-essential amino acids (henceforth referred to as complete RPMI 1640). All these reagents were purchased from Gibco-Invitrogen (Carlsbad, CA, USA).

Mice. All animal experiments were carried out in compliance with French and European laws and guidelines and regulations. The local institutional board approved all mouse experiments (permission number: 2016-109-7450). All mouse experiments were performed at the animal facility in Gustave Roussy Cancer Campus where animals were housed in specific pathogen-free conditions. Female C57BL/6 were purchased from Harlan (Gannat, France). Mice were used at an age between 7 and 12 weeks of age.

Antibiotic treatments. Mice were treated during 3 days (biotinylated) an antibiotic (ATB) solution containing ampicillin (1 mg/mL), streptomycin (5 mg/mL), colistin (1 mg/mL) and vancomycin (0.25 mg/mL) (Sigma-Aldrich) added to the sterile drinking water of mice. Antibiotic activity was confirmed by cultivating fecal pellets resuspended in brain heart infusion (BHI) broth + 15% glycerol at 0.1 g/mL on COS (BD Columbia agar with 5% sheep blood, BioMérieux) plates for 48h at 37°C in aerobic and anaerobic conditions. In the context of bacterial or fecal transplantation, mice received 3 days of ATB before undergoing bacterial or fecal transplantation the next day by oral gavage using animal feeding needles. ATB were not used for Figure 3E and Figure S6E.

Tumor challenge and treatment. Syngeneic C57BL/6 mice were inoculated subcutaneously (s.c.) with 1×10^6 MC38 colon cancer cells, 0.8×10^6 MCA205 sarcoma cells or 0.8×10^6 TC1 lung cancer cells. When tumors reached 20 to 35 mm² in size, the mice were treated intraperitoneally (i.p.) with cyclophosphamide (CTX, 100mg/kg) (Endoxan Baxter, was provided by Institut de Cancérologie Gustave Roussy, Villejuif, France) or anti-PD-1 mAb (250µg/mouse; clone RMP1-14) or isotype control (clone 2A3) (BioXcell, NH, USA). Depending on the experimental setting, mice were injected with CTX once or 3 times at 1-week intervals. Mice

were injected 4 times at 3-day intervals with anti-PD-1 mAb. Tumor size was routinely monitored every 3 days by means of a caliper.

Gut colonization with dedicated commensal species. *Enterococcus hirae* 13144 were originally isolated from spleens of SPF mice treated with CTX in our laboratory. *E. hirae* 708 was provided by INRA (P. Langella), while *E. hirae* 13344, ATCC9790 were provided by Prof. Cattoir, CHU de Caen, France. *L. plantarum* was provided by Prof. Ivo Gomperts Boneca from the Institut Pasteur strain repository, France. All *E. hirae* IGR strains were isolated from the stools of NSCLC patients in our laboratory, according to patient informed consent and local IRB approval (ancillary study "Oncobiotics"). All bacteria were grown in COS plates for 24 to 48 hours at 37°C in aerobic conditions. Colonization of ATB pre-treated mice was performed by oral gavage with 100 µl of suspension containing 1×10^9 bacteria. For bacterial gavage, we used suspensions of 10^{10} CFU/mL, monitored using a fluorescence spectrophotometer (Eppendorf) at an optical density of 600 nm in PBS. Depending on the experimental setting, 2 or 6 bacterial gavages were performed for each mouse: the first, the same day as CTX injection, and then 24 hours after the injection of CTX. For anti-PD1 mAb, 5 bacterial oral gavages were performed for each mouse: the first, the same day and 24h before the first anti-PD1 injection, and the same day for the three other injections of anti-PD1 Abs. The efficacy of colonization was confirmed by culturing the feces 48 hours post-gavage. Fecal pellets were harvested and resuspended in BHI+15% glycerol at 0.1 g/mL. Serial dilutions of feces were plated onto COS plates and incubated for 48 hours at 37°C in aerobic and anaerobic conditions. After 48 hours, the identification of specific bacteria was accomplished using a Matrix-Assisted Laser Desorption/Ionisation Time of Flight (MALDI-TOF) mass spectrometer (Andromas, Beckman Coulter, France).

Culture and propagation of bone marrow-derived dendritic cells. Bone marrow-derived dendritic cells (BM-DCs) were generated by flushing bone marrow precursors from the femurs and tibia of female C57Bl/6 WT mice aged between 8 and 12 weeks. Bones were collected in sterile PBS, washed in alcohol and Iscove's medium (IMDM, Sigma-Aldrich) baths, extremities of bones were cut and flushed using a 26G needle. After red blood cell lysis, cells were cultured in IMDM supplemented with 10% of FCS + 2mM L-glutamine + 100 UI/mL penicillin/streptomycin + 50µM 2-mercaptoethanol (Sigma-Aldrich) (referred herein as complete

IMDM medium) at 0.5×10^6 /mL and treated with 40ng/mL of GM-CSF (supernatant of GM-CSF transfected-cells J558) and 10 ng/mL of recombinant interleukin-4 (IL-4) for BM-DCs (from Peprotech). Cells were split at day 3 and used in experiments on day 7 or 8.

Test of memory TC1 immune response and H-2K^b restricted-peptides on splenic CD8⁺ T cells. Interferon- γ (IFN- γ) ELISPOT assay were performed in 96-well PVDF bottomed sterile plates (Millipore MSIP S4510) by means of a commercial kit (Cell sciences, Newburyport, US) according to the manufacturer's instructions. After PVDF membrane activation with ethanol 35%, plates were coated overnight with capture antibody to IFN- γ and washed before incubation of blocking buffer during 2 hours. BM-DC (1×10^5 /well) were exposed to heat-inactivated (2 hours at 65°C) bacterial strains (*E. hirae* 13144, *E.hirae* 708, *E.hirae* 13344 and *L.plantarum* at a multiplicity of infection [MOI] of 1:10) or pulsed with peptides (20 μ g/mL) and were added to CD8⁺ T cells (2×10^5 /well) for 20 hours at 37°C. Cells were then removed and plates were developed with a biotinylated antibody specific for IFN- γ during 1 hour and 30 minutes, followed by streptavidin-alkaline phosphatase during 1 hour. Finally, the substrate of streptavidin (BCIP/NBT buffer) was added for 5-20 min. Spots were counted by means of a CTL Immunospot Analyzer (Cellular Technology Limited, Cleveland, OH).

Vaccination of mice. BM-DCs were activated with poly I:C (10 μ g/mL, Invivogen) overnight before infection with heat-inactivated (2 hours at 65°C) bacterial strains (MOI 10) or pulsed with peptides (20 μ g/mL, peptide 2.0). After 6 hours of incubation with bacteria or 1 hour of incubation with peptides, BM-DCs were washed 3 times with PBS before subcutaneous injection in the right flank of mice (1.5×10^5 cells per mice). Mice were vaccinated twice at 10 days apart and challenged 4 weeks after the second vaccination with the minimal tumorigenic dose of MCA205 tumor cells in left flank.

Flow cytometry analyses. In experiments without tumor, spleens were harvested 7 days after the injection of CTX. In tumor growth experiments, spleens, tumors and tumor draining lymph node were harvested at different time points, 7, 14 and 21 days after the first injection of CTX into mice bearing MCA205 tumors. Excised tumors were cut into small pieces and digested in RPMI medium containing LiberaseTM at 25 μ g/mL and DNase1 at 150 UI/mL (Roche) for 30 minutes at

37°C and then crushed and filtered twice using 100 and 40µm cell strainers (Becton & Dickinson, BD). Lymph nodes and spleen were crushed in RPMI medium and subsequently filtered through a 70 µm cell strainer. Two million splenocytes, tumor cells or lymph node cells were pre-incubated with purified antimouse CD16/CD32 (clone 93; eBioscience) for 15 minutes at 4°C, before membrane staining. Dead cells were excluded using the Live/Dead Fixable Yellow dead cell stain kit (Life Technologies). Anti-mouse antibodies for CD3 (145-2C11), CD4 (GK1.5), CD8 (eBioH35-17.2), CXCR3 (CXCR3-173), CCR9 (CW-1.2), and TMP specific tetramer (BD, BioLegend, eBioscience and Cliniscience). Stained samples were acquired on Canto II 7 colors cytometer (BD) and analyses were performed with FlowJo software (Tree Star, Ashland, OR, USA).

Human T cell responses to HLA-A*0201 restricted-TMP epitopes. Cytapheresis cones were collected from healthy volunteers (Etablissement français du sang, EFS) and peripheral blood mononuclear cells (PBMC) were separated using a Ficoll Hypaque (Sigma Aldrich) gradient. We selected only donors with the HLA-A02*01 haplotype determined by immunofluorescence and flow cytometry. PBMC were washed and resuspended in the separation medium (PBS, 1mM ethylenediaminetetraacetic acid, 2% human AB⁺ serum) for magnetic bead separation. CD14⁺ monocytic cells (human CD14 MicroBeads, Miltenyi) were enriched from 75×10^6 peripheral blood mononuclear cells (PBMC) and cultured at 0.5×10^6 /mL in IMDM supplemented with 10% human AB⁺ serum, 1% of 2 mmol/L glutamine (GIBCO Invitrogen), 1000 IU/mL GM-CSF and 1000 IU/mL IL-4 (Miltenyi). Cells were split at day 3 and used in experiments on day 6 or 7. Such (DC-like) cells were seeded in 96-well plates at 1×10^5 cells/well either alone or in the presence of peptides (20µg/mL) for 2 hours at 37°C, 5% CO₂. The remaining autologous PBMC fractions were enriched for CD8⁺ T cells (CD8⁺ T Cell Isolation Kit, human, Miltenyi). The enriched CD8⁺ T cells were washed, counted and resuspended at 1×10^5 cells/well in RPMI-1640 supplemented with 10% human AB⁺ serum, 1% 2 mMol/L glutamine, 1% penicillin/streptomycin (GIBCO Invitrogen) and 50 U/mL IL-2 (Proleukin). DC-peptide/ T cell co-cultures were incubated for one week at 37°C, 5% CO₂ (medium was changed every 2 days). Then, the pools of cells were seeded in 96-well ELISpot plates at 2×10^5 cells/well and restimulated with or without peptides (20µg/mL) or anti-CD3/anti-CD28 coated beads (1µL/mL, Dynabeads T-Activator, Invitrogen) as a positive control for 20 hours at 37°C. IFN-γ ELISPOT

assays were performed in 96-well PVDF bottomed sterile plates (Millipore MSIP S4510) by using a IFN- γ ELISPOT kit (Cell sciences, Newburyport, Etats-Unis) according to the manufacturer's instructions.

In vitro stimulation of PBMCs from healthy volunteers and cancer patients with HLA-A2-restricted phage and cancer peptides. Cytapheresis cones were collected from healthy volunteers (EFS) and peripheral blood mononuclear cells (PBMC) were separated using a Ficoll Hypaque gradient. We selected only donors with HLA-A02*01 haplotype determined by flow cytometry with anti-HLA-A2 antibody (BB7-2 clone). 4×10^7 PBMC were seeded in 96 well/plates at 2×10^5 cells/well in RPMI 1640 medium supplemented with 8% human serum (HS), 50 IU/mL of IL-2 (Proleukin, Novartis) and stimulated with 5 μ M of KLAKFASVV peptide. After 14 days, each microculture was evaluated for the percentage of specific CD8⁺ T lymphocytes by double staining with a HLA-A2- KLAKFASVV peptide tetramer and anti-CD8 mAb (Clone RPA-T8, Biolegend) using a FACS Canto HTS. Cross-reactivity of positive microcultures was evaluated by double staining with a HLA-A2 KLQKFASTV peptide tetramer and anti-CD8. HLA-A2 /peptide monomers were produced by the recombinant platform facility P2R, from SFR Santé, as previously described (1). Microcultures that contained at least 0.5% of specific T cells were selected, pooled and sorted with the relevant tetramer-coated beads and amplified as previously described (2). After the amplification step, purity and cross-reactivity of sorted T cell lines were evaluated by tetramer/CD8 double labeling. CD107A mobilization and TNF α production were evaluated after stimulation of sorted T cells with 5 μ M of each peptide (KLAKFASVV or KLQKFASTV). After a 5 hour-stimulation period in the presence of brefeldin A at 10 μ g/mL (Sigma, B7651), T cells were labeled with phycoerythrin (PE)-conjugated anti-CD8 antibody (Clone RPA-T8, Biolegend) and fixed with PBS 4% paraformaldehyde (VWR, 100504-858). Lymphocytes were then stained for TNF production using APC conjugated anti-TNF α (clone Mab11, Biolegend). Concerning CD107A labeling, specific T cells were stimulated for 3 hours at 37°C in the presence of Alexa-F647-conjugated mAb specific for CD107A (clone H4A3, Biolegend). T cells were then stained with anti-CD8 antibody (Clone RPA-T8, Biolegend) and analyzed by flow cytometry.

Short-term Ag-specific T cell lines from HLA-A*0201 lung cancer patients. Peripheral blood was collected from non-small cell lung cancer (NSCLC) patients at the time of surgery, after informed consent and PBMC were isolated on a Ficoll Hypaque gradient. Only PBMC from patients bearing the HLA-A*02*01 haplotype were used for *in vitro* short-term Ag-specific stimulation. CD8⁺ T cells were positively enriched using an anti-CD8-coated magnetic microbeads (Miltenyi Biotec) selection process resulting in more than 95% purity. CD8⁺ T cells were seeded in 96-well plates at 2×10^5 cells/well in RPMI-1640 medium supplemented with 10% human serum. Autologous CD8-depleted PBMC were used as antigen presenting cells (APCs), irradiated, pulsed with TMP epitope 10 (KLAKFASVV) (5 µg/mL) for 2 hours at 37°C in 5% CO₂ and plated with CD8⁺ T cells at a 1:3 ratio. After 24 hours, human recombinant IL-2 (Miltenyi Biotec) and IL-7 (PeproTech Inc.) (25 U/mL and 5 ng/mL, respectively) were added to the culture wells. After one week, cells were restimulated with the soluble TMP epitope 10 peptide (1 µg/mL) for an additional week before functional analysis.

HLA-A2/peptide tetramer staining. Phycoerythrin (PE)-labeled HLA-A*0201/peptide (KLAKFASVV) tetramers were used. FITC-CD8 monoclonal antibody was purchased from Miltenyi Biotec (BW135/80). Briefly, $2-3 \times 10^5$ short-term *in-vitro* expanded T cells were first incubated with tetramer (10 µg/mL, 30 minutes, room temperature). After washing, cells were incubated with FITC-CD8 mAb (20 minutes, 4°C). Dead cells were excluded using propidium iodide staining (MP Biomedicals). Cells were immediately acquired on BD FACS Celesta and analyzed using FACS Diva software (BD).

Interferon (IFN)- γ and granzyme B (GrB) production. $3-4 \times 10^5$ autologous CD8-depleted PBMC isolated from NSCLC patients were pulsed with the relevant TMP epitope 10 (KLAKFASVV) or the irrelevant TMP epitope 14 (KMAALAASA) (1 µg/mL) (Figure S13E). Alternatively, instead of autologous APC, T2 cell lines (purchased from the American Type Culture Collection and routinely checked for mycoplasma using Mycoplasma PCR Reagent, Euroclone, Italy) were used at 1×10^5 cells/well and pulsed (or not) with the relevant TMP epitope 10, the irrelevant TMP epitope 14, or with irrelevant MART-1 A27L (ELAGIGILTV) and gp100 209-217 (IMDQVPFSV) peptides (1 µg/mL) (Figure S13F). Whenever indicated, the HLA Class I-blocking antibody (W6/32 mAb) was added (3). After 1 hour incubation at 37°C in

5% CO₂, autologous 2-3 x 10⁵ CD8⁺ T-cell lines were added to monitor antigen-specific activation markers, IFN- γ and GrB production. Cells were co-cultured for 5 hours at 37°C in 5% CO₂, in the presence of the protein transport inhibitor GolgiStop (BD). After 5 h, T cells were collected, washed in PBS and incubated for 30 minutes at 4°C with the following mAbs from BD Biosciences: PE-CD3 (SP34-2), APC-H7-CD8 (SK1), FITC-CD4 (SK3), BV786-CD137 (4B4-1). After washing, cells were fixed and permeabilized by means of the Cytotfix/Cytoperm kit (BD Biosciences), following the manufacturer's instructions. Intracellular staining was performed for 30 minutes at room temperature by the use of following mAbs from BD Biosciences: PE-Cy7-IFN- γ (B27), and Alexa Fluor647-GrB (GB11). Cells were immediately acquired on a FACS Celesta and analyzed using FACS Diva software (BD).

MART-1 and MELOE-1-specific T cell clones and responses to bacterial peptides.

Principles.

We identified microbial analogs of non-mutated tumor-associated antigens relevant to human malignancies (such as the MART-1/Melan-A melanoma differentiation antigen or MELOE-1 aberrantly expressed antigen). In the public microbiome database (metaHIT), 5 and 11 microbial sequences shared more than 78% homology with EAAGIGILTV (from MART-1/Melan-A) (Figure S14A) and TLNDECWPA (from MELOE-1) (Figure S15A), respectively. The cross-reactivity of 11 MART-1/Melan-A -specific T-cell clones to each of the 5 bacterial peptides was measured. All the 11 T cell clones recognized 2 out of the 5 bacterial peptides with EC₅₀ values similar to those found for the MART-1/Melan-A-AA27L peptide (Figure S14B). Another bacterial peptide was recognized by 2 of the 11 T cell clones, though with a low affinity (Fig S14B). The cross-reactivity of all the MART-1/Melan-A-specific T-cell clones tested might be linked to the frequent occurrence of TRAV12-2 segments (which are highly flexible) (33) within the alpha chains of their TCRs (Table S8). We also evaluated 11 microbial peptides for their capacity to stimulate 10 MELOE-1 specific T cell clones, which exhibited a bias towards another TRAV segment (TRAV19, Table S9). Four out of 10 MELOE-1 specific T cell clones responded to at least 1 bacterial peptide (Figure S15B). One of these peptides, differing from the cognate peptide at positions 6 (P6) and 8 (P8) (predicted as weak binder to the HLA-A2 molecule), was recognized by 3 MELOE-1-specific T-cell clones with EC₅₀ values similar to the one observed for the WT MELOE-1 peptide (Figure S15B). The two other analogous peptides with different P6

and P8 residues were also recognized by two T cell clones, with an EC_{50} around 10^{-9} M (Figure S15B), suggesting that these two positions are not essential for TCR recognition.

Calculation of EC_{50} for bacterial epitopes.

EC_{50} of MART-1 and MELOE-1-specific T-cell clones were evaluated for each bacterial peptide, by measuring TNF α production after co-culture with TAP-deficient T2 cells loaded with a range of peptides, at an effector/target ratio of 1:2. After a 5h-stimulation period in the presence of brefeldin A at 10 μ g/mL (Sigma, B7651), T cells were labeled with PE-conjugated specific anti-CD8 antibody (Clone RPA-T8, BioLegend) and fixed with PBS 4% paraformaldehyde (VWR, 100504-858). Lymphocytes were then stained for cytokine production using APC conjugated anti-TNF α (clone cA2, Miltenyi Biotec).

Stool detection of phage TMP sequence by PCR in human or mouse samples (Figure S10).

We cultivated the stools (from cancer patients) or ileal material (mice) after several dilutions in aerobic conditions and permissive medium to allow for the isolation of enterococci colonies (according to a procedure described in (4)). We performed a PCR of the TMP sequence in each single cultivatable *Enterococcus* colony. One colony was placed into 100 μ l of nuclease-free water to release the bacterial DNA and PCR was performed with 5 μ l of DNA, 12.5 μ l of PCR master mix (ThermoFischer Scientific), 5 μ l nuclease-free water and 1.25 μ l of pairs of TMP primers (20 μ M) (refer to Figure S10 for the position of the probe sets). PCR products were separated on 1.5% agarose gel containing ethidium bromide and revealed by UV exposure. The sequence of primers are: forward 5'-ACTGCAGCCGTA AAAATGGGA-3' and reverse 5'-TCCGTATCGTTTGCCAGCTT-3' (amplicon 1026 bp).

Lysogenic conversion of *E. gallinarum* by the *E. hirae* 13144 phage.

In vivo. To investigate the capacity of the *E. hirae* 13144 phage to lysogenize other bacterial species *in vivo*, we performed culturomic analyses of the ileal content from C57BL/6 mice subjected to oral gavage with *E. hirae* 13144 and systemic CTX therapy, followed by PCR analyses seeking TMP sequences (Figure S8A-B). We tested 7 to 18 bacterial colonies from each animal and a total of 76 colonies. We only found lysogenic conversion of *E. gallinarum* by the *E. hirae*-temperate phage *in vivo*, as confirmed by sequencing of the phage genome in the second

host (Figure 4F, Figure S8B-C). In contrast, none of the 90 colonies (mostly of *E. gallinarum*) isolated from naive mice harbored the TMP sequence (Figure S8A).

In vitro. One *E. gallinarum* strain (isolated from naïve mice) were incubated at a ratio of 1:1 (10^7 of each bacteria), in the presence of small intestinal organoids, during one hour before treatment with mafosfomide (25µg/mL). Six and twenty hours post-incubation, organoid supernatants were plated to allow for the isolation of *E.gallinarum* colonies followed by PCR-based detection of the TMP sequence on each *E.gallinarum* colony. For preparation of small intestine organoids, ileal intestinal crypts were isolated and enriched from 8-12 week old C57BL/6 mice as previously described (5) with the following modifications. Briefly, pieces of ileum washed in PBS were incubated in PBS containing 2mM EDTA for 30 minutes on ice. Fragments were then rinsed 3 times with PBS containing 10% FCS and filtered through a 70-µm cell strainer. Crypts were pelleted, washed with Advanced DMEM/F12 (ADF) (Invitrogen), resuspended in 1mL of Cultrex PathClear Reduced Growth Factor BME (Bio-Techne, Minnesota, United States) and 50µL drops were pipetted into a 24 well plate. Drops were overlaid with ADF containing the following: 100 U/mL penicillin G sodium, 100 µg/mL streptomycin sulfate, 2 mM L-glutamine, 10 mM HEPES, 1x N2 supplement, 1x B27 supplement, 50 ng/mL mEGF, 100 ng/mL mNoggin (Peprotech, Hamburg, Germany), N-acetylcysteine (Sigma) (reagents from Invitrogen unless otherwise indicated) and 10% conditioned medium of Cultrex® HA-R-Spondin1-Fc 293T Cells (Bio-Techne). Organoids were passaged once per week and utilized in experiments 7 days post splitting.

Transmission electron micrographs of the bacteriophage. The negative staining of particles was realized on supernatant of a co-culture of *E. hirae* 13144 and *E. gallinarum* admixed at a 1:10 ratio for 20 hours. Staining was performed using a 5% solution of ammonium molybdate. Images were acquired using a Tecnai G2, operating at 200 keV. Scale bars are shown on micrograph pictures.

Generation of TMP-expressing *E. coli*. A DNA fragment containing the P23 promoter sequence was generated by annealing two complementary primers (5'-CAATAAAAATCAGACCTAAGACTGATGACAAAAAGAGCAAATTTTGATAAAATAG TATTAGAATTAAATTA AAAAGGGAGGCCAAATATAG-3' and 5'-

GATCCTATATTTGGCCTCCCTTTTTAATTTAATTCTAATACTATTTTATCAAAATTTGC TCTTTTTGTCATCAGTCTTAGGTCTGATTTTTTATTGCATG-3'). The sequence was then inserted into SphI/BamHI-digested vector pDL278 (Addgene 46882, gift from Gary Dunny) (6) to generate vector pDL278-P23. A part of the TMP gene (N-terminal 1185 nucleotides of TMP, including the epitope TSLARFANI, fused to a C-terminal FLAG-tag) was amplified from *E. hirae* 13144 genomic DNA (5'-TCCGGATCCATGGCACAAAGTAAAACAGTCAAAGCG-3', 5'-

CAGGAATTCTTACTTGTCGTCATCGTCTTTGTAGTCACGTAGTAAACTATCACGTAAT CGAACTTC-3') and inserted into BamHI/EcoRI-digested vector pDL278-P23 to generate vector pDL278-P23-TMP-FLAG. Mutations in the epitope were introduced using the QuikChange Lightning Kit (Agilent). Primers 5'-AACGAGCTAAGGCAGTAGCAGCTGTATCTGCAGAC-3' and 5'-GTCTGCAGATACAGCTGCTACTGCCTTAGCTCGTT-3' were used to mutate position 2 (S to A, pDL278-P23-TMP-mut2-FLAG), primers 5'-ATTAGCAAACGAGCGAAGGAAGTAGCAGCTGTATCTG-3' and 5'-CAGATACAGCTGCTACTTCCTTCGCTCGTTTTGCTAAT-3' were used to mutate position 3 (L to F, pDL278-P23-TMP-mut3-FLAG). To generate the control plasmid pDL278-P23-EGFP, EGFP was amplified from pCIB1(deltaNLS)-pmGFP (Addgene 28240, gift from Chandra Tucker (7)) using primers 5'-CTTGATCCATGGTGAGCAAGGGCGAG-3' and 5'-CAGGAATTCCTACATAATTACACACTTTGTC-3' and inserted into BamHI/EcoRI-digested vector pDL278-P23. Plasmids were transformed into chemically competent *E. coli* DH5 α (NEB) and the presence of plasmids with the correct insert was verified by sequence analysis (5'-CCCAGTCACGACGTTGTAAAACG-3' and 5'-GAGCGGATAACAATTTACACAGG-3'). Expression of EGFP and TMP-FLAG in *E. coli* was verified by western blot analysis using antibodies targeting GFP (Cell Signaling, 2956) or FLAG (Sigma-Aldrich, F7425), respectively.

CRISPR/Cas9-mediated mutations of mouse *Psemb4* in MCA205 and TC1 lung cancer cells.

Wild type MCA205 cell line was purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA) and was maintained in RPMI-1640 medium (Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 10% FBS (Thermo Fisher Scientific, Inc), 100 U/mL penicillin and 100 μ g/mL streptomycin (Thermo Fisher Scientific, Inc.) at 37°C. For the CRISPR knock in mutations, we designed the gRNA (sequence AGATATTGCGGAAACGAGCC) by using the CRISPR design tool developed by the Zhang lab (<http://crispr.mit.edu/>).

Oligonucleotides containing the designed sequence were synthesized (Sigma) and ligated into the pX458 backbone (Addgene #48138, (8)) containing the Cas9 gene (human codon-optimised and fused with 2A-GFP allowing for selection) under a CBh promoter and the cloned sgRNA under a U6 promoter. Homology templates (sequence attached) containing the mutation sites were synthesized by Invitrogen GeneArt Gene Synthesis (Thermo Fisher Scientific, Inc.). The cloned pX458 plasmid and synthesized homology arms were cotransfected into MCA205 cells by means of lipofectamine 3000 (Thermo Fisher Scientific, Inc.) following the manufacturer's protocol. Forty-eight hours after transfection, GFP-positive cells were sorted to 96-well plates as single cells before surviving clones were expanded in duplicated conditions, one for frozen storage at -80 and the other for genomic DNA extraction. The targeted region in genomic DNA from clones was further amplified by PCR using the Phusion® High-Fidelity PCR Master Mix (New England BioLabs; Ipswich, MA, USA) and primers 5'CTCAGGGACCCTTTTCACGA 3' and 5'CCCACTCCCTGTTCTACACA 3', and purified with the Monarch® DNA Gel Extraction Kit (New England BioLabs) before being sent to Eurofins Genomics GmbH (BERSBERG GERMANY) for sequencing with the primer 5'GGACCCTTTTCACGATTCAGG 3'. Positive clones were expanded and subjected to DNA extraction for further validating the mutations.

Genome sequencing and analysis. The whole genome sequence of 5 *E. hirae* (13144, 708, 13152, 13344 and EH-17) strains was determined with PacBio technology (GATC Biotech, Konstanz, Germany). Genomic DNA was isolated from 15 other *E. hirae* isolates using the Quick-DNA fungal/bacterial miniprep kit (Zymo Research, Irvine, CA) according to the manufacturer's recommendations. After DNA shearing, the DNA libraries were prepared using the NEBNext Ultra DNA library prep kit for Illumina (New England Biolabs, Ipswich, MA) and sequenced as paired-end reads (2 x 300 bp) using an Illumina MiSeq platform and the MiSeq reagent kit version 3. The Illumina reads were trimmed using Trimmomatic (9), quality filtered with the Fastx-toolkit (http://hannonlab.cshl.edu/fastx_toolkit/) and assembled using SPAdes (10). Protein sequences were predicted using prokka v1.11 software (11). Prophage regions were detected using PHAST software. Predicted proteins were annotated using BLASTp against the National Center for Biotechnology Information (NCBI) non-redundant (NR) database.

Phylogenomic and comparative genomics. Single nucleotide polymorphisms in 20 *E. hirae* genomes was investigated using the parsnp program (12) and using the genome of strain 13144

genome as a reference. Phylogenetic analysis was performed by considering the 47,303 polymorphic sites retained in the core genome of the 20 genomes. Maximum likelihood phylogeny was constructed using Fasttree (13). Phylogenetic tree was visualized using figtree (<http://tree.bio.ed.ac.uk/software/figtree/>). Complete proteome sequences of 20 *E. hirae* strains were compared using BlastP and pairwise alignments using ClustalW. We clustered the *E. hirae* homologous genes using orthoMCL (14) on the translated protein sequences of all predicted genes with a conservative parameter value of 70% amino acid sequence identity and 50% sequence coverage. The determination of the different unique core genomes was based on the homology clusters found by orthoMCL.

TCR sequencing of TMP1- and PSMB4-specific CD8⁺ T cells. H-2K^b-TMP1 tetramer binding CD8⁺T cells were isolated from spleen, tumor draining lymph nodes and MCA205 tumor beds after animal exposure to CTX+*E.hirae* 13144, using FACS cell sorting and were pooled into two fractions (positive or negative for the H-2K^b-TMP1 staining, regardless of tissue location). H-2K^b-PSMB4 tetramer binding CD8⁺T cells from tumor draining lymph nodes and MCA205 tumor beds were cell sorted by FACS after exposure of the animal to CTX+*E. hirae* 13144 and were pooled into 2 fractions (positive or negative for the H-2K^b-PSMB4 staining, regardless of tissue location). Moreover, H-2K^b-PSMB4 tetramer binding CD8⁺T cells were harvested (cell sorted by FACS) from vaccine draining lymph nodes after immunization of naive mice with PSMB4 peptides admixed with TLR3 ligands. RNA from those T cell pools (positive and negative fractions) were isolated by means of lysis buffer with the RNAqueous-Kit (Invitrogen®) extraction kit, according to the manufacturer's protocol. The RNA concentration and sample integrity were determined on Nanodrop (ThermoFisher). T cell receptor (TCR) libraries were prepared with the RNA from each sample with SMARTer Human TCR α/β Profiling Kit (TakaraBio) following the provider's protocol. Briefly, the reverse transcription was performed using TRBC reverse primers and further extended with a template-switching oligonucleotide (SMART-Seq v4). cDNAs were then amplified following two semi-nested PCR: a first PCR with TRBC and TRAC reverse primers as well as a forward primer hybridizing to the SMART-Seqv4 sequence added by template-switching and a second PCR targeting the PCR1 amplicons with reverse and forward primer including Illumina Indexes allowing for sample barcoding. PCR2 are then purified using AMPure beads (Beckman-Coulter). The quantification and integrity of cDNA

samples was carried out using DNA electrophoresis performed on Agilent 2100 Bioanalyser System in combination with the Agilent DNA 1000 kit, according to the manufacturer's protocol. Sequencing has been performed with Miseq (Illumina) SR-300 protocols at Institut du Cerveau et de la Moelle (Paris, France). FASTQ raw data files were processed for TRAs and TRBs sequences annotation using MiXCR (15) software (v2.1.10) with RNA-Seq parameter. MiXCR extracts TRA and TRB providing corrections of PCR and sequencing errors. Generation of datasets was done by concatenating the FASTQ raw data files based on the specificity of the different sorted cell population samples from the different organs. We obtained 4 datasets representing $CD8^+TMP^+$ (meaning CD8 binding to H-2K^b-TMP1 tetramer) $CD8^+TMP^-$, $CD8^+PSMB4^+$ and $CD8^+PSMB4^-$ TCR repertoires. These repertoires were respectively composed of 40.734, 416.541, 208 and 532.360 unique clonotypes. Venn diagrams and samples comparisons were performed using R software version 3.5.0 (www.r-project.org) and Prism (GraphPad Software, LaJolla, CA). To compare the TCR sharing of $PSMB4^+$ with TMP^+ vs TMP^- TCRs, a random sampling of 13842 α TCRs and 25057 β TCRs was performed 10 times within the TMP^- repertoire (Fig. 4H).

Statistical analyses. A statistical report has been written for each panel (online material). Data analyses and representations were performed with Prism 6 software (GraphPad, San Diego, CA, USA). Tumor size differences were calculated either using Anova or a dedicated software (<https://kroemerlab.shinyapps.io/TumGrowth/>). Briefly, tumor growth was subjected to a linear mixed effect modeling applied to log pre-processed tumor surfaces. P-values were calculated by testing jointly whether both tumor growth slopes and intercepts (on a log scale) were different between treatment groups of interests. Survival probabilities were estimated using the Kaplan-Meier method. Cutoffs for continuous variables were chosen using the median value or an optimal cutoff approach. Survival curves were evaluated using the log-rank test. All reported tests are two-tailed and were considered significant at P-values <0.05.

References:

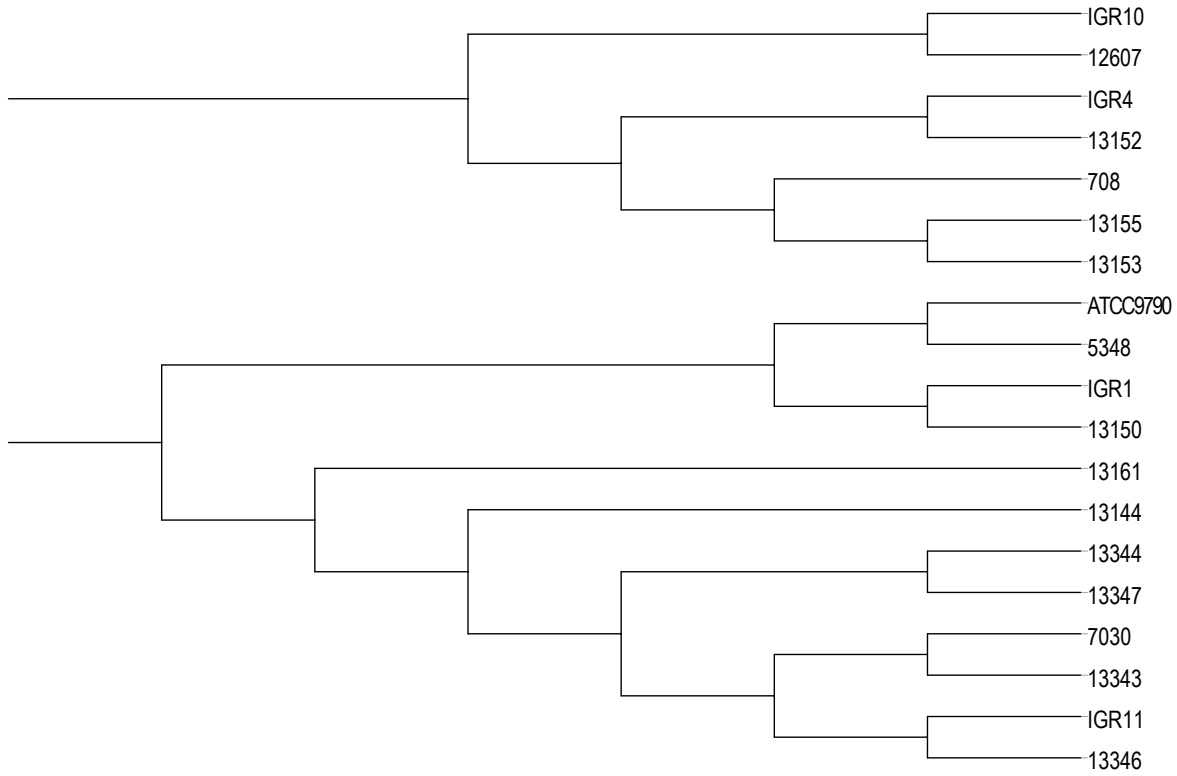
1. M. Bodinier, M. A. Peyrat, C. Tournay, F. Davodeau, F. Romagne, M. Bonneville, F. Lang, Efficient detection and immunomagnetic sorting of specific T cells using multimers of MHC class I and peptide with reduced CD8 binding. *Nat. Med.* **6**, 707–710 (2000).

2. N. Labarriere, A. Fortun, A. Bellec, A. Khammari, B. Dreno, S. Saïagh, F. Lang, A full GMP process to select and amplify epitope-specific T lymphocytes for adoptive immunotherapy of metastatic melanoma. *Clin. Dev. Immunol.* **2013**, 932318 (2013).
3. C. J. Barnstable, W. F. Bodmer, G. Brown, G. Galfre, C. Milstein, A. F. Williams, A. Ziegler, Production of monoclonal antibodies to group A erythrocytes, HLA and other human cell surface antigens-new tools for genetic analysis. *Cell.* **14**, 9–20 (1978).
4. B. Samb-Ba, C. Mazenot, A. Gassama-Sow, G. Dubourg, H. Richet, P. Hugon, J.-C. Lagier, D. Raoult, F. Fenollar, MALDI-TOF identification of the human Gut microbiome in people with and without diarrhea in Senegal. *PLoS One.* **9**, e87419 (2014).
5. T. Sato, R. G. Vries, H. J. Snippert, M. van de Wetering, N. Barker, D. E. Stange, J. H. van Es, A. Abo, P. Kujala, P. J. Peters, H. Clevers, Single Lgr5 stem cells build crypt-villus structures in vitro without a mesenchymal niche. *Nature.* **459**, 262–265 (2009).
6. D. J. LeBlanc, L. N. Lee, A. Abu-Al-Jaibat, Molecular, genetic, and functional analysis of the basic replicon of pVA380-1, a plasmid of oral streptococcal origin. *Plasmid.* **28**, 130–145 (1992).
7. M. J. Kennedy, R. M. Hughes, L. A. Peteya, J. W. Schwartz, M. D. Ehlers, C. L. Tucker, Rapid blue-light-mediated induction of protein interactions in living cells. *Nat. Methods.* **7**, 973–975 (2010).
8. F. A. Ran, P. D. Hsu, J. Wright, V. Agarwala, D. A. Scott, F. Zhang, Genome engineering using the CRISPR-Cas9 system. *Nat. Protoc.* **8**, 2281–2308 (2013).
9. A. M. Bolger, M. Lohse, B. Usadel, Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinforma. Oxf. Engl.* **30**, 2114–2120 (2014).
10. A. Bankevich, S. Nurk, D. Antipov, A. A. Gurevich, M. Dvorkin, A. S. Kulikov, V. M. Lesin, S. I. Nikolenko, S. Pham, A. D. Prjibelski, A. V. Pyshkin, A. V. Sirotkin, N. Vyahhi, G. Tesler, M. A. Alekseyev, P. A. Pevzner, SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J. Comput. Biol. J. Comput. Mol. Cell Biol.* **19**, 455–477 (2012).
11. T. Seemann, Prokka: rapid prokaryotic genome annotation. *Bioinforma. Oxf. Engl.* **30**, 2068–2069 (2014).
12. T. J. Treangen, B. D. Ondov, S. Koren, A. M. Phillippy, The Harvest suite for rapid core-genome alignment and visualization of thousands of intraspecific microbial genomes. *Genome Biol.* **15**, 524 (2014).
13. S. Guindon, J.-F. Dufayard, V. Lefort, M. Anisimova, W. Hordijk, O. Gascuel, New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. *Syst. Biol.* **59**, 307–321 (2010).

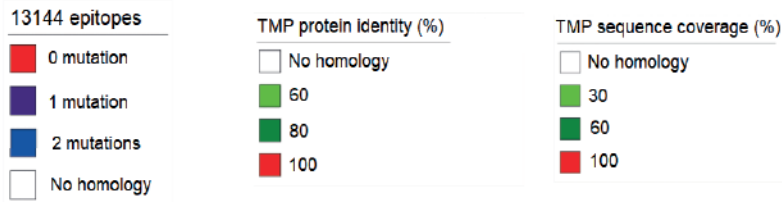
14. L. Li, C. J. Stoeckert, D. S. Roos, OrthoMCL: identification of ortholog groups for eukaryotic genomes. *Genome Res.* **13**, 2178–2189 (2003).
15. D. A. Bolotin, S. Poslavsky, I. Mitrophanov, M. Shugay, I. Z. Mamedov, E. V. Putintseva, D. M. Chudakov, MiXCR: software for comprehensive adaptive immunity profiling. *Nat. Methods.* **12**, 380–381 (2015).
16. B. Routy, E. Le Chatelier, L. Derosa, C. P. M. Duong, M. T. Alou, R. Daillère, A. Fluckiger, M. Messaoudene, C. Rauber, M. P. Roberti, M. Fidelle, C. Flament, V. Poirier-Colame, P. Opolon, C. Klein, K. Iribarren, L. Mondragón, N. Jacquelot, B. Qu, G. Ferrere, C. Clémenson, L. Mezquita, J. R. Masip, C. Naltet, S. Brosseau, C. Kaderbhai, C. Richard, H. Rizvi, F. Levenez, N. Galleron, B. Quinquis, N. Pons, B. Ryffel, V. Minard-Colin, P. Gonin, J.-C. Soria, E. Deutsch, Y. Loriot, F. Ghiringhelli, G. Zalcman, F. Goldwasser, B. Escudier, M. D. Hellmann, A. Eggermont, D. Raoult, L. Albiges, G. Kroemer, L. Zitvogel, Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science.* **359**, 91–97 (2018).
17. S. Simon, Z. Wu, J. Cruard, V. Vignard, A. Fortun, A. Khammari, B. Dreno, F. Lang, S. J. Rulli, N. Labarriere, TCR Analyses of Two Vast and Shared Melanoma Antigen-Specific T Cell Repertoires: Common and Specific Features. *Front. Immunol.* **9**, 1962 (2018).

Figure S1

A



B



Prophage 2 proteins (cutoff identity 60%, coverage 70%)

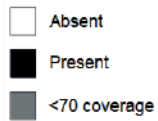


Figure S1. Clading and comparative analysis of *E. hirae* 13144 39.2kb-prophage protein sequence with other *E. hirae* strains.

A. Phylogenetic tree of 19 *E.hirae* genomes based on SNP alignments.

B. A particular genomic trait of *E. hirae* 13144 is that it encodes two intact prophage regions (of 40.6 kb and 39.2 kb) showing weak sequence identities with the most common *Enterococcus* phage phiEf11 *vB_EfaS_IME197* (14% and 11% of shared genes, respectively) (Table S3). Comparative analysis of the 39.2kb prophage of *E. hirae* 13144 with 18 other sequenced *E. hirae* genomes showed that this phage was strain-specific, although portions of its genome were detectable in other *E.hirae* strains. Comparative analysis through a “heatmap” clustering based on a matrix of presence (black) and absence (white) of the *E.hirae* 13144 39.2kb-prophage protein sequence or the TMP1 epitope and HLA-A2 TMP epitope 10 without mutation (red), with 1 mutation (violet) or 2 mutations (blue).

Figure S2

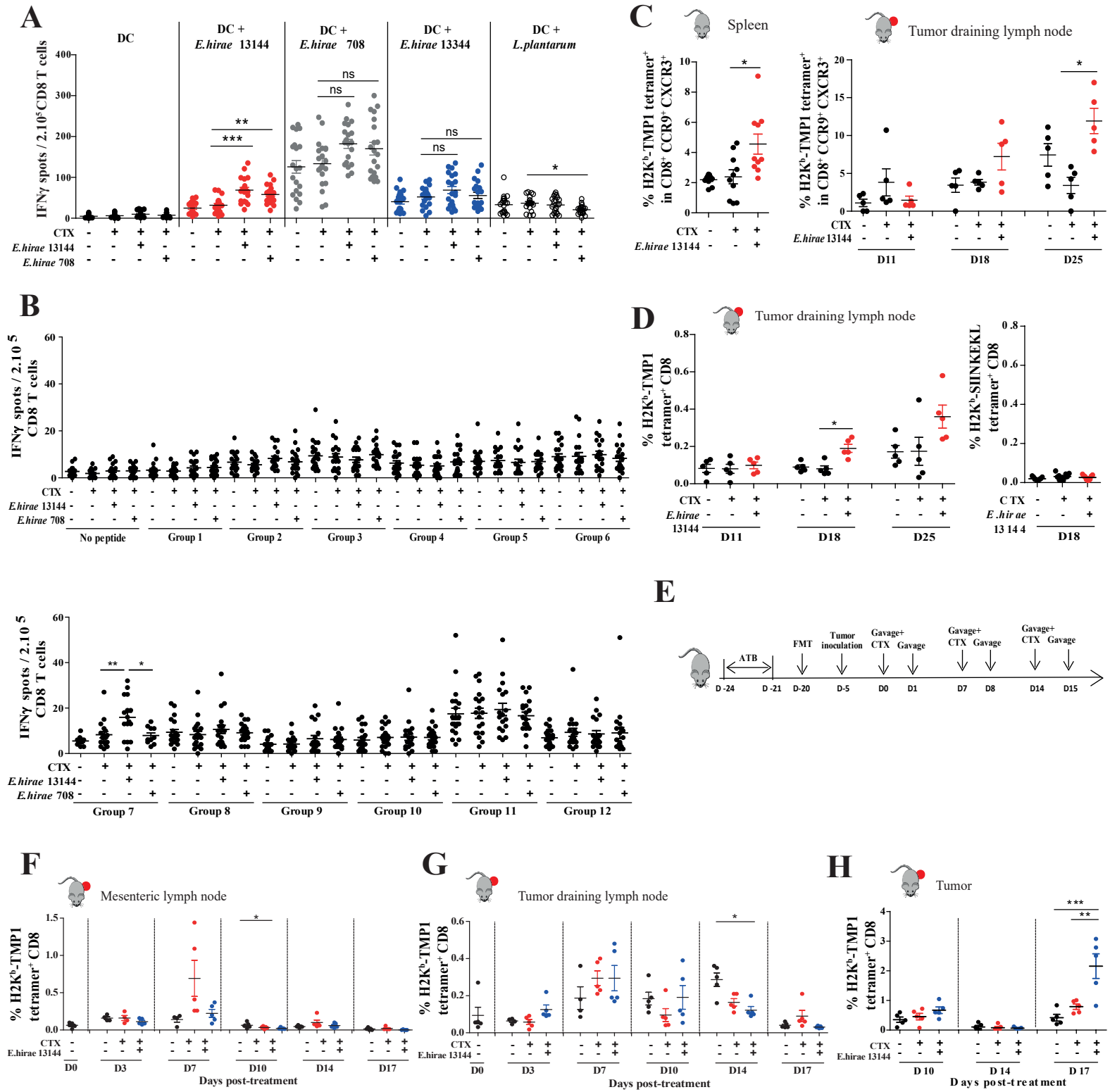


Figure S2. Identification of group 7 as the only group of peptides containing an immunogenic one.

A-B. Naive mice were treated with broad spectrum antibiotics (streptomycin, colistin, ampicillin, vancomycin) for 3 days before oral gavage with *E. hirae* strain 13144 or 708 (1×10^9 bacteria) was performed prior to and after systemic administration of cyclophosphamide (ip CTX - 100 mg/kg) or saline solution (NaCl) at day 4 once (like in Fig. 1C). One week later, purified CD8⁺ T splenocytes were restimulated *ex vivo* in a recall assay with bone marrow-derived DC loaded with saline or distinct heat killed (65°C during 2 hours) bacterial strains (A) or groups of peptides (Table S2) (B). IFN γ -secreting CD8⁺T cells (spots) were determined after 24h of coculture. Each dot represents one mouse. The experiment (with 5 mice/group) was performed three times. Statistical analyses revealed that only group 7 of peptides induced a significant response (B). C. Flow cytometric determination of H-2K^b/TSLARFANI tetramer-binding CD8⁺ T cells in the spleen of naive mice (left panel) and tumor draining lymph node (right panel) of tumor bearers in the gate of CXCR3⁺CCR9⁺ double positive cells. D. Flow cytometry analyses of H-2K^b/TSLARFANI (left panel) or H-2K^b/SIINFEKL (right panel) tetramer binding CTL in tumor draining lymph nodes at various time points in tumor bearing mice treated with CTX and *E. hirae* 13144 (like in Fig. 1A). One representative experiment out of two yielding similar conclusions is shown. Each dot represents one animal, each experiment containing 5-6 mice/group. Data from two independent experiments are depicted. E-H. Avatar mice are SPF C57BL/6 animals treated with 3 days of ATB to allow establishment of a fecal microbial transplant (FMT from a breast cancer patient) 21 days prior to CTX+*E. hirae* 13144 therapy (E). Kinetic study of H-2K^b/TSLARFANI tetramer-binding CD8⁺ T cells by flow cytometry analyses in various organs (mesenteric lymph nodes (mLN) (F), tumor draining lymph nodes (tdLN) (G) and tumor beds (MCA205) (H) during a therapy of established MCA205 with the combination of CTX+*E. hirae* 13144 in human fecal material (FMT) subjected avatar mice according to the experimental setting detailed in E. Each dot represents one animal, each experiment containing 5-6 mice/group. Data from one representative experiment are depicted. ANOVA statistical analyses (Kruskal-wallis test): * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. Refer to the statistical report.

Figure S3

TMP 39.2-kb prophage (1506 aa)

> *E.hirae*_13144_02029

TMP epitope 10 (HLA-A0201)

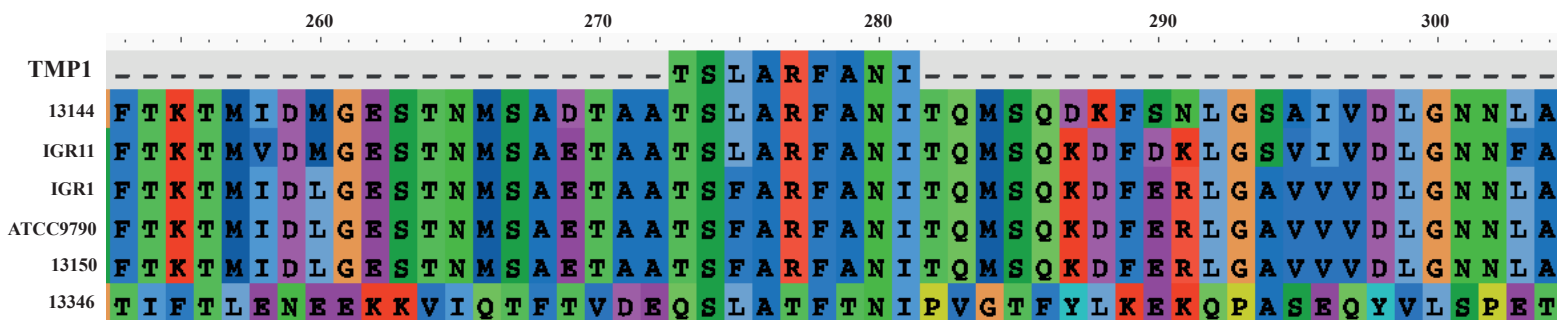
TMP1 (H2-kB)

MAQSKTVKAVLTAIDKGFQTMTGSAATSSLKKLSSNASDIPSNLNTVSGAMKSFSGDKTASIGQSIEKVGGSMTKGITLPIAGAVGAVTTAAVKWESA
FTGVKKTNDDEMVDVSDNGKVIYSYDDLEKGLRDLAKELPTSHEEIAKVAEAAAGQLGIKTDKVVGFCTKTMIDMGESTNMSADTAA**TSLARFANI**ITQMS
QDKFSNLGSAIVDLGNNLATTESIEITMGLRLAGAGKQIGMTEGDIVGFAAALSSVGI EAEAGGS AFSRLMVQMQLATETGVKAFEP LKQAVAIQ
GVSWEFVHAVNWWGGKELTAVSKQMGVPASELKKLYKEASKASGSLEDFANVTGRTGEFEAE LFKSNPSQAMIEFIQGLKDEKHGISAIVLDD
MGITEVRLRDSLLRAANASDVFE GAVKRGNEAF NENTALAE EAGKRYGTTESQLKILRGQLNDVAITFGGPLVAALNSAISAAKPMIEALANMAEA
FASADPKTQEFILKMAALAASAGPVLKVF GKMTSVFGKTISTMFEKAGNIDSKWKQFIVTPIKNGSSSALQAVKGFVSKYKSNLAGLESAGINVNLL
TRFTTLKDTIVGLFPTLDTFGANLRASQRQLNMLGEGNKVTNFRSFSASLQLSNS **KLAKFASVV**INPIGSLRNLSSAAGKSGTVLSGLGVAASKAGG
GFRTFAATGIRSIASLTGAMLSNPITAILVAITTTIVGVVQAWKSNFMNIQGYVKTAFSGIVKSFKSVLPSSASVTKTIKGLGNIFKWLGTGTLVGVTFA
IAGFVDGLRAIITVGKTAVNAIMAIANGVKGLWQRLKGDSKGDADKSKFDVKKSLADIGKDWDTMFSDSALKKAAKSTEELGKKSDDTTKAMSMN
MEEVSNVENYSSKLEAKQAMTELSQQNGSTAGVEAYFNHTLDLVTNLKEQQKKA VETYNKQIEAAEGKSEA EKQKIFANASTEYMKAVQSN
NSDLLKVYTDYSNQLKNNKTVEGQELTDQQRATLQ NQTNIIRDQLDQKQFVEAGV NKLNNQALSEQEKEQLTSSLKTFGEIQAAQQVQENNA
QIQQLTQKNQAKTES EKAAFQ NQITQLQTQNDQIRQSELEQGAQLLAIISQNGANKIAVTADNLAQLKG VTDQQLLGIYQSYVNNGASIDQQM
ALLAGMLRQRGIDGSNGLVQGLQSNDPKLWANMSKADIVNTLQSLPPDLFKNGQDGK NKLIDGLNSGKVEINN VGOELMNQMN SGVKNKKA
EAEKTSGDVASSGAKGAKSKGKEYNSGGNSNAGEYNTGLAKQKSNKQKGAELGSAPVEGVKTKASAMRSVGEQLGRSFVQGLASQVGSANNA
GRELGNAVKSGAGSVNMTSVGSNMAKGVASGIRASQGEAVSAMQNLVAAVNAEAQKKAKIKSPSRLLYDVG VFLAQGVAA GIREDTSVAVQS
AKDMISSIHQSITGSRLIKRSNAIEVKHSIDNTPMGKMEILEEIRH LTVM DTGQVVGALGSPMNLNLAEQQKQDGRYRS

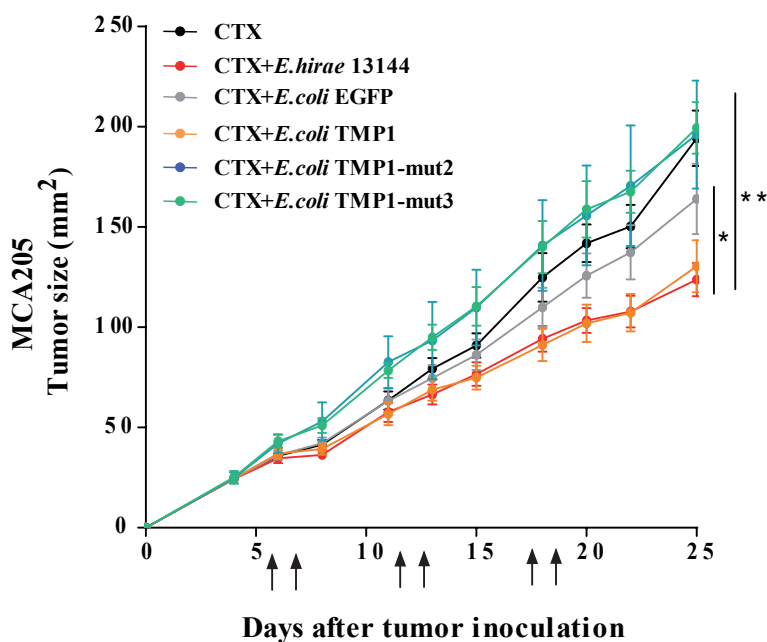
Figure S3. *E. hirae* 13144 39.2kb-prophage sequence alignment with location of the TMP protein and TMP epitopes. The position of the H-2K^b-restricted TMP1 epitope and HLA-A2*0201 TMP epitope 10 are indicated in red and green, respectively.

Figure S4

A



B



C



Figure S4. Sequence alignment of the immunogenic epitope region within 39.2kb-prophage of *E. hirae* 13144. A. The immunogenic peptide TMP1 (TSLARFANI, A) from *E. hirae* 13144 identified in Figure 1 and Figure S2 is aligned to sequences from other *E.hirae* strains tested in this study. B. Complete tumor growth curves of the experiment shown in Fig. 2D. C57BL/6 mice bearing MCA205 sarcomas were treated with CTX and gavaged with *E. hirae* 13144 or *E.coli* genetically modified to express TMP1 (TSLARFANI), TMP1 mut2 (TALARFANI), TMP1 mut3 (TSFARFANI) or EGFP sequence (as control). The means \pm SEM of tumor sizes at different time points for 12-18 animals, gathered from 2-3 independent experiments are shown. C. The immunogenic peptide HLA-A*0201 TMP epitope 10 (KLAKFASVV) from *E. hirae* 13144 identified is aligned to the sequences from other *E.hirae* strains.

Figure S5

A

TMP-FLAG = TSLARFANI

MAQSKTVKAVLTAIDKGFTQTMGSATSSLKKLSSNASDIPSNLNTVSGAMKSFSGDKTASIGQSIEKVGGSMTKGITLPIAGAVGAVTTAA
VKWESAFTGVKKTNDDEMVDNSNGKVIYSYDDLEKGLRDLAKELPTSHEEIAKVAEAAGQLGIKTDKVVGFCTMIDMGESTNMSADTAA
TSLARFANITQMSQDKFSNLGSAIVDLGNNLATTESEITEMGLRLAGAGKQIGMTEGDIVGFAAALSSVGIEAEAGGSAFSRLMVQMQ
LATETGVKAFEPLKQAVAIQGVSWEKVHAVNWGGKELTAVSKQMGVPASELKKLYKEASKASGSLEDFANVTGRTGEEFAELFKSNP
SQAMIEFIQGLKDSEKHGISAIVLDMGITEVRLRDSLLR**DYKDDDDK**

TMP-mut2-FLAG (mutation in position 2) = TSLARFANI to TALARFANI

MAQSKTVKAVLTAIDKGFTQTMGSATSSLKKLSSNASDIPSNLNTVSGAMKSFSGDKTASIGQSIEKVGGSMTKGITLPIAGAVGAVTTAA
VKWESAFTGVKKTNDDEMVDNSNGKVIYSYDDLEKGLRDLAKELPTSHEEIAKVAEAAGQLGIKTDKVVGFCTMIDMGESTNMSADTAA
TALARFANITQMSQDKFSNLGSAIVDLGNNLATTESEITEMGLRLAGAGKQIGMTEGDIVGFAAALSSVGIEAEAGGSAFSRLMVQMQ
LATETGVKAFEPLKQAVAIQGVSWEKVHAVNWGGKELTAVSKQMGVPASELKKLYKEASKASGSLEDFANVTGRTGEEFAELFKSNP
SQAMIEFIQGLKDSEKHGISAIVLDMGITEVRLRDSLLR**DYKDDDDK**

TMP-mut3-FLAG (mutation in position 3) = TSLARFANI to TSFARFANI

MAQSKTVKAVLTAIDKGFTQTMGSATSSLKKLSSNASDIPSNLNTVSGAMKSFSGDKTASIGQSIEKVGGSMTKGITLPIAGAVGAVTTAA
VKWESAFTGVKKTNDDEMVDNSNGKVIYSYDDLEKGLRDLAKELPTSHEEIAKVAEAAGQLGIKTDKVVGFCTMIDMGESTNMSADTAA
TSFARFANITQMSQDKFSNLGSAIVDLGNNLATTESEITEMGLRLAGAGKQIGMTEGDIVGFAAALSSVGIEAEAGGSAFSRLMVQMQ
LATETGVKAFEPLKQAVAIQGVSWEKVHAVNWGGKELTAVSKQMGVPASELKKLYKEASKASGSLEDFANVTGRTGEEFAELFKSNP
SQAMIEFIQGLKDSEKHGISAIVLDMGITEVRLRDSLLR**DYKDDDDK**

B

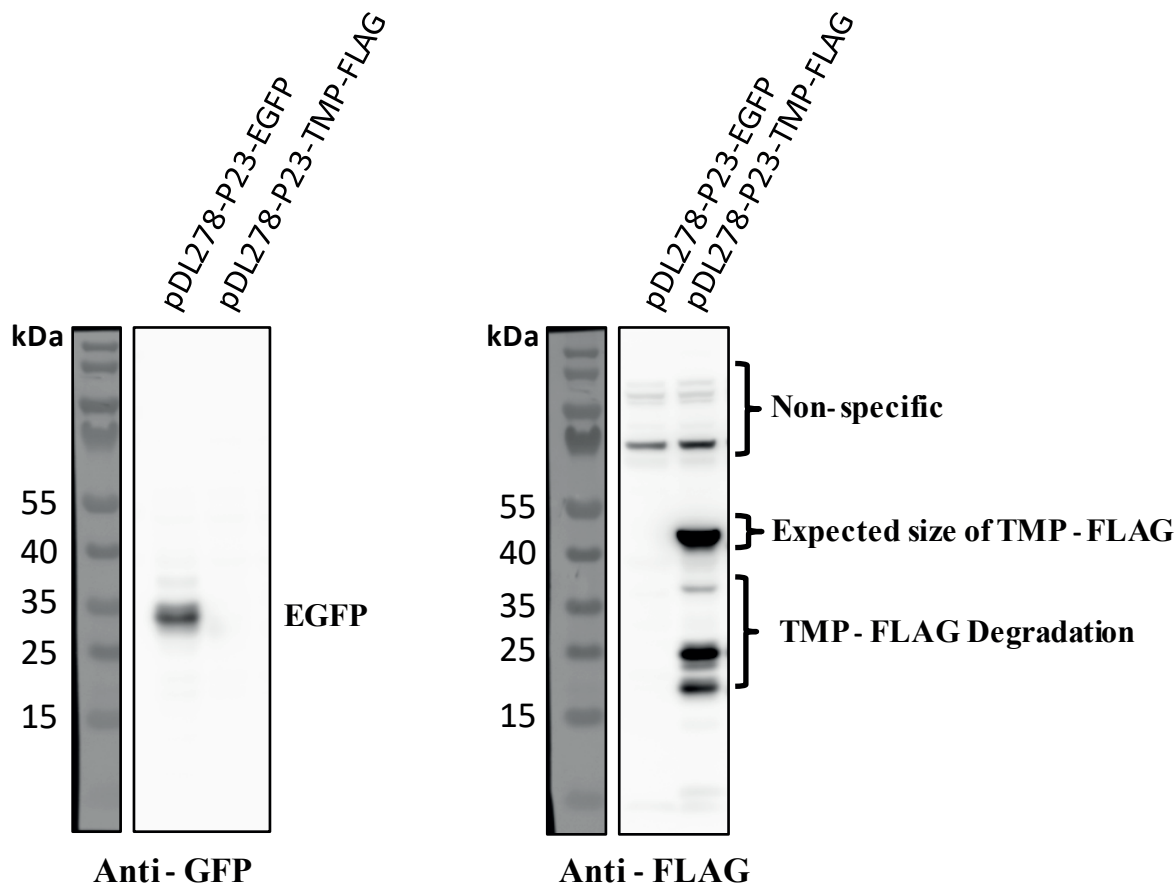


Figure S5. Sub-cloning expression of part of the TMP gene in *E.coli*.

A. Amino acid sequences of TMP-FLAG, TMP-mut2-FLAG and TMP-mut3-FLAG expressed in *E. coli* DH5 α . Note that only the N-terminal part of the TMP protein, including the indicated variants of the epitope (green), was expressed as fusion protein with a C-terminal FLAG tag (blue). B. Western blot analysis demonstrating expression of EGFP and TMP-FLAG in *E. coli* strains transformed with pDL28-P23-EGFP or pDL28-P23-TMP-FLAG, respectively.

Figure S6

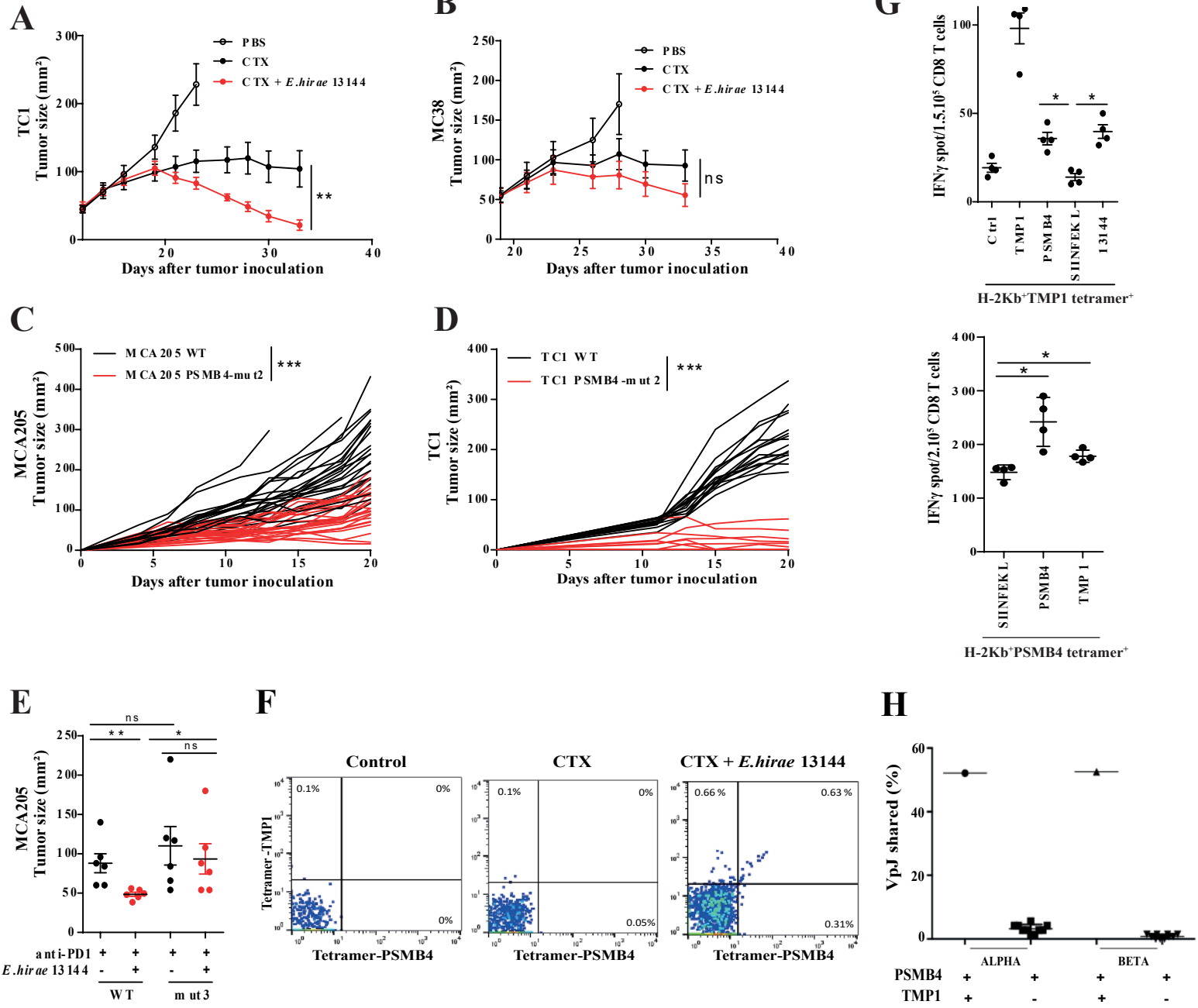


Figure S6. Molecular mimicry between the TMP1 phage and the PSMB4 oncogenic driver.

A-B. Tumor growth of TC1 (A) and MC38 (B) cancers with or without therapy combining CTX \pm *E. hirae* 13144 (or saline) following the experimental setting described in Fig1A. C-D. Tumor growth of WT clones or clone harboring a knock-in mutation in position 2 (mut2) of GSLARFRNI for MCA205 (C) and TC1 (D). Each line represents one animal. Two concatenated experiments comprising each 6 mice/group are depicted. E. Tumor sizes at day 9 of SPF mice implanted with WT or mut3 MCA205 clones treated every 3 days three times with anti-PD1 \pm *E. hirae* 13144. F. Flow cytometric determination of splenic T cells co-staining with two different tetramers (TMP1 related H-2K^b/TSLARFANI or PSMB4-related GSLARFRNI/H-2K^b complexes). Representative dot plot of CD3⁺CD8⁺ splenic T lymphocytes staining with either or both tetramers in one representative tumor bearing animal treated with PBS, or CTX or CTX+*E.hirae* 13144. G. Cells were prepared following a protocol of *in vitro* expansion (detailed in Figure 4B) in which BM-DC were pulsed with TMP1 or PSMB4 peptide. The number of IFN γ secreting CD8⁺ GSLARFRNI/H-2K^b (reexpanded after *in vitro* stimulation with PSMB4 peptides, and apostrophed "CD8⁺PSMB4⁺" on the graph) after stimulation with BM-DC pulsed with the three different peptides, and the number of IFN γ secreting CD8⁺ H-2K^b/TSLARFANI⁺ (reexpanded after *in vitro* stimulation with TMP1 peptides and apostrophed "CD8⁺TMP1⁺" on the graph) after stimulation with BM-DC pulsed with the three different peptides are both depicted in two representative experiments (lower and upper graph respectively). Of note, the binding affinity for H2-K^b of GSLARFRNI peptide is lower than that of TSLARFANI (EC50 :216.85 versus 7.81 nM respectively). We did not observe any IFN γ secretion by the negative fraction CD8⁺ H-2K^b/TSLARFANI⁻ cells. H. TCR α/β sequences shared between PSMB4-specific TCRs and TCRs from CD8⁺ T cells that bind H-2K^bTMP1-tetramers or fail to do so. Each point represents the result of a random sampling of the same number of clonotypes as for the comparison with TMP1-specific TCRs. Mann Whitney test or ANOVA statistical analyses (Kruskal-wallis test): * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. Refer to the statistical report.

Figure S7

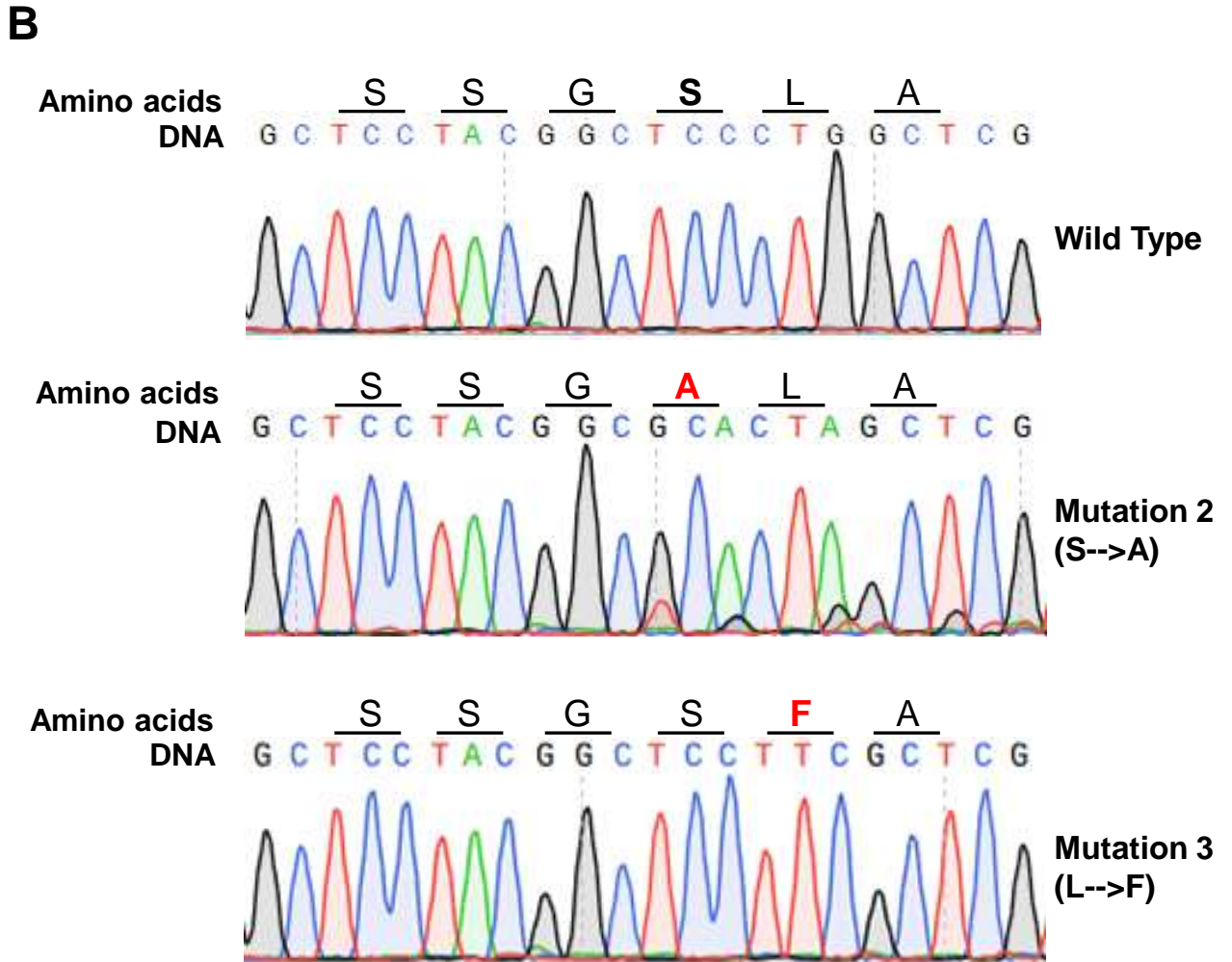
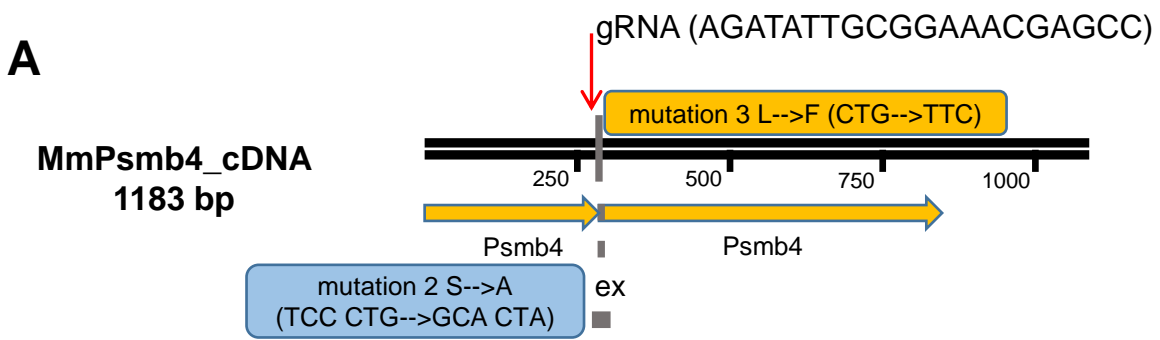
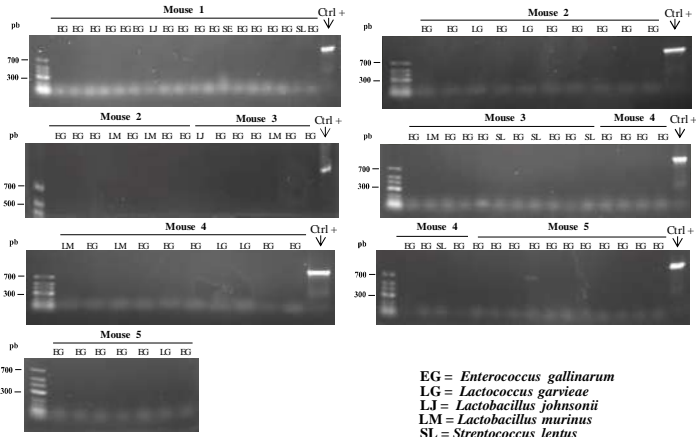


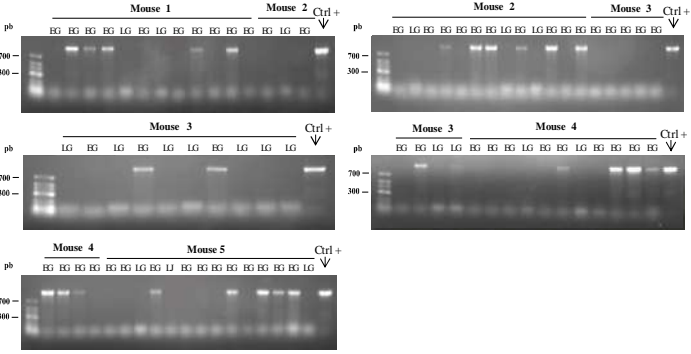
Figure S7. Generation of *pmsb4*-mutated MCA205 cell lines by means of the CRISPR/Cas9 technology. A. Schematic diagrams of *Psmb4* cDNA, and the designed mutation sites. The target site of sgRNA and point mutations are indicated. B. Representative sequence electropherograms for the validation of *Pmsb4* mutation 2 and mutation 3 introduced by CRISPR/Cas9. Mutated amino acids are highlighted in red. Similar methods were used for engineering TC1 cells.

Figure S8

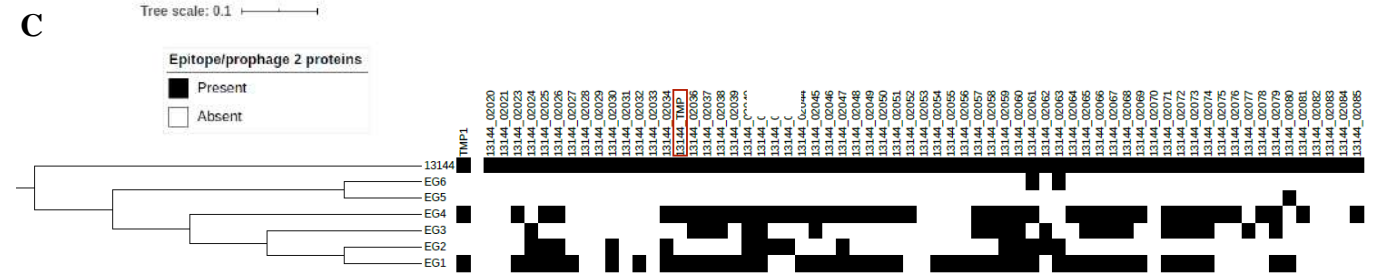
A Naïve mice



B + *E.hirae* 13144 mice



C



D

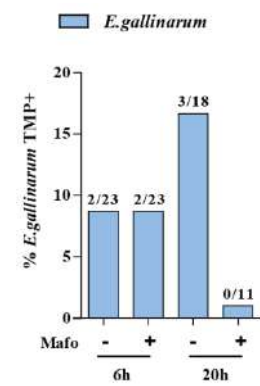
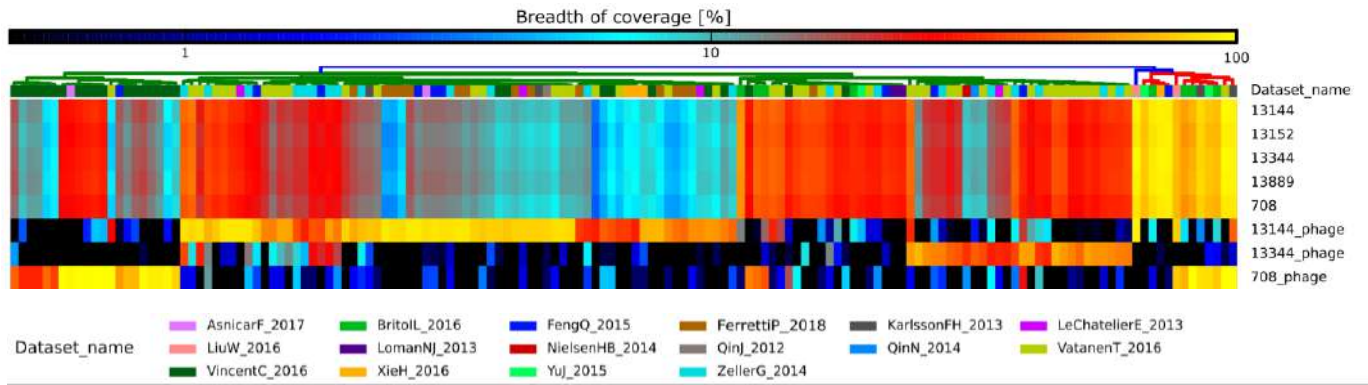


Figure S8. Identification of ileal bacterial colonies after treatment with CTX+oral gavage with *E. hirae* 13144.

A-B. PCR amplification of the TMP sequence in each colony growing after seeding of ileal content in aerobic conditions to isolate Gram⁺ bacteria. A photograph of each agarose electrophoresis gel is shown for each animal. A depicts the results in 5 naive mice (A) and B depicts the findings after CTX+oral gavage with the phage encoding bacterium *E. hirae* 13144 (10⁹ cfu) (B). Each vertical lane corresponds to the bacterium identified in MALDI-TOF. Initials are detailed in the lower part of panel A. The positive control (Ctl+) represents the DNA of *E. hirae* 13144. C. Sequence alignment of the prophage harbored by *E. hirae* 13144 in 6 strains of *E. gallinarum* (EG1 to EG6) harvested from ileal material after oral gavage of naive mice with *E. hirae* 13144 and therapy with CTX. Comparative analysis through a “heatmap” clustering based on a matrix of presence (black) and absence (white) of the *E. hirae* 13144 39.2kb-prophage protein sequence or the TMP1 epitope and TMP protein. D. Small intestine stem cell crypt derived-organoids were incubated with *E. hirae* 13144 and *E. gallinarum* at a 1:1 ratio for 6 or 20 hrs ± the CTX derivative mafosfamide. Then, live colonies of *E. gallinarum* were harvested and analyzed by PCR. The percentages of *E. gallinarum* which turned positive for TMP detection are indicated in a representative experiment out of two yielding similar results.

Figure S9

A



B

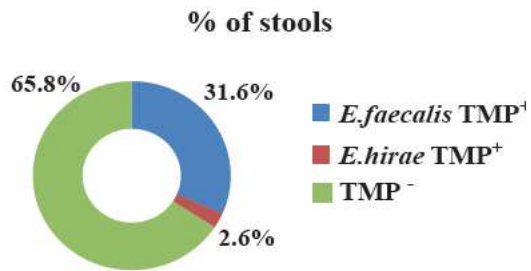


Figure S9. Breadth of coverage of the *E. hirae* genome and its phage in the MG reference catalog and culturomics analyses of patients stools.

A. Breadth of coverage (BOC) of different *E. hirae* sequences (*E. hirae* 13144, 708, ATCC9790) and its phage in 3,027 adult and mother-infant metagenomes (mostly from human stools but also from various mucosae) by reference-based mapping of metagenomic reads from 17 publicly available datasets annotated in curatedMetagenomicData. The BOC measures the fraction of the genome that is covered by the reads in the metagenomes. The color code is indicated with highest BOC towards yellow/red colors and no BOC in black. We first screened a total of 3,027 adult and mother-infant metagenomes (mostly from human stools but also from other mucosae) by reference-based mapping of metagenomic reads from 17 publicly available datasets annotated in curatedMetagenomicData to assess the breadth of coverage (BOC) of the *E. hirae* genome and its phages. *E. hirae* was present with 100% confidence (i.e. BOC > 80%) in 13 samples from disparate geography, age and datasets. In another ~40 cases, the presence of *E. hirae* was very likely but could not be confirmed with high confidence because of insufficient sequencing depth. In 70% of the samples in which *E. hirae* was confidently found, one of the three phage sequences (from *E. hirae* 13144, 708 or 13344) were also detected, though in a partially mutually exclusive fashion (Figure S9A). Of note, the *E. hirae* 13144 phage was detectable in many samples lacking the presence of the *E. hirae* core genome, suggesting that other bacteria than *E. hirae* can host this phage. Analysis of the global prevalence of these phages (irrespective of the presence of *E. hirae*) further confirmed their mutually co-exclusion in the microbiome. We could detect the presence of phage 13144 at 0.66 BOC in three mother-infant paired stool specimens and in the infants at 1, 3, and 7 days after birth suggesting that this phage (and its host) can be vertically transmitted from mothers to infants and then colonize the neonate. We complemented this analysis by a metagenomic-assembly based screening of 9,428 metagenomes, confirming the presence of phage 13144 in humans across the world at a low prevalence (272 positive samples), though possibly with a overrepresentation among a non-Westernized population from Madagascar (19 positive samples). Importantly, this analysis highlighted an increased prevalence of the phage (57%) in fecal microbiomes from children (representing 16% of all metagenomes, Fisher's test p-value <0.00001). We confirmed the integration of the phage into the genome of distinct bacterial species for 128 positive samples (47%), when re-evaluating 154,723 microbial genomes reconstructed from the same 9,428 samples. All host genomes belonged to the

Enterococcus genus (except two assigned to *Coprobacillus*), in particular *E. faecalis* (80 genomes), *E. faecium* (23 genomes), and *E. hirae* (15 genomes), suggesting that phage 13144 (and its homologues from *E. hirae* 708, and 13344) are genus-specific but not species-specific. B. Percentages of stools with detectable TMP sequences in *E. hirae* and/or *E. faecalis* colonies assessed by culturomics followed by PCR in 76 NSCLC and RCC bearing patients (cohort described in (16)), the corresponding Kaplan Meier curves indicating overall survival featuring in Figure S11E-F. Up to 5 colonies per species and individual (either colonies from *E. avium*, *E. casseliflavus*, *E. durans*, *E. faecium*, *E. faecalis*, *E. gallinarum*, *E. hirae*) from 76 cancer patients led to the detection of the TMP sequence encompassing the TMP1 peptide (aligned in Figure S10) in 34% of the patients. PCR detected TMP sequences only in *E. faecalis*, with 29 colonies positive out of 118 colonies that were tested (not in *E. faecium* nor *E. durans*).

Figure S10

TCAACTACGATACCTCCCATCTTGCTTTTGTGTTCTGCTAAATTAAGATTCATCGGGCTACCTAGTGCTCCCCTACTTGCCAGTATCC
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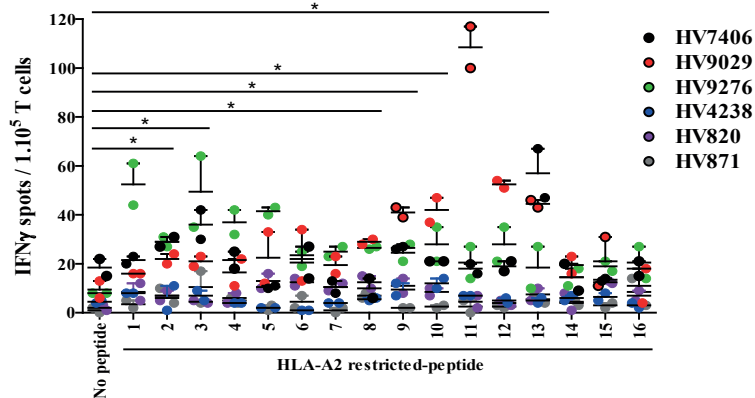
Primers

Sequence of TMP1

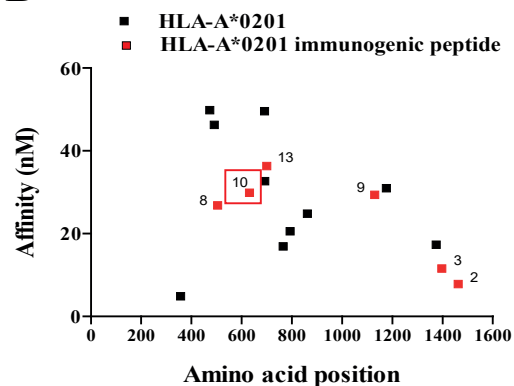
Figure S10. Sequence of the Phage Tail Length Tape Measure Protein in *E. hirae*.
Nucleotide sequence of the whole TMP protein as well as binding area for PCR primers indicated in green and TMP1 epitope sequence indicated in red.

Figure S11

A



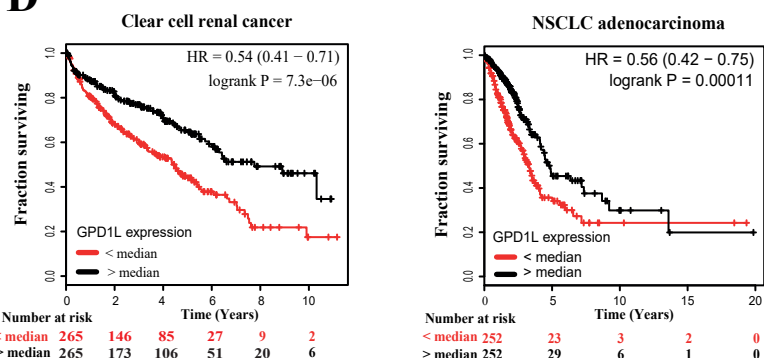
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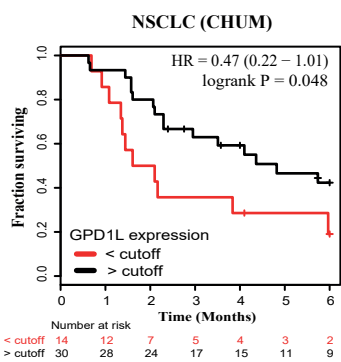
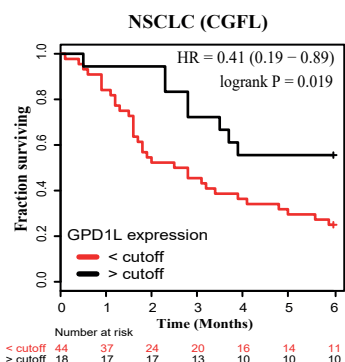
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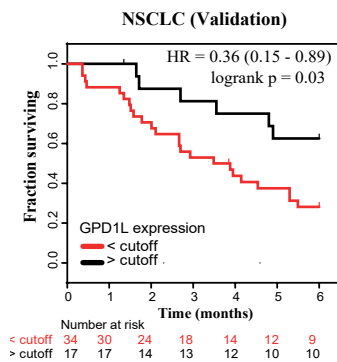
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G

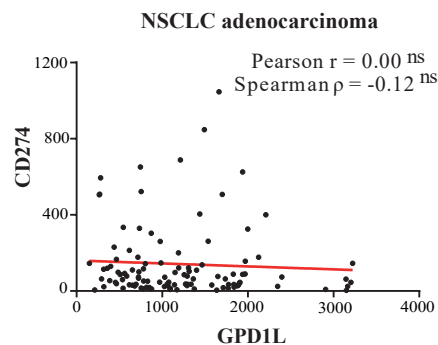


Figure S11. Identification and functional impact of TMP-crossreactive epitopes in the GPD1-L protein .

A. Priming of naive CD8⁺ T cells from six HLA-A*0201 healthy volunteers with autologous monocyte-derived DC pulsed (or not) with 16 HLA-A*0201 binding TMP epitopes (Table S6). Restimulation at day 7 with each of the 16 TMP peptides for IFN γ ELISpot assays and enumeration of positive spots. ANOVA statistical analyses: $*p < 0.05$. B. The HLA-A*0201 binding and immunogenic epitopes (Table S6) are located in defined domains of the TMP protein, as indicated by the color code (red: 6 peptides with significant reactivity in A) and the amino acid sequence position, as a function of their binding affinity to the MHC class I allele (calculated *in silico*). C. Blast sequence alignment of the immunogenic HLA-A*0201-restricted TMP epitope 10 (KLAKFASVV) with a sequence belonging to the GPD1-L protein (KLQKFASTV). D. Impact of GPD1-L mRNA expression on survival in 530 clear renal cell cancers (left panel) and lung adenocarcinoma (right panel) from the TCGA. Patients were segregated according to the median value of GPD1-L expression, and Kaplan Meier curves of overall survival were compared by Cox regression univariate analysis. E-F. Kaplan Meier curves for time to progression following PD-1 blockade in second line therapy in 44 stage IIIC/IV NSCLC patients (CHUM test cohort, E) corroborated with a second cohort of 62 stage IIIC/IV NSCLC patients (CGFL cohort, E) and then a validation cohort of stage IIIC/IV NSCLC patients (F, n=51) using an optimal cut-off value for GPD1-L tumor expression obtained in RNA sequencing for each cohort (Table S7 for patients description). G. Absence of correlations between GPD1-L and CD274/PD-L1 mRNA expression in lung cancers (TCGA, CHUM and CGFL cohorts together), as determined by Spearman and Pearson calculations.

Figure S12

A GPD1L

>sp|Q8N335|GPD1L_HUMAN Glycerol-3-phosphate dehydrogenase 1-like protein

MAAAPLKVCIVGSGNWGSAVAKIIGNNVKKLQKFASTVKMWVFEETVNGRKLTDIINNDHENVKYLPGHKL
PENVVAMSNLSEAVQDADLLVFPVPHQFIHRICDEITGRVPKALGITLIKIGIDEGPEGLKLISDIIREKMGIDISVL
MGANIANEVAAEKFCETTIGSKVMENGLLFKELLQTPNFRITVVDADTVELCGALKNIVAVGAGFCDGLRCG
DNTKAAVIRLGLMEMIAFARIFCKGQVSTATFLESCGVADLITTCYGGRNRRVAEAFARTGKTIEELEKEMLNQ
KLQGPQTSAEVYRILKQKGLLDKFLFTAVYQICYESRPVQEMLSCLQSHPEHT

³⁰KLQKFASTV³⁸

B

Gene Name	Sample Name	AA Mutation	Primary Tissue
GPD1L	TCGA-DZ-6132-01	K30T	Kidney
GPD1L	TCGA-17-Z053-01	L31F	Lung
GPD1L	U343	K33N	Central nervous system

C

Gene Name	Sample Name	AA Mutation	Primary Tissue
GPD1L	TCGA-AA-3667-01	K29N	Large intestine
GPD1L	TCGA-KN-8431-01	K39_W41del	Kidney
GPD1L	TCGA-KN-8431-01	K39_V42delinsl	Kidney
GPD1L	T593	M40V	Large intestine
GPD1L	EGC3	W41R	Stomach

D

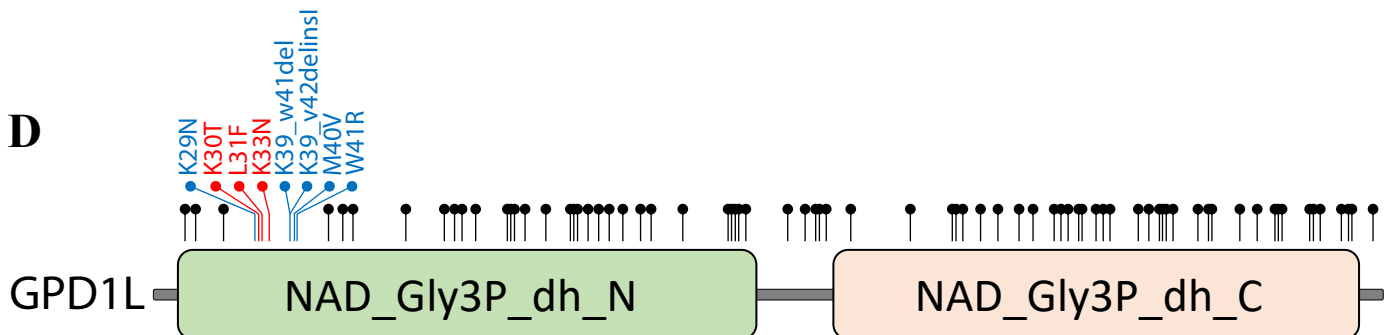
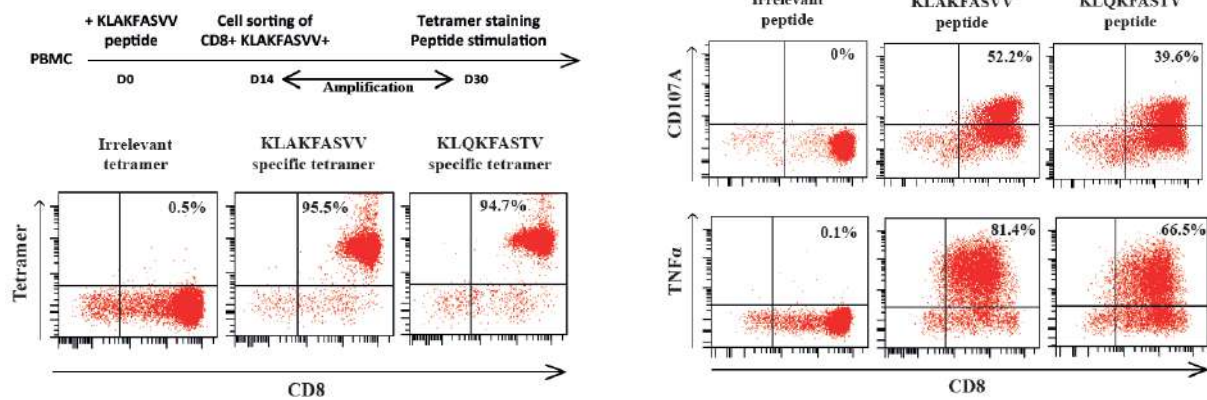


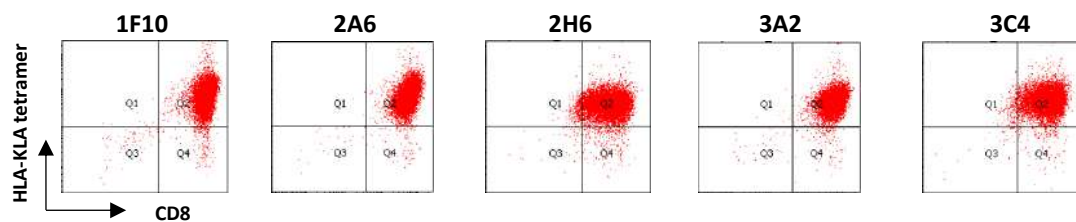
Figure S12. Cancer-associated mutations in GPD1-L from cBIOPORTAL and COSMIC. A. Protein sequence of GPD1-L. B. Mutations annotated in the conserved sequence KLQKFASTV (highlighted in red). C. Mutations annotated in positions adjacent to the conserved sequence KLQKFASTV (highlighted in blue). Gray background indicates two mutations found in the same sample. D. Distribution of all cancer-associated GPD1-L mutations. Mutations in the conserved sequence KLQKFASTV are highlighted in red and adjacent mutations are highlighted in blue.

Figure S13

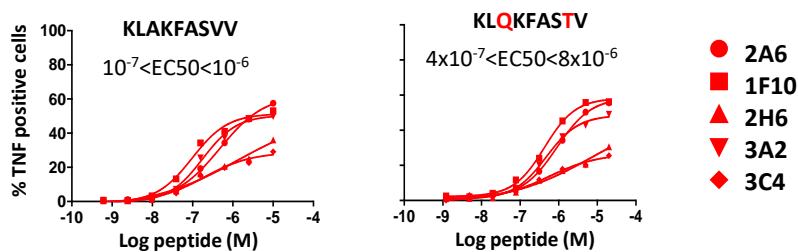
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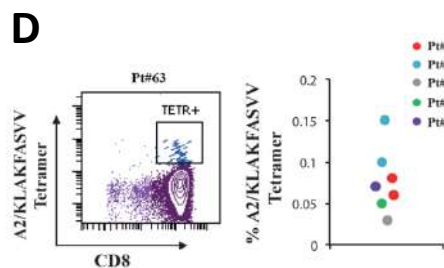
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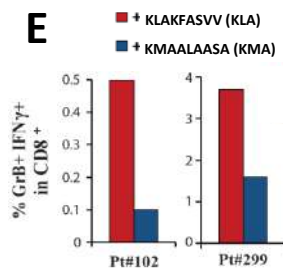
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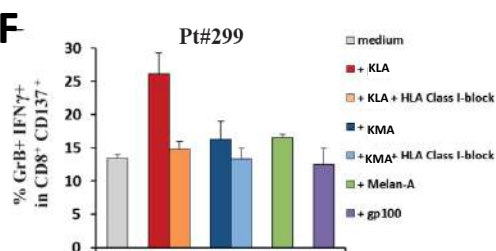
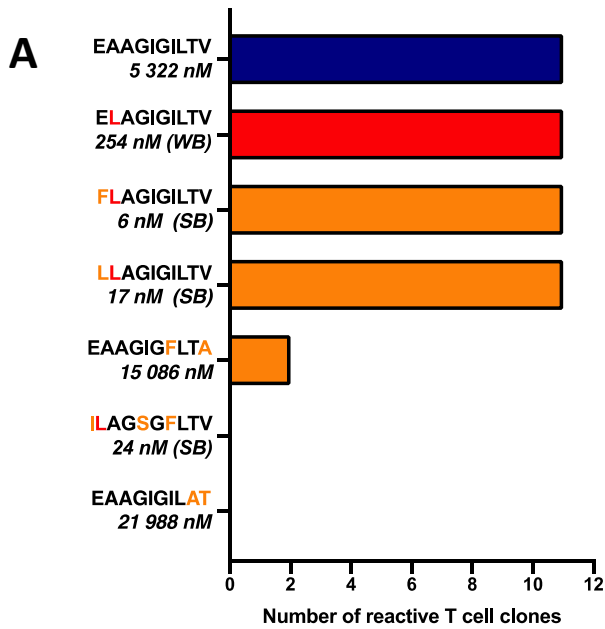


Figure S13. Crossreactive T cells recognizing HLA-A*0201-restricted peptides from TMP and GPD1-L proteins.

A-C. *In vitro* stimulation of T cells from normal volunteers. A. Experimental setting (upper left panel). HLA-A*0201⁺ PBMC extracted from a healthy volunteer were stimulated in microculture assays for 14 days, *ex vivo*, with KLAKFASVV peptide. Each well was screened for costaining with both tetramers (HLA-A*0201/KLAKFASVV from TMP and HLA-A*0201 /KLQKFASTV from GPD1-L). One well out of 192 tested was positive and subjected to cell sorting using HLA-A*0201/KLAKFASVV multimer-coated beads and re-expanded for 15 days. The resulting T cell line was further characterized by flow cytometric analyses for binding to each and both tetramers (A, lower panels) and for its functional cross-reactivity with both epitopes (KLAKFASVV from TMP and KLQKFASTV from GPD1-L), as measured by degranulation (CD107A surface expression) and TNF α release assays. One representative dot plot is depicted for each experimental condition of stimulation (A, upper right panels). B. After cloning of the T cell line by limiting dilution assays, we studied the five KLAKFASVV-specific CD8⁺ CTL clones (1F10, 2A6, 2H6, 3A2 and 3C4) for their capacity to bind the HLA-A*0201/KLAKFASVV tetramer (B) and to secrete TNF α after exposure to increasing concentrations of the two peptides. Three clones exhibited lower or similar affinity for KLQKFASTV (KLQ) compared with KLAKFASVV (KLA) peptides (C). D-F. *In vitro* stimulation of T cells from NSCLC patients. Representative flow cytometry dot plot analyses of PBMC from one of the five HLA-A*0201 NSCLC patients tested, after short term *ex vivo* restimulation with KLAKFASVV peptide followed by tetramer staining using HLA-A*0201/KLAKFASVV tetramers (D, left). Results from 5 different NSCLC patients (D, right panel), each dot representing the percentages of tetramer⁺CD8⁺ T cells for each patient evaluated. Dots with a similar color represent values from two independent experiments performed using the same patient's PBMC. E-F. Percentages of co-production of GrB and IFN- γ , as measured by multicolor intracellular staining, in CD8⁺ T-cell lines obtained from three HLA-A*0201 NSCLC patients (out of 6 tested) after priming with TMP epitope 10 (KLAKFASVV) and final exposure to the same TMP epitope p10 or irrelevant TMP epitope p14 (KMAALAASA) (E) or irrelevant MART-1/MelanA or pg100 peptides with or without neutralizing antibodies blocking MHC class I molecules (W6/32) (F). The percentages of effector cells in CD8⁺ T cells (Pt#102, Pt#299, panel E) or in CD8⁺ CD137⁺ T cells (means \pm SEM of Pt#297 and Pt#299, panel F) are shown.

Figure S14



B

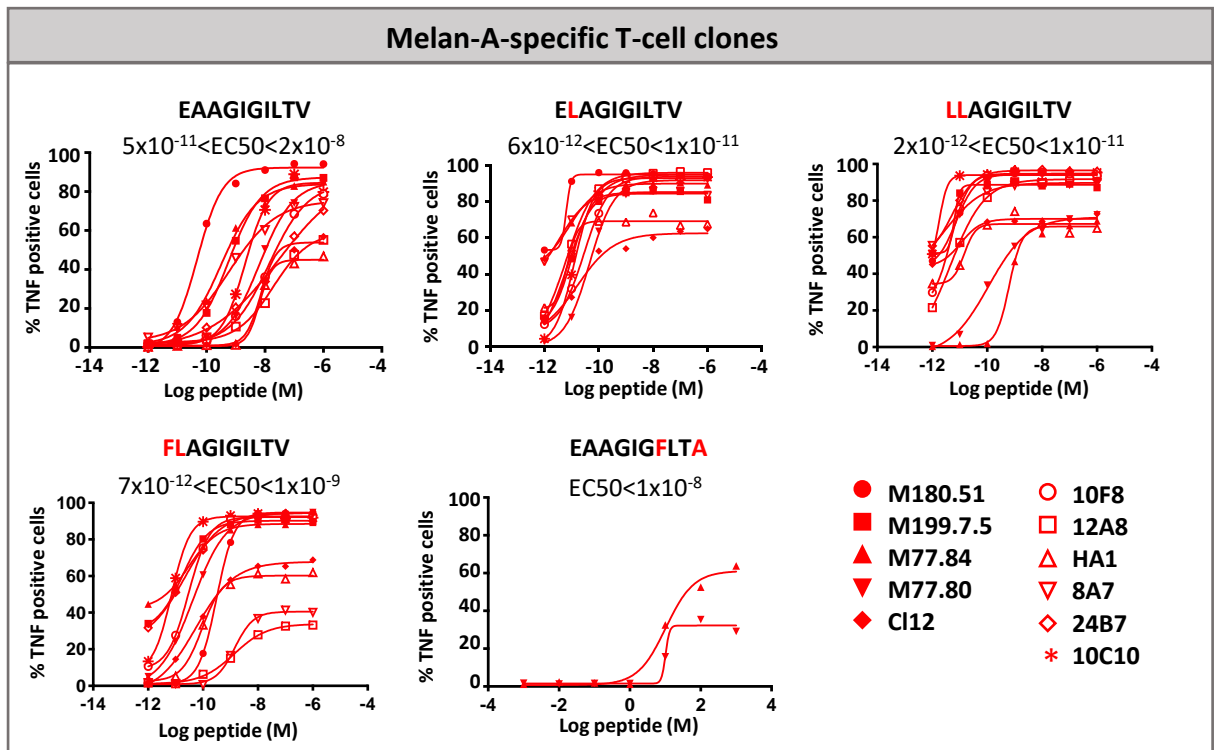
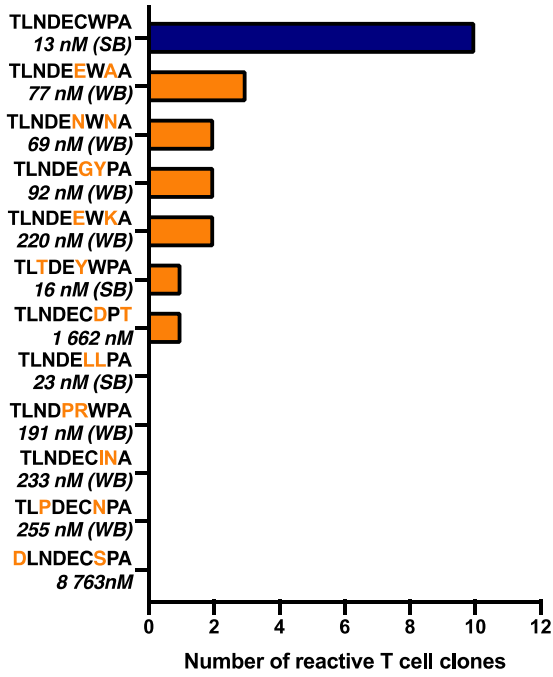


Figure S14. Crossreactivity of T cell clones specific for the MART-1 melanoma peptide with microbial antigens.

A. Numbers of Melan-A specific CD8⁺T cell clones reactive against bacterial peptides. The blue histogram represents the number of clones reactive against their cognate/naturally processed epitope (11/11 T cell clones). The red histogram represents the number of Melan-A-specific T cell clones reactive against the high affinity analog peptide Melan-A A27L. Orange histograms represent the number of crossreactive T-cell clones against each bacterial peptide (as selected by an *in silico* approach, Table S8), the differences in the decapeptide amino-acid sequence being highlighted with orange letters. HLA-A*0201-predicted binding affinities (NetMHCprediction) are indicated for each peptide (strong (SB) *versus* weak (WB) binding affinity). B. Functional avidities of MART-1/Melan-A specific CD8⁺T cell clones in response to naturally processed versus synthetic versus bacterial analogs. Red curves represent T cell clones crossreactive against at least one bacterial peptide. Functional avidities were evaluated by measuring TNF α production in response to T2 cells loaded with a dose range of each indicated peptide, at an E:T ratio of 1:2, by intracellular staining in flow cytometry. Ranges of EC₅₀ for each peptide were calculated using PRISM software.

Figure S15

A



B

MELOE-1-specific T-cell clones

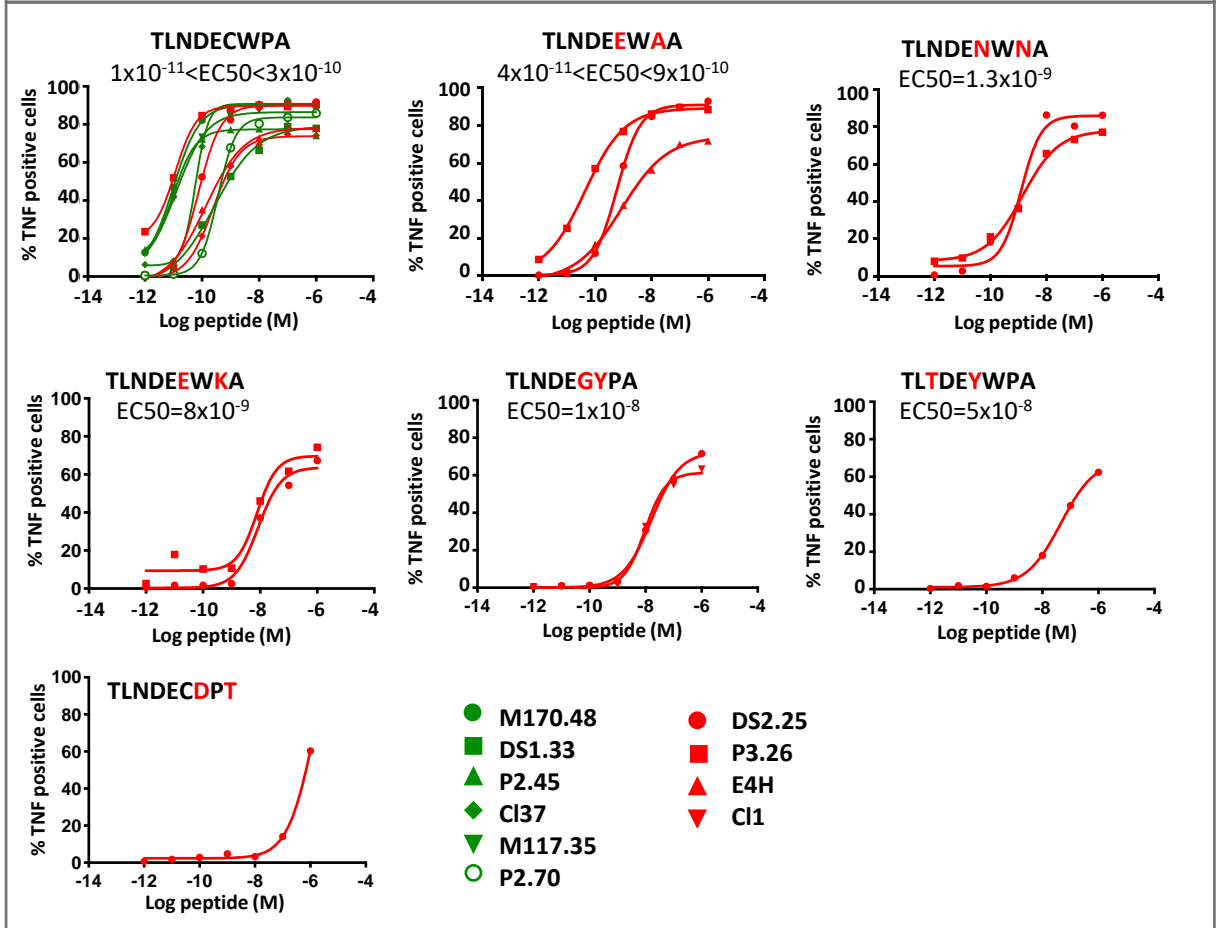


Figure S15. Cross-reactivity of T cell clones specific for the MELOE-1 melanoma peptide with microbial antigens.

A. Numbers of MELOE-1 specific CD8⁺T cell clones reactive against bacterial peptides. The blue histogram represents the number of clones reactive against their cognate/naturally processed epitope (10 T cell clones). Orange histograms represent the number of crossreactive T-cell clones against each bacterial peptide (designed upon *in silico* selection, Table S8), the differences in the decapeptide amino-acid sequence being highlighted with orange letters. HLA-A*0201-predicted binding affinities (NetMHCprediction) are indicated for each peptide (strong (SB) versus weak (WB) binding affinity). B. Functional avidities of MELOE-1 specific CD8⁺T cell clones in response to naturally processed versus bacterial analogs. Red curves represent T cell clones cross-reactive against at least one bacterial peptide and green curves represent MELOE-1-specific T-cell clones reactive only to the cognate peptide TLNDECWPA. Functional avidities were evaluated by measuring TNF α production in response to T2 cells loaded with a dose range of each indicated peptide, at an E:T ratio of 1:2, by flow cytometry. Ranges of EC₅₀ for each peptide were calculated using PRISM software.

Supplemental Tables:**Table S1. Description of *E. hirae* strains.****Table S2. H-2K^b restricted-*E. hirae* epitopes (<50nM).**

We performed sequence alignments of bacterial genes encoding putative cell wall and secreted proteins for immunogenic (13144) *versus* non-immunogenic (708 and 13344) *E. hirae* strains (using the PSORT software), followed by a selection of high affinity epitopes for the MHC class I H-2K^b protein (<50 nM binding affinity) using the NetMHC software.

Table S3. Seeking prophage sequences in *E. hirae* 13144 genomes.**Table S4. List of TRA sequences shared between TMP1 and PSMB4-specific TCRs.****Table S5. List of TRB sequences shared between TMP1 and PSMB4-specific TCRs.****Table S6. List of TMP epitopes selected *in silico* to bind with high affinity (<50nM) HLA-A*0201 molecules.****Table S7. Description of cancer patients treated with anti-PD1 Abs in three independent cohorts (corresponding to Figure S11E-F).****Table S8. Sequence of peptides tested in MART-1 and MELOE-1-specific T cell clones.****Table S9. TCR sequence of MART-1-specific T cell clones.** Recurrent motifs already described in the CDR3 β of MART-1-specific T-cell clones (17) are indicated in bold.**Table S10. TCR sequence of MELOE-1-specific T cell clones.** Recurrent motifs already described in the CDR3 α of MELOE-1 specific –T-cell clones (17) are indicated in bold.

Table S1. Description of *E.hirae* strains

Species	Origin	Cancer	Patient outcome
<i>Enterococcus hirae</i> 13144	Murine – CTX-treated		
<i>Enterococcus hirae</i> 708	Human - Unknown		
<i>Enterococcus hirae</i> 13344	Human - Blood		
<i>Enterococcus hirae</i> ATCC9790	Type strain CIP 53.48T		
<i>Enterococcus hirae</i> 5348	Human - Unknown		
<i>Enterococcus hirae</i> 7030	Human – Liver abscess		
<i>Enterococcus hirae</i> 12607	Environmental – RiskManche project		
<i>Enterococcus hirae</i> 13150	Environmental - Water		
<i>Enterococcus hirae</i> 13152	Environmental - Water		
<i>Enterococcus hirae</i> 13153	Environmental - Water		
<i>Enterococcus hirae</i> 13155	Environmental – RiskManche project		
<i>Enterococcus hirae</i> 13161	Environmental - Cockle		
<i>Enterococcus hirae</i> 13343	Conservation liquid of kidney		
<i>Enterococcus hirae</i> 13346	Human - Urine		
<i>Enterococcus hirae</i> 13347	Blood culture		
<i>Enterococcus hirae</i> IGR1	Human (stool)	Lung	Responder
<i>Enterococcus hirae</i> IGR4	Human (stool)	Lung	Complete Responder
<i>Enterococcus hirae</i> IGR10	Human (stool)	Lung	Responder
<i>Enterococcus hirae</i> IGR11	Human (stool)	Lung	Responder

Table S2. H-2K^b restricted-*E.hirae* epitopes (<50nM binding affinity)

Group	Hirae	sequence	names of the proteins	
Group 1	708	INAKFSSQL	Membrane proteins related to metalloendopeptidases	
	708	YIYNHYKDM	Membrane proteins related to metalloendopeptidases	
	708	YVYGKSRMTM	Membrane proteins related to metalloendopeptidases	
	708	IAFLSYKLF	cell surface protein precursor	
Group 2	708	IMYEYMPV	hypothetical protein	
	708	SSMEYFLKV	Phage tail length tape-measure protein	
	708	ISFFQENQL	Collagen adhesin	
	708	TNLLFMTSL	extracellular protein	
Group 3	708	KIFSIFMLL	Phosphatidylinositol-specific phospholipase C	
	708	LNIFKFNRF	Chitinase	
	708	MTYDYRGGF	Chitinase	
	708	PSYMFRTSF	Chitinase	
Group 4	708	QSYTYMTA	cell wall surface anchor family protein	
	708	ITFSHYEPT	cell wall surface anchor family protein	
	13144	SAFPYEQEL	C3 family ADP-ribosyltransferase	
	13144	YNYSKSYPV	hypothetical protein	
Group 5	13144	VFSHYRPG	hypothetical protein	
	13144	VTFLGYNAF	cell surface protein	
	13144	TVYTFHVNI	cell surface protein	
	13144	TSYSPLFLL	cell surface protein (putative)	
Group 6	13144	TNYIYPNIL	2',3'-cyclic-nucleotide 2'-phosphodiesterase	
	13144	VVPILFLGL	FmtB protein	
	13144	KNYKAYVEL	hypothetical protein	
	13144	SAMKYGIPL	hypothetical protein	
Group 7	13144	TSLARFANI	Phage tail length tape-measure protein	→ TMP1
	13144	AMIEFIQGL	Phage tail length tape-measure protein	→ TMP2
	13144	VAITFGGPL	Phage tail length tape-measure protein	→ TMP3
	13144	VSTNHYGLL	hypothetical protein	
Group 8	13144	VMFGLFITI	cell surface protein precursor	
	13144	TVFSLVSL	Chitinase	
	13144	SIYNLEKPL	IgA1 protease	
	13144	YTIIRYGNL	IgA1 protease	
Group 9	13144	SNGLLYTPM	IgA1 protease	
	13144	NNYHYVGGGL	IgA1 protease	
	13144	SMFLNCNNL	hypothetical protein	
	13144	IAFQGYSSL	hypothetical protein	
Group 10	13144	QVTNFFNMF	hypothetical protein	
	13144	IMLGLFMTM	cell surface protein precursor	
	EH17	MSFTFFSST	hypothetical protein	
	EH17	IAFQNFVNL	Chitinase	
Group 11	EH17	SMFIAFQNF	Chitinase	
	EH17	LNYDYGRI	Chitinase	
	EH17	AGICFFTGV	Peptidoglycan N-acetylglucosamine deacetylase	
	EH17	VEYTYFPTL	Membrane proteins related to metalloendopeptidases	
Group 12	EH17	AAYVFEMNF	Membrane proteins related to metalloendopeptidases	
	EH17	EMYRKLSTL	Membrane proteins related to metalloendopeptidases	
	EH17	YNYGYSVL	enhancin family protein	
	EH17	VIHELNSL	bacteriocin immunity protein	

Table S3. Seeking prophage sequence in *E.hirae* 13144 genome

Region	Region Length	Completeness	Score	# Total Proteins	Region Position	Most Common Phage	GC%
1	40.6Kb	intact	150	58	481066-521729	PHAGE_Enterо_phiEf11_NC_013696(9)	33.79%
2	39.2Kb	intact	140	59	2123983-2163272	PHAGE_Enterо_vB_IME197_NC_028671(6)	34.95%

Table S4. List of TRA sequences shared between TMP1 and PSMB4-specific TCRs

V	J	aaSeqCDR3	CDR3dna	VpJ	VJ	cloneCount
TRAV14D-2	TRAJ22	CAASASSGSWQLIF	TGTGCAGCAAGCGCATCTTCTGGCAGCTGGCAACTCATCTTT	TRAV14D-2 CAASASSGSWQLIF TRAJ22	TRAV14D-2 TRAJ22	57
TRAV14-1	TRAJ26	CAASDNIAQGLTF	TGTGCAGCAAGTGATAACTATGCCAGGGATTAACCTTC	TRAV14-1 CAASDNIAQGLTF TRAJ26	TRAV14-1 TRAJ26	405
TRAV16D-DV11	TRAJ17	CAMRDLSAGNKLTF	TGTGCTATGAGAGACCTTAACAGTGCAGGGAAACAGCTAACTTTT	TRAV16D-DV11 CAMRDLSAGNKLTF TRAJ17	TRAV16D-DV11 TRAJ17	19
TRAV16D-DV11	TRAJ27	CAMREDTNTGKLF	TGTGCTATGAGAGAGGACCAATAACAGGCAAAATAACCTTT	TRAV16D-DV11 CAMREDTNTGKLF TRAJ27	TRAV16D-DV11 TRAJ27	123
TRAV3D-3	TRAJ30	CAVSDTNAYKVI	TGCGCAGTCAGTGACACAAATGCTTACAAGTCATCTTT	TRAV3D-3 CAVSDTNAYKVI TRAJ30	TRAV3D-3 TRAJ30	297
TRAV7-3	TRAJ9	CAVSNMGGYKLF	TGTGCAGTGAGCAACATGGGCTACAACTTACCTTC	TRAV7-3 CAVSNMGGYKLF TRAJ9	TRAV7-3 TRAJ9	208
TRAV8-1	TRAJ18	CATGDRGSALGRHF	TGTGCTACTGGAGATAGAGGTTTTCAGCTTAGGAGGGCTGCATTT	TRAV8-1 CATGDRGSALGRHF TRAJ18	TRAV8-1 TRAJ18	19
TRAV6D-7	TRAJ31	CALGGNSNRRIF	TGTGCTCTGGGGGGGAATAGCAATAACAGAATCTCTTT	TRAV6D-7 CALGGNSNRRIF TRAJ31	TRAV6D-7 TRAJ31	17
TRAV14D-2	TRAJ22	CAASASSGSWQLIF	TGTGCAGCCTCTGCATCTTCTGGCAGCTGGCAACTCATCTTT	TRAV14D-2 CAASASSGSWQLIF TRAJ22	TRAV14D-2 TRAJ22	14
TRAV16N	TRAJ52	CAMRENTGANTGKLF	TGTGCTATGAGAGAGAACTGGAGCTAACACTGGAAAGCTCACGTTT	TRAV16N CAMRENTGANTGKLF TRAJ52	TRAV16N TRAJ52	14
TRAV3D-3	TRAJ21	CAVRDLSNRYLVYF	TGCGCAGTCAGGGATTGTCTAATTACAACGTGCTTACTTC	TRAV3D-3 CAVRDLSNRYLVYF TRAJ21	TRAV3D-3 TRAJ21	10
TRAV14D-1	TRAJ26	CAARNNYAQLTF	TGTGCAGCAAGAAATAACTATGCCAGGGATTAACCTTC	TRAV14D-1 CAARNNYAQLTF TRAJ26	TRAV14D-1 TRAJ26	156
TRAV12D-3	TRAJ33	CALSNYQLIW	TGTGCTCTGAGCAACTATCAGTTGATCTGG	TRAV12D-3 CALSNYQLIW TRAJ33	TRAV12D-3 TRAJ33	8
TRAV12D-3	TRAJ31	CALSDRDSNRRIF	TGTGCTCTGAGTGATCGAGATAGCAATAACAGAATCTCTTT	TRAV12D-3 CALSDRDSNRRIF TRAJ31	TRAV12D-3 TRAJ31	5
TRAV6D-7	TRAJ31	CALGGNSNRRIF	TGTGCTCTGGGTGGGAATAGCAATAACAGAATCTCTTT	TRAV6D-7 CALGGNSNRRIF TRAJ31	TRAV6D-7 TRAJ31	4
TRAV12D-3	TRAJ31	CALSDRDSNRRIF	TGTGCTCTGAGTGATCGGGATAGCAATAACAGAATCTCTTT	TRAV12D-3 CALSDRDSNRRIF TRAJ31	TRAV12D-3 TRAJ31	255
TRAV14D-3-DV8	TRAJ22	CAASASSGSWQLIF	TGTGCAGCAAGTGCAAGTTCTGGCAGCTGGCAACTCATCTTT	TRAV14D-3-DV8 CAASASSGSWQLIF TRAJ22	TRAV14D-3-DV8 TRAJ22	2
TRAV12D-3	TRAJ31	CALSDRHSNRRIF	TGTGCTCTGAGTGTGCACATAGCAATAACAGAATCTCTTT	TRAV12D-3 CALSDRHSNRRIF TRAJ31	TRAV12D-3 TRAJ31	1
TRAV8-1	TRAJ50	CATDPLASSFSKLVF	TGTGCTACTGACCCCTAGCATCTCTCTCTTCAGCAAGCTGGTGTTT	TRAV8-1 CATDPLASSFSKLVF TRAJ50	TRAV8-1 TRAJ50	49
TRAV10	TRAJ27	CAASRGNTGKLF	TGTGCAGCAAGCAGAGGCAACATAACAGGCAAAATAACCTTT	TRAV10 CAASRGNTGKLF TRAJ27	TRAV10 TRAJ27	446
TRAV7-2	TRAJ12	CAAPGTGGYKVV	TGTGCAGCCCGGGGACTGGAGGCTATAAAGTGGTCTTT	TRAV7-2 CAAPGTGGYKVV TRAJ12	TRAV7-2 TRAJ12	360
TRAV6N-6	TRAJ22	CALRAASSGSWQLIF	TGCGCTCTGAGGGCAGCATCTTCTGGCAGCTGGCAACTCATCTTT	TRAV6N-6 CALRAASSGSWQLIF TRAJ22	TRAV6N-6 TRAJ22	332
TRAV13-2	TRAJ26	CAIDQYAQGLTF	TGTGCTATAGACCAATATGCCAGGGATTAACCTTC	TRAV13-2 CAIDQYAQGLTF TRAJ26	TRAV13-2 TRAJ26	296
TRAV14-2	TRAJ44	CAGTGSGGKLT	TGTGCAGGGACTGGCAGTGGTGGAAAACTCACTTTG	TRAV14-2 CAGTGSGGKLT TRAJ44	TRAV14-2 TRAJ44	276
TRAV12D-3	TRAJ23	CALSGENYQGLIF	TGTGCTCTGAGTGGGAGAAATAACAGGGGAAGCTTATCTTT	TRAV12D-3 CALSGENYQGLIF TRAJ23	TRAV12D-3 TRAJ23	256
TRAV19	TRAJ50	CAAGVASSFSKLVF	TGCGCAGCAGGGGGGGTAGCATCTCTCTCTCAGCAAGCTGGTGTTT	TRAV19 CAAGVASSFSKLVF TRAJ50	TRAV19 TRAJ50	236
TRAV13-1	TRAJ6	CALVLTSGGNYKPTF	TGTGCTTTGGTCTAACCTCAGGAGGAACTACAACTCAGTTT	TRAV13-1 CALVLTSGGNYKPTF TRAJ6	TRAV13-1 TRAJ6	228
TRAV6N-6	TRAJ22	CALSVASSGSWQLIF	TGCGCTCTGAGTGATCGCATCTTCTGGCAGCTGGCAACTCATCTTT	TRAV6N-6 CALSVASSGSWQLIF TRAJ22	TRAV6N-6 TRAJ22	230
TRAV12-2	TRAJ58	CALSDPGTGSKLSF	TGTGCTTTGAGTGATCCAGGCACTGGGTCTAAGCTGTCACTT	TRAV12-2 CALSDPGTGSKLSF TRAJ58	TRAV12-2 TRAJ58	182
TRAV5D-4	TRAJ22	CAASTSSGSWQLIF	TGTGCTGCAAGTACATCTTCTGGCAGCTGGCAACTCATCTTT	TRAV5D-4 CAASTSSGSWQLIF TRAJ22	TRAV5D-4 TRAJ22	128
TRAV14D-2	TRAJ23	CAASEDYNQGLIF	TGTGCAGCAAGTGAGGATTATAACAGGGGAAGCTTACTTT	TRAV14D-2 CAASEDYNQGLIF TRAJ23	TRAV14D-2 TRAJ23	100
TRAV13N-1	TRAJ27	CAMEPGTNTGKLF	TGTGCTATGGAACCGGGCAACATAACAGGCAAAATAACCTTT	TRAV13N-1 CAMEPGTNTGKLF TRAJ27	TRAV13N-1 TRAJ27	80
TRAV12D-3	TRAJ31	CALSDRHSNRRIF	TGTGCTCTGAGTGATCGACACAGCAATAACAGAATCTCTTT	TRAV12D-3 CALSDRHSNRRIF TRAJ31	TRAV12D-3 TRAJ31	58
TRAV13N-4	TRAJ28	CVLSLLPGTGSNRLTF	TGTGTTCTGAGTCTGCTACCAGGCACTGGGAGTAACAGGCTCACTTTT	TRAV13N-4 CVLSLLPGTGSNRLTF TRAJ28	TRAV13N-4 TRAJ28	2
TRAV8-1	TRAJ50	CATDPLASSFSKLVF	TGTGCTACTGACCCCTAGCATCTCTCTCTTCAGCAAGCTGGTGTTT	TRAV8-1 CATDPLASSFSKLVF TRAJ50	TRAV8-1 TRAJ50	2
TRAV12-2	TRAJ43	CVRNNNAPRF	TGTGTTGCAATAACAACAATGCCACGATTT	TRAV12-2 CVRNNNAPRF TRAJ43	TRAV12-2 TRAJ43	346
TRAV16N	TRAJ40	CAMRENTGNYKYVF	TGTGCTATGAGAGAGAAATACAGGAACTACAAATACGCTCTT	TRAV16N CAMRENTGNYKYVF TRAJ40	TRAV16N TRAJ40	234
TRAV8D-2	TRAJ9	CATDVGYKLF	TGTGCTACAGATGGGGCTACAACTTACCTTC	TRAV8D-2 CATDVGYKLF TRAJ9	TRAV8D-2 TRAJ9	108
TRAV12-2	TRAJ43	CVRNNNAPRF	TGTGTTGCAATAACAACAATGCCCAAGATT	TRAV12-2 CVRNNNAPRF TRAJ43	TRAV12-2 TRAJ43	30
TRAV16D-DV11	TRAJ17	CAMRDLSAGNKLTF	TGTGCTATGAGAGACCTTAACAGTGCAGGGAAACAGCTAACTTTT	TRAV16D-DV11 CAMRDLSAGNKLTF TRAJ17	TRAV16D-DV11 TRAJ17	2
TRAV3D-3	TRAJ21	CAVRDLSNRYLVYF	TGCGCAGTCAGGGATTGTCTAATTACAACGTGCTTACTTC	TRAV3D-3 CAVRDLSNRYLVYF TRAJ21	TRAV3D-3 TRAJ21	2
TRAV4D-3	TRAJ15	CAADQGRALIF	TGTGCTGCTGACCAAGGAGGAGCAGCTCTGATATT	TRAV4D-3 CAADQGRALIF TRAJ15	TRAV4D-3 TRAJ15	2
TRAV14D-2	TRAJ22	CAASASSGSWQLIF	TGTGCAGCAAGTGCTCTTCTGGCAGCTGGCAACTCATCTTT	TRAV14D-2 CAASASSGSWQLIF TRAJ22	TRAV14D-2 TRAJ22	180
TRAV14D-3-DV8	TRAJ22	CAASASSGSWQLIF	TGTGCAGCAAGTGATCTTCTGGCAGCTGGCAACTCATCTTT	TRAV14D-3-DV8 CAASASSGSWQLIF TRAJ22	TRAV14D-3-DV8 TRAJ22	180
TRAV12D-3	TRAJ33	CALSNYQLIW	TGTGCTCTGAGCAACTATCAGTTGATCTGG	TRAV12D-3 CALSNYQLIW TRAJ33	TRAV12D-3 TRAJ33	135
TRAV6D-7	TRAJ31	CALGGNSNRRIF	TGTGCTCTGGGTGGGAATAGCAATAACAGAATCTCTTT	TRAV6D-7 CALGGNSNRRIF TRAJ31	TRAV6D-7 TRAJ31	37

Table S5. List of TRB sequences shared between TMP1 and PSMB4-specific TCRs

V	J	aaSeqCDR3	CDR3dna	VpJ	VJ	count
TRBV13-3	TRBJ2-3	CARDGAETLYF	TGTGCCAGGCGACGGTGCAGAAACGCTGTATTTT	TRBV13-3 CARDGAETLYF TRBJ2-3	TRBV13-3 TRBJ2-3	318
TRBV12-2	TRBJ2-5	CASFANQDTQYF	TGTGCCAGCGCTTTAAACCAAGACACCCAGTACTTT	TRBV12-2 ASFANQDTQYF TRBJ2-5	TRBV12-2 TRBJ2-5	696
TRBV13-2	TRBJ2-7	CASGDFYEQYF	TGTGCCAGCGGGGACTTTGAAACAGTACTTC	TRBV13-2 CASGDFYEQYF TRBJ2-7	TRBV13-2 TRBJ2-7	16
TRBV13-2	TRBJ2-7	CASGDFYEQYF	TGTGCCAGCGGTGATTTCTATGAACAGTACTTC	TRBV13-2 CASGDFYEQYF TRBJ2-7	TRBV13-2 TRBJ2-7	138
TRBV13-2	TRBJ2-1	CASGDNANSYDF	TGTGCCAGCGGGGCAATGCAAACTCCGACTACCCCTTC	TRBV13-2 CASGDNANSYDF TRBJ2-1	TRBV13-2 TRBJ2-1	42
TRBV13-2	TRBJ2-1	CASGDNANSYDF	TGTGCCAGCGGGGCAATGCAAACTCCGACTACCCCTTC	TRBV13-2 CASGDNANSYDF TRBJ2-1	TRBV13-2 TRBJ2-1	372
TRBV13-2	TRBJ2-4	CASGDRGSQNTLYF	TGTGCCAGCGGTGACAGGGGTAGTCAAACACCTTGACTTT	TRBV13-2 CASGDRGSQNTLYF TRBJ2-4	TRBV13-2 TRBJ2-4	1
TRBV13-2	TRBJ2-4	CASGDRGSQNTLYF	TGTGCCAGCGGTGATCGGGGTAGTCAAACACCTTGACTTT	TRBV13-2 CASGDRGSQNTLYF TRBJ2-4	TRBV13-2 TRBJ2-4	166
TRBV13-2	TRBJ1-4	CASGDSNERLFF	TGTGCCAGCGGGGATCCAACGAAAGATATTTTTT	TRBV13-2 CASGDSNERLFF TRBJ1-4	TRBV13-2 TRBJ1-4	6
TRBV13-2	TRBJ1-4	CASGDSNERLFF	TGTGCCAGCGGTGATCCAACGAAAGATATTTTTT	TRBV13-2 CASGDSNERLFF TRBJ1-4	TRBV13-2 TRBJ1-4	1
TRBV13-2	TRBJ1-4	CASGDSNERLFF	TGTGCCAGCGGTGACAGCAAGAAATATTTTTT	TRBV13-2 CASGDSNERLFF TRBJ1-4	TRBV13-2 TRBJ1-4	45
TRBV13-2	TRBJ2-5	CASGGDRGQDTQYF	TGTGCCAGCGGTGGGACAGGGGGCAAGACACCCAGTACTTT	TRBV13-2 CASGGDRGQDTQYF TRBJ2-5	TRBV13-2 TRBJ2-5	14
TRBV13-2	TRBJ2-1	CASGGTAPIAEQFF	TGTGCCAGCGGTGGGACAGCTCTCTATGCTGAGCAGCTCTTC	TRBV13-2 CASGGTAPIAEQFF TRBJ2-1	TRBV13-2 TRBJ2-1	18
TRBV17	TRBJ2-5	CASGTGTQDTQYF	TGTGCTAGCGGGATGGGACCAAGACACCCAGTACTTT	TRBV17 CASGTGTQDTQYF TRBJ2-5	TRBV17 TRBJ2-5	300
TRBV19	TRBJ1-3	CASRNRRSGNTLYF	TGTGCCAGCAGAAACAGGGGTTCTGGAATACGCTCTATTTT	TRBV19 CASRNRRSGNTLYF TRBJ1-3	TRBV19 TRBJ1-3	584
TRBV13-1	TRBJ2-7	CASSDAAGQYF	TGTGCCAGCAGTGATCGGCTGGGCACTACTTC	TRBV13-1 CASSDAAGQYF TRBJ2-7	TRBV13-1 TRBJ2-7	268
TRBV13-3	TRBJ1-2	CASSDARGQSDYTF	TGTGCCAGAGTATGACCGGGGGCAGACTCCGACTACACCTTC	TRBV13-3 CASSDARGQSDYTF TRBJ1-2	TRBV13-3 TRBJ1-2	618
TRBV13-3	TRBJ2-1	CASSDGAEQFF	TGTGCCAGCAGTGAAGGTGCTGAGCAGTCTTC	TRBV13-3 CASSDGAEQFF TRBJ2-1	TRBV13-3 TRBJ2-1	5
TRBV13-3	TRBJ2-1	CASSDGAEQFF	TGTGCCAGCAGTGAAGGTGCTGAGCAGTCTTC	TRBV13-3 CASSDGAEQFF TRBJ2-1	TRBV13-3 TRBJ2-1	254
TRBV13-1	TRBJ1-4	CASSDGGSNRERLFF	TGTGCCAGCAGTGATGGGGGTTCAAACGAAAGATATTTTTT	TRBV13-1 CASSDGGSNRERLFF TRBJ1-4	TRBV13-1 TRBJ1-4	1
TRBV13-1	TRBJ1-4	CASSDGGSNRERLFF	TGTGCCAGCAGTGATGGGGGTTCAAACGAAAGATATTTTTT	TRBV13-1 CASSDGGSNRERLFF TRBJ1-4	TRBV13-1 TRBJ1-4	228
TRBV13-3	TRBJ1-2	CASSDHANSYTF	TGTGCCAGCAGTGAACCTGCAAACTCCGACTACACCTTC	TRBV13-3 CASSDHANSYTF TRBJ1-2	TRBV13-3 TRBJ1-2	37
TRBV13-1	TRBJ2-7	CASSDRDWWYEQYF	TGTGCCAGCAGTCCGGGACTGGGTCTATGAACAGTACTTC	TRBV13-1 CASSDRDWWYEQYF TRBJ2-7	TRBV13-1 TRBJ2-7	186
TRBV13-1	TRBJ2-4	CASSDRTGGSQNTLYF	TGTGCCAGTGAAGCTGACAGGGGCTCAGTCAAACACCTTGACTTT	TRBV13-1 CASSDRTGGSQNTLYF TRBJ2-4	TRBV13-1 TRBJ2-4	1474
TRBV13-1	TRBJ2-1	CASSDWGNAYEQFF	TGTGCCAGCAGCAGCTGGGGGAACTGCTGAGCAGTCTTC	TRBV13-1 CASSDWGNAYEQFF TRBJ2-1	TRBV13-1 TRBJ2-1	8
TRBV13-1	TRBJ2-1	CASSDWGNAYEQFF	TGTGCCAGCAGTGAAGGTGCTGAGCAGTCTTC	TRBV13-1 CASSDWGNAYEQFF TRBJ2-1	TRBV13-1 TRBJ2-1	128
TRBV13-1	TRBJ2-5	CASSELWGGQDTQYF	TGTGCCAGCAGTGAACCTGGGGGGCAAGACACCCAGTACTTT	TRBV13-1 CASSELWGGQDTQYF TRBJ2-5	TRBV13-1 TRBJ2-5	346
TRBV13-1	TRBJ2-7	CASSEPEYEQYF	TGTGCCAGCAGTGAACCAAGAAATGAACAGTACTTT	TRBV13-1 CASSEPEYEQYF TRBJ2-7	TRBV13-1 TRBJ2-7	130
TRBV12-1	TRBJ2-7	CASSFRDISYEQYF	TGTGCCAGCTCTCCGGGACATCTCTATGAACAGTACTTC	TRBV12-1 CASSFRDISYEQYF TRBJ2-7	TRBV12-1 TRBJ2-7	124
TRBV12-1	TRBJ2-7	CASSFRDISYEQYF	TGTGCCAGCTCTCCGGGACAGCTCTATGAACAGTACTTC	TRBV12-1 CASSFRDISYEQYF TRBJ2-7	TRBV12-1 TRBJ2-7	38
TRBV14	TRBJ2-7	CASSFRPGYEQYF	TGTGCCAGCAGTTTCAGGGGTCCTATGAACAGTACTTC	TRBV14 CASSFRPGYEQYF TRBJ2-7	TRBV14 TRBJ2-7	62
TRBV14	TRBJ2-7	CASSFRVPYEQYF	TGTGCCAGCAGTTTCAGGGGTCCTATGAACAGTACTTC	TRBV14 CASSFRVPYEQYF TRBJ2-7	TRBV14 TRBJ2-7	288
TRBV12-1	TRBJ2-4	CASSGDRDKNTLYC	TGTGCCAGCTTGGGCGACAGGGGACAAAACACCTTGACTGT	TRBV12-1 CASSGDRDKNTLYC TRBJ2-4	TRBV12-1 TRBJ2-4	2
TRBV12-1	TRBJ2-4	CASSGDRDKNTLYC	TGTGCCAGCTTGGGCGACAGGGGACAAAACACCTTGACTTT	TRBV12-1 CASSGDRDKNTLYC TRBJ2-4	TRBV12-1 TRBJ2-4	326
TRBV13-3	TRBJ2-1	CASSGTRNSDYTF	TGTGCCAGCAGTGAACCAAGAACTCCGACTACACCTTC	TRBV13-3 CASSGTRNSDYTF TRBJ2-1	TRBV13-3 TRBJ2-1	12
TRBV13-3	TRBJ2-1	CASSGTRNSDYTF	TGTGCCAGCAGTGAACCAAGAACTCCGACTACACCTTC	TRBV13-3 CASSGTRNSDYTF TRBJ2-1	TRBV13-3 TRBJ2-1	288
TRBV12-1	TRBJ2-3	CASSGTTSAETLYF	TGTGCCAGCTCCGGGACAACTAGTGCAGAACCTTGACTTT	TRBV12-1 CASSGTTSAETLYF TRBJ2-3	TRBV12-1 TRBJ2-3	2
TRBV19	TRBJ2-3	CASSIGGTTSAETLYF	TGTGCCAGCAGTATAGGGGGGACTCTAGTGCAGAAACGCTGTATTTT	TRBV19 CASSIGGTTSAETLYF TRBJ2-3	TRBV19 TRBJ2-3	532
TRBV19	TRBJ2-3	CASSIGGTTSAETLYF	TGTGCCAGCAGTATAGGGGGGACTCTAGTGCAGAAACGCTGTATTTT	TRBV19 CASSIGGTTSAETLYF TRBJ2-3	TRBV19 TRBJ2-3	1
TRBV17	TRBJ2-7	CASSITGAYEQYF	TGTGCTAGCAGTATAGGGGACAGGGGCTATGAACAGTACTTC	TRBV17 CASSITGAYEQYF TRBJ2-7	TRBV17 TRBJ2-7	228
TRBV12-2	TRBJ2-5	CASSLKDQTYF	TGTGCCAGCTCTCCGCAAGAACACCCAGTACTTT	TRBV12-2 CASSLKDQTYF TRBJ2-5	TRBV12-2 TRBJ2-5	14
TRBV12-2	TRBJ2-5	CASSLKDQTYF	TGTGCCAGCTCTCCGCAAGAACACCCAGTACTTT	TRBV12-2 CASSLKDQTYF TRBJ2-5	TRBV12-2 TRBJ2-5	2
TRBV12-2	TRBJ2-3	CASSLSSAETLYF	TGTGCCAGCTCTCGACTAGTGCAGAAACGCTGTATTTT	TRBV12-2 CASSLSSAETLYF TRBJ2-3	TRBV12-2 TRBJ2-3	386
TRBV16	TRBJ2-4	CASSLERGASQNTLYF	TGTGCAAGCAGTTAGAAAGGGGAGTGTCAAACACCTTGACTTT	TRBV16 CASSLERGASQNTLYF TRBJ2-4	TRBV16 TRBJ2-4	454
TRBV16	TRBJ2-4	CASSLETGGARQNTLYF	TGTGCAAGCAGTTAGAAACTGGGGGGGCGCTCAAACACCTTGACTTT	TRBV16 CASSLETGGARQNTLYF TRBJ2-4	TRBV16 TRBJ2-4	114
TRBV16	TRBJ2-5	CASSLGGQDTQYF	TGTGCAAGCAGTCTGGGGGGCAAGACACCCAGTACTTT	TRBV16 CASSLGGQDTQYF TRBJ2-5	TRBV16 TRBJ2-5	33
TRBV16	TRBJ2-5	CASSLGGQDTQYF	TGTGCAAGCAGTCTGGGGGGCAAGACACCCAGTACTTT	TRBV16 CASSLGGQDTQYF TRBJ2-5	TRBV16 TRBJ2-5	7
TRBV16	TRBJ2-5	CASSLGGQDTQYF	TGTGCAAGCAGTCTAGGGGGCAAGACACCCAGTACTTT	TRBV16 CASSLGGQDTQYF TRBJ2-5	TRBV16 TRBJ2-5	260
TRBV26	TRBJ1-1	CASSLGGQINTEVFF	TGTGCCAGCAGTCTGGGCAAAACCAAGAAAGTCTCTTT	TRBV26 CASSLGGQINTEVFF TRBJ1-1	TRBV26 TRBJ1-1	434
TRBV26	TRBJ1-1	CASSLGGQINTEVFF	TGTGCCAGCAGTCTGGGCAAAACCAAGAAAGTCTCTTT	TRBV26 CASSLGGQINTEVFF TRBJ1-1	TRBV26 TRBJ1-1	4
TRBV26	TRBJ1-1	CASSLGGQINTEVFF	TGTGCCAGCAGTCTGGGCAAAACCAAGAAAGTCTCTTT	TRBV26 CASSLGGQINTEVFF TRBJ1-1	TRBV26 TRBJ1-1	28
TRBV12-2	TRBJ2-5	CASSLMGNQDTQYF	TGTGCCAGCTCTCATGGGGAACCAAGACACCCAGTACTTT	TRBV12-2 CASSLMGNQDTQYF TRBJ2-5	TRBV12-2 TRBJ2-5	9
TRBV12-2	TRBJ2-5	CASSLMGNQDTQYF	TGTGCCAGCTCTCATGGGGAACCAAGACACCCAGTACTTT	TRBV12-2 CASSLMGNQDTQYF TRBJ2-5	TRBV12-2 TRBJ2-5	2
TRBV29	TRBJ1-4	CASSLSSNERLFF	TGTGCTAGCAGTTAAAGTATCCAAACGAAATATTTTTT	TRBV29 CASSLSSNERLFF TRBJ1-4	TRBV29 TRBJ1-4	370
TRBV19	TRBJ1-3	CASSMEETSSGNTLYF	TGTGCCAGCAGTATGAGGGAGACATCTGAAACACCTTGACTTT	TRBV19 CASSMEETSSGNTLYF TRBJ1-3	TRBV19 TRBJ1-3	2
TRBV19	TRBJ1-1	CASSNRENTVEFF	TGTGCCAGCAGCAACAGGGGAAACACAGAAAGTCTCTTT	TRBV19 CASSNRENTVEFF TRBJ1-1	TRBV19 TRBJ1-1	25
TRBV19	TRBJ1-1	CASSNRENTVEFF	TGTGCCAGCAGCAACAGGGGAAACACAGAAAGTCTCTTT	TRBV19 CASSNRENTVEFF TRBJ1-1	TRBV19 TRBJ1-1	2
TRBV14	TRBJ2-1	CASSPRDGYAEQFF	TGTGCCAGCAGCAAGCAGAGGGGATGCTGAGCAGTCTTC	TRBV14 CASSPRDGYAEQFF TRBJ2-1	TRBV14 TRBJ2-1	18
TRBV14	TRBJ2-1	CASSPRDGYAEQFF	TGTGCCAGCAGCAGCAGAGGGGATGCTGAGCAGTCTTC	TRBV14 CASSPRDGYAEQFF TRBJ2-1	TRBV14 TRBJ2-1	2
TRBV21	TRBJ2-4	CASSPGQASQNTLYF	TGTGCTAGCAGTCCGGGACAGGGGCGACTCAAACACCTTGACTTT	TRBV21 CASSPGQASQNTLYF TRBJ2-4	TRBV21 TRBJ2-4	165
TRBV29	TRBJ2-7	CASSPGTGGYEQYF	TGTGCTAGCAGTCCGGGACAGGGGGTATGAACAGTACTTC	TRBV29 CASSPGTGGYEQYF TRBJ2-7	TRBV29 TRBJ2-7	566
TRBV29	TRBJ2-5	CASSPGTGNQDTQYF	TGTGCTAGCAGCCCGGGAACAGGAAACCAAGACACCCAGTACTTT	TRBV29 CASSPGTGNQDTQYF TRBJ2-5	TRBV29 TRBJ2-5	47
TRBV29	TRBJ2-5	CASSPGTGNQDTQYF	TGTGCTAGCAGCCCGGGAACAGGAAACCAAGACACCCAGTACTTT	TRBV29 CASSPGTGNQDTQYF TRBJ2-5	TRBV29 TRBJ2-5	2
TRBV29	TRBJ2-3	CASSPGTNSAETLYF	TGTGCTAGCAGCCCGGGAACAAATAGTGCAGAAACGCTGTATTTT	TRBV29 CASSPGTNSAETLYF TRBJ2-3	TRBV29 TRBJ2-3	262
TRBV4	TRBJ1-1	CASSPQDTEVFF	TGTGCCAGCAGCCCCAGGACACAGAAAGTCTCTTT	TRBV4 CASSPQDTEVFF TRBJ1-1	TRBV4 TRBJ1-1	28
TRBV4	TRBJ1-1	CASSPQDTEVFF	TGTGCCAGCAGCCCCAGGACACAGAAAGTCTCTTT	TRBV4 CASSPQDTEVFF TRBJ1-1	TRBV4 TRBJ1-1	2
TRBV2	TRBJ2-7	CASSQDLGGRWEQYF	TGTGCCAGCAGCAAGCAGTGGGGGGCTGGGAAACAGTACTTC	TRBV2 CASSQDLGGRWEQYF TRBJ2-7	TRBV2 TRBJ2-7	658
TRBV5	TRBJ2-4	CASSQENGGSQNTLYF	TGTGCCAGCAGCAAGAAAGTGGGGGACTGAAACACCTTGACTTT	TRBV5 CASSQENGGSQNTLYF TRBJ2-4	TRBV5 TRBJ2-4	762
TRBV5	TRBJ1-3	CASSQRDRGSGNTLYF	TGTGCCAGCAGCAACGGGACAGGGGATCTGGAATACGCTCTATTTT	TRBV5 CASSQRDRGSGNTLYF TRBJ1-3	TRBV5 TRBJ1-3	59
TRBV5	TRBJ1-3	CASSQRDRGSGNTLYF	TGTGCCAGCAGCAACGGGACAGGGGATCTGGAATACGCTCTATTTT	TRBV5 CASSQRDRGSGNTLYF TRBJ1-3	TRBV5 TRBJ1-3	2
TRBV5	TRBJ2-4	CASSQLVGSQNTLYF	TGTGCCAGCAGCAAGTCTGGGGGAGTCAAACACCTTGACTTT	TRBV5 CASSQLVGSQNTLYF TRBJ2-4	TRBV5 TRBJ2-4	282
TRBV17	TRBJ2-7	CASSRDRSYEQYF	TGTGCTAGCAGTGAAGGAGACATCTATGAACAGTACTTC	TRBV17 CASSRDRSYEQYF TRBJ2-7	TRBV17 TRBJ2-7	4
TRBV17	TRBJ2-7	CASSRDRSYEQYF	TGTGCTAGCAGTGAAGGAGACATCTATGAACAGTACTTC	TRBV17 CASSRDRSYEQYF TRBJ2-7	TRBV17 TRBJ2-7	28
TRBV17	TRBJ2-7	CASSRGGEQYF	TGTGCTAGCAGTGGGGGGTGAACAGTACTTC	TRBV17 CASSRGGEQYF TRBJ2-7	TRBV17 TRBJ2-7	130
TRBV12-1	TRBJ2-4	CASSRGLGGRQNTLYF	TGTGCCAGCTCTCGGGAAGTGGGGGGCGCAAAACACCTTGACTTT	TRBV12-1 CASSRGLGGRQNTLYF TRBJ2-4	TRBV12-1 TRBJ2-4	162
TRBV12-1	TRBJ1-4	CASSRPNRERLFF	TGTGCCAGCTCTCGCCAAACGAAAGATATTTTTT	TRBV12-1 CASSRPNRERLFF TRBJ1-4	TRBV12-1 TRBJ1-4	572
TRBV15	TRBJ2-4	CASSRRESQNTLYF	TGTGCCAGCAGCCCGGGAAGTCAAACACCTTGACTTT	TRBV15 CASSRRESQNTLYF TRBJ2-4	TRBV15 TRBJ2-4	138
TRBV15	TRBJ2-4	CASSRRESQNTLYF	TGTGCCAGCAGCCCGGGAAGTCAAACACCTTGACTTT	TRBV15 CASSRRESQNTLYF TRBJ2-4	TRBV15 TRBJ2-4	2
TRBV19	TRBJ2-4	CASSRTGGQNTLYF	TGTGCCAGCAGTGAAGTGGGGTCAAACACCTTGACTTT	TRBV19 CASSRTGGQNTLYF TRBJ2-4	TRBV19 TRBJ2-4	136
TRBV17	TRBJ2-4	CASSSGGGQNTLYF	TGTGCTAGCAGTCCGGGGGGTCAAACACCTTGACTTT	TRBV17 CASSSGGGQNTLYF TRBJ2-4	TRBV17 TRBJ2-4	312
TRBV29	TRBJ1-6	CASSSGGNSPLYF	TGTGCTAGCAGTCCGGGGGAAATCGCCCTCTACTTT	TRBV29 CASSSGGNSPLYF TRBJ1-6	TRBV29 TRBJ1-6	9
TRBV29	TRBJ1-6	CASSSGGNSPLYF	TGTGCTAGCAGTCCGGGGGAAATCGCCCTCTACTTT	TRBV29 CASSSGGNSPLYF TRBJ1-6	TRBV29 TRBJ1-6	616
TRBV29	TRBJ1-6	CASSSGGNSPLYF	TGTGCTAGCAGTCCGGGGGAAATCGCCCTCTACTTT	TRBV29 CASSSGGNSPLYF TRBJ1-6	TRBV29 TRBJ1-6	58
TRBV19	TRBJ1-4	CASSSGQSERLFF	TGTGCCAGCAGTCCGGGACAGGGAAGAAAGATATTTTTT	TRBV19 CASSSGQSERLFF TRBJ1-4	TRBV19 TRBJ1-4	24
TRBV12-1	TRBJ2-1	CASSSTGGSDYTF	TGTGCCAGCTCTCCGGGACAGGGGGTCCGACTACACCTTC	TRBV12-1 CASSSTGGSDYTF TRBJ2-1	TRBV12-1 TRBJ2-1	312
TRBV17	TRBJ2-4	CASSTLGQNTLYF	TGTGCTAGCAGTCCGGGGTAAAGTCAAACACCTTGACTTT	TRBV17 CASSTLGQNTLYF TRBJ2-4	TRBV17 TRBJ2-4	412
TRBV19	TRBJ1-3	CASSWDSSGNTLYF	TGTGCCAGCAGTGGGACAGCTCTGGAATACGCTCTATTTT	TRBV19 CASSWDSSGNTLYF TRBJ1-3	TRBV19 TRBJ1-3	7788
TRBV31	TRBJ1-1	CASSLRGANTVEFF	TGTGCTGGAAGTAAAGGGTGAACAAACAGAAAGTCTCTTT	TRBV31 CASSLRGANTVEFF TRBJ1-1	TRBV31 TRBJ1-1	1
TRBV31	TRBJ1-1	CASSLRGANTVEFF	TGTGCTGGAAGTAAAGGGTGAACAAACAGAAAGTCTCTTT	TRBV31 CASSLRGANTVEFF TRBJ1-1	TRBV31 TRBJ1-1	432
TRBV1	TRBJ2-7	CTCSADRAGGYEQYF	TGCACTTGCAGTGCAGATAGGGGACGGGGTGAACAGTACTTC	TRBV1 CTCSADRAGGYEQYF TRBJ2-7	TRBV1 TRBJ2-7	200
TRBV29	TRBJ1-3	CVSSPGTGGYEQYF	TGTGCTAGCAGTCCGGGACAGGGGCTGAGCAGTACTTC	TRBV29 CVSSPGTGGYEQYF TRBJ1-3	TRBV29 TRBJ1-3	2
TRBV19	TRBJ1-3	GDSSWESSGNTRYF	GGGGACAGCAGTGGGAAAGCTGGAATACGCGATATTTT	TRBV19 GDSSWESSGNTRYF TRBJ1-3	TRBV19 TRBJ1-3	2
TRBV21	TRBJ2-1	YAEQFF	TATGCTGAGCAGTCTTC	TRBV21 YAEQFF TRBJ2-1	TRBV21 TRBJ2-1	366

Table S6. List of TMP epitopes selected in silico to bind HLA-A2 with high affinity (<50nM)

Peptide	Start	Stop	HLA	Sequence	Affinity(nM)
1	357	365	HLA-A0201	AMIEFIQGL	4.88
2	1462	1470	HLA-A0201	KMVEILEEI	7.8
3	1397	1405	HLA-A0201	RLLKYDVGV	11.55
4	765	773	HLA-A0201	TLVGVTFAI	16.94
5	1374	1382	HLA-A0201	AMQNLVAAV	17.32
6	793	801	HLA-A0201	AIMAIANGV	20.56
7	862	870	HLA-A0201	AMSMNMEEV	24.83
8	504	512	HLA-A0201	KVFGKMTSV	26.84
9	1130	1138	HLA-A0201	LLGIYQSYV	29.4
10	631	639	HLA-A0201	KLAKFASVV	29.89
11	1176	1184	HLA-A0201	KLWANMSKA	30.99
12	692	700	HLA-A0201	MLSNPITAI	32.68
13	700	708	HLA-A0201	ILVAITTTI	36.32
14	491	499	HLA-A0201	KMAALAASA	46.27
15	691	699	HLA-A0201	AMLSNPITA	49.58
16	473	481	HLA-A0201	NMAEAFASA	49.85



Table S7. Patient characteristics. Description corresponding to Figure 4E-F

Stage IIIC/IV NSCLC	Cohort (CHUM)	Cohort (CGFL)	Cohort (validation)
Numbers (n)	44	62	51
Age (mean, range)	65 (45-81)	65,5 (46-85)	65 (42-83)
Gender (n)			
Male	20 (45.5%)	48 (77.4%)	25 (49%)
Female	24 (54.5%)	14 (22.6%)	26 (51%)
Smokers (n)			
Yes	41 (93.2%)	54 (87.1%)	41 (80.4%)
No	3 (6.8%)	6 (9.7%)	10 (19.6%)
NA		2 (3.2%)	0 (0%)
Histology (n)			
Adenocarcinoma	32 (72.7%)	30 (48.4%)	47 (92.2%)
Squamous cell carcinoma	8 (18.2%)	31 (50%)	4 (7.8%)
Other	4 (9.1%)	1 (1.6%)	0 (0%)
Immunotherapy (n)			
Pembrolizumab	23 (52.3%)	0 (0%)	20 (39.2%)
Nivolumab	21 (47.7%)	62 (100%)	28 (55%)
Atezolizumab	0 (0%)	0 (0%)	3 (5.8%)
Line of therapy (n)			
1L	7 (15.9%)	0 (0%)	8 (15.7%)
2L	36 (81.8%)	62 (100%)	43 (84.3%)
NA	1 (2.3%)	0 (0%)	0 (0%)
PDL-1 status (n)			
>50%	20 (45.5%)	16 (25.8%)	14 (27.5%)
<50%	11 (25%)	33 (53.2%)	37 (72.5%)
NA	13 (29.5%)	13 (21%)	0 (0%)

Table S8. Peptide sequences

Peptide_Name	Peptide_Sequence
MART-1_A2_26-35_WT	EAAGIGILIV
MART-1_A2_26-35_Mut	ELAGIGILIV
MART-1_A2_26-35_B1	EAAGIGILAT
MART-1_A2_26-35_B2	EAAGIGFLTA
MART-1_A2_26-35_B3	FLAGIGILTV
MART-1_A2_26-35_B4	ILAGSGILTV
MART-1_A2_26-35_B5	LLAGIGILTV
MELOE-1_A2_36-44_WT	TLNDECWPA
MELOE-1_A2_36-44_B1	DLNDECSPA
MELOE-1_A2_36-44_B2	TLNDEC DPT
MELOE-1_A2_36-44_B3	TLNDECINA
MELOE-1_A2_36-44_B4	TLNDEEWAA
MELOE-1_A2_36-44_B5	TLNDEEWKA
MELOE-1_A2_36-44_B6	TLNDEGYPA
MELOE-1_A2_36-44_B7	TLNDELLPA
MELOE-1_A2_36-44_B8	TLNDE N WNA
MELOE-1_A2_36-44_B9	TLNDPRWPA
MELOE-1_A2_36-44_B10	TL P DEC N PA
MELOE-1_A2_36-44_B11	TL T DE Y WPA

Table S9. CDR3 alpha and beta sequences of MART-1-specific T-cell clones

MART-1-specific T-cell clones					
T-cell clone	TRBV	CDR3beta	TRAV	CDR3alpha	origin ¹
10C10	4-3	CASSPGTLSDTQYFG	12-2	CAVNLEGNNRLAFG	Patient PBMC
12A8	20-1	CSARD GLG ELFFG ²	12-2	CAVNFDQGTGANNLFFG	
24B7	4-2	CASSQDRGGAETQYFG	12-2	CAASQGFQKLVFG	
CI12	19	CASRWGYLSNQPQHFG	35	CAGLGAQKLVFG	
10F8	20-1	CSARD GLG ELFFG	12-2	CAVNLEGNNRLAFG	
8A7	5-5	CASSSGEGLDTQYFG	12-2	CAVKAIYFG	HV PBMC
HA1	25-1	CASSEPYKETQYFG	12-2	CAVGTGTYKYIFG	TIL
M77-84	6-1	CASSEVAWGRAETQYFG	39	CAVDIVPTNDYKLSFG	
M77-80	28	CASTSALLAGGEQYFG	29	CAASVNARLMFG	
M199.75	28	CASSLQ GLG TEAFFG	12-2	CALNQAGTALIFG	
M180.51	28	CASSFE GLG TEAFFG	19	CALSDNTNAGKSTFG	

¹ MART-1 specific T-cell clones were obtained either from PBMC of melanoma patients or healthy volunteers, after a step of peptide stimulation, followed by HLA-p/multimer sorting and cloning by limiting dilution, or directly from TIL spontaneously enriched in Melan-A specific T lymphocytes (Godet et al., Eur J Immunol. 2010).

² In bold are indicated recurrent motifs already described in the CDR3 β of MART-1-specific T-cell clones (Simon et al, Front Immunol., 2018).

Table S10. CDR3 alpha and beta sequences of MELOE-1-specific T-cell clones

MELOE-1-specific T-cell clones					
T-cell clone	TRBV	CDR3beta	TRAV	CDR3alpha	origin ¹
P2.70	10-3	CAISESWGRDTEAFFG	19	CALSEAKYNQGGKLIFG	Patient PBMC
P2.45	14	CASSQPSRDRKDNEQFFG	19	CALSGPLLGTSYGKLTFG ²	
P3.26	3-1	CASSQSGTSGRRDNEQFFG	19	CALSGPISGGGADGLTFG	
CI1	10-3	CAIARTANYGYTFG	24	CAFIQGNNDMRFG	
CI37	14	CASSQERDRGRTNEQFFG	19	CALSGPILTG GGNKLTFG	
E4H	11-1	CASSVQVSGANVLTFG	17	CASRGTPLVFG	HV PBMC
DS1.33	7-2	CASSSGLAGTRNYEQYFG	19	CALRGPMDTGRRALTFG	
DS2.25	20-1	CSATSLAGIDYGYTFG	19	CALSGPFSGGYNKLIFG	TIL
M170.48	3-1	CASSHKWKREPTDTQYFG	19	CALSGPFS DGQKLLFA	
M117.35	19	CASSISEPARRDNEQFFG	19	CALRGPILTG GGNKLTFG	

¹ MELOE-1 specific T-cell clones were obtained either from PBMC of melanoma patients or healthy volunteers, after a step of peptide stimulation, followed by HLA-p/multimer sorting and cloning by limiting dilution, or directly from TIL spontaneously enriched in MELOE-1 specific T lymphocytes (Godet et al., Eur J Immunol. 2010).

² In bold are indicated recurrent motifs already described in the CDR3 α of MELOE-1 specific –T-cell clones (Simon et al, Front Immunol., 2018).

In red are indicated T cell clones cross-reactive against at least one bacterial peptide.

Statistical Report

“Cross-reactivity between MHC class I-restricted antigens from cancer cells and an enterococcal bacteriophage”

(All tests are non-parametric, due to the absence of Gaussian distribution assumption)

---- Figure 1 ----

Fig.1B

Table Analyzed Fig 1B

Kruskal-Wallis test

P value	0,0023
Exact or approximate P value?	Approximate
P value summary	**
Do the medians vary signif. (P < 0.05)	Yes
Number of groups	5
Kruskal-Wallis statistic	16,62

Table Analyzed Fig 1B

Column B	CTX + 13144
vs.	vs,
Column A	CTX
Mann Whitney test	
P value	0,0030
Exact or approximate P value?	Exact
P value summary	**
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,B	302,0 , 163,0
Mann-Whitney U	43,00
Difference between medians	
Median of column A	140,0
Median of column B	72,00
Difference: Actual	-68,00
Difference: Hodges-Lehmann	-64,25

Table Analyzed Fig 1B

Column E	CTX + IGR11
vs.	vs,
Column A	CTX
Mann Whitney test	
P value	0,0202
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,E	288,0 , 177,0
Mann-Whitney U	57,00
Difference between medians	
Median of column A	140,0
Median of column E	88,00
Difference: Actual	-52,00
Difference: Hodges-Lehmann	-52,00

Table Analyzed Fig 1B

Column C	CTX + 13344
vs.	vs,
Column A	CTX
Mann Whitney test	
P value	0,1698
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed

Sum of ranks in column A,C	266,0 , 199,0
Mann-Whitney U	79,00
Difference between medians	
Median of column A	140,0
Median of column C	110,5
Difference: Actual	-29,50
Difference: Hodges-Lehmann	-26,00

Table Analyzed	Fig 1B
Column D	CTX + ATCC9790
vs.	vs,
Column A	CTX
Mann Whitney test	
P value	0,1512
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,D	267,5 , 197,5
Mann-Whitney U	77,50
Difference between medians	
Median of column A	140,0
Median of column D	115,0
Difference: Actual	-25,00
Difference: Hodges-Lehmann	-30,00

Table Analyzed	Fig 1B
Column D	CTX + ATCC9790
vs.	vs,
Column B	CTX + 13144
Mann Whitney test	
P value	0,0055
Exact or approximate P value?	Exact
P value summary	**
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column B,D	167,0 , 298,0
Mann-Whitney U	47,00
Difference between medians	
Median of column B	72,00
Median of column D	115,0
Difference: Actual	43,00
Difference: Hodges-Lehmann	36,00

Table Analyzed	Fig 1B
Column C	CTX + 13344
vs.	vs,
Column B	CTX + 13144
Mann Whitney test	
P value	0,0063
Exact or approximate P value?	Exact
P value summary	**
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column B,C	168,0 , 297,0
Mann-Whitney U	48,00
Difference between medians	
Median of column B	72,00
Median of column C	110,5
Difference: Actual	38,50
Difference: Hodges-Lehmann	39,00

Table Analyzed	Fig 1B
Column E	CTX + IGR11
vs.	vs,
Column D	CTX + ATCC9790
Mann Whitney test	

P value	0,0992
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column D,E	272,5 , 192,5
Mann-Whitney U	72,50
Difference between medians	
Median of column D	115,0
Median of column E	88,00
Difference: Actual	-27,00
Difference: Hodges-Lehmann	-23,00

Table Analyzed	Fig 1B
Column E	CTX + IGR11
vs.	vs,
Column C	CTX + 13344
Mann Whitney test	
P value	0,0179
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column C,E	289,0 , 176,0
Mann-Whitney U	56,00
Difference between medians	
Median of column C	110,5
Median of column E	88,00
Difference: Actual	-22,50
Difference: Hodges-Lehmann	-22,50

Fig.1D

Table Analyzed	Fig 1D - CTX		
Kruskal-Wallis test			
P value	0,1836		
Exact or approximate P value?	Gaussian Approximation		
P value summary	ns		
Do the medians vary signif. (P < 0.05)	No		
Number of groups	5		
Kruskal-Wallis statistic	6,216		
Dunn's Multiple Comparison Test	Difference in rank sum	Significant? P < 0.05?	Summary
No peptide vs TMP1	-22,02	No	ns
No peptide vs TMP2	-11,30	No	ns
No peptide vs TMP3	-19,74	No	ns
No peptide vs Hypothetical protein	-13,89	No	ns
TMP1 vs TMP2	10,72	No	ns
TMP1 vs TMP3	2,283	No	ns
TMP1 vs Hypothetical protein	8,130	No	ns
TMP2 vs TMP3	-8,435	No	ns
TMP2 vs Hypothetical protein	-2,587	No	ns
TMP3 vs Hypothetical protein	5,848	No	ns

Table Analyzed	Fig 1D – CTX + Eh13144		
Kruskal-Wallis test			
P value	0,0028		
Exact or approximate P value?	Gaussian Approximation		
P value summary	**		
Do the medians vary signif. (P < 0.05)	Yes		
Number of groups	5		
Kruskal-Wallis statistic	16,17		
Dunn's Multiple Comparison Test	Difference in rank sum	Significant? P < 0.05?	Summary
No peptide vs TMP1	-31,42	Yes	**
No peptide vs TMP2	-22,03	No	ns

No peptide vs TMP3	-11,11	No	ns
No peptide vs Hypothetical protein	-7,667	No	ns
TMP1 vs TMP2	9,389	No	ns
TMP1 vs TMP3	20,31	No	ns
TMP1 vs Hypothetical protein	23,75	No	ns
TMP2 vs TMP3	10,92	No	ns
TMP2 vs Hypothetical protein	14,36	No	ns
TMP3 vs Hypothetical protein	3,444	No	ns

Table Analyzed Fig 1D – CTX + Eh IGR11

Kruskal-Wallis test			
P value	0,0131		
Exact or approximate P value?	Gaussian Approximation		
P value summary *			
Do the medians vary signif. (P < 0.05)	Yes		
Number of groups	5		
Kruskal-Wallis statistic	12,66		
Dunn's Multiple Comparison Test			
	Difference in rank sum	Significant? P < 0.05?	Summary
No peptide vs TMP1	-20,90	Yes	*
No peptide vs TMP2	-18,00	No	ns
No peptide vs TMP3	-9,200	No	ns
No peptide vs Hypothetical protein	-11,40	No	ns
TMP1 vs TMP2	2,900	No	ns
TMP1 vs TMP3	11,70	No	ns
TMP1 vs Hypothetical protein	9,500	No	ns
TMP2 vs TMP3	8,800	No	ns
TMP2 vs Hypothetical protein	6,600	No	ns
TMP3 vs Hypothetical protein	-2,200	No	ns

Table Analyzed Fig 1D – CTX + ATCC9790

Kruskal-Wallis test			
P value	0,0711		
Exact or approximate P value?	Gaussian Approximation		
P value summary	ns		
Do the medians vary signif. (P < 0.05)	No		
Number of groups	5		
Kruskal-Wallis statistic	8,627		
Dunn's Multiple Comparison Test			
	Difference in rank sum	Significant? P < 0.05?	Summary
No peptide vs TMP1	-14,60	No	ns
No peptide vs TMP2	-12,45	No	ns
No peptide vs TMP3	-2,850	No	ns
No peptide vs Hypothetical protein	-1,350	No	ns
TMP1 vs TMP2	2,150	No	ns
TMP1 vs TMP3	11,75	No	ns
TMP1 vs Hypothetical protein	13,25	No	ns
TMP2 vs TMP3	9,600	No	ns
TMP2 vs Hypothetical protein	11,10	No	ns
TMP3 vs Hypothetical protein	1,500	No	ns

Table Analyzed Fig 1D – CTX + Eh13344

Kruskal-Wallis test			
P value	0,0149		
Exact or approximate P value?	Gaussian Approximation		
P value summary *			
Do the medians vary signif. (P < 0.05)	Yes		
Number of groups	5		
Kruskal-Wallis statistic	12,36		
Dunn's Multiple Comparison Test			
	Difference in rank sum	Significant? P < 0.05?	Summary
No peptide vs TMP1	-14,40	No	ns
No peptide vs TMP2	-7,300	No	ns
No peptide vs TMP3	0,3000	No	ns
No peptide vs Hypothetical protein	6,650	No	ns
TMP1 vs TMP2	7,100	No	ns

TMP1 vs TMP3	14,70	No	ns
TMP1 vs Hypothetical protein	21,05	Yes	*
TMP2 vs TMP3	7,600	No	ns
TMP2 vs Hypothetical protein	13,95	No	ns
TMP3 vs Hypothetical protein	6,350	No	ns

Table Analyzed Fig 1D – CTX + Eh708

Kruskal-Wallis test			
P value	0,1359		
Exact or approximate P value?	Gaussian Approximation		
P value summary	ns		
Do the medians vary signif. (P < 0.05)	No		
Number of groups	5		
Kruskal-Wallis statistic	6,999		
Dunn's Multiple Comparison Test			
	Difference in rank sum	Significant? P < 0.05?	Summary
No peptide vs TMP1	-19,43	No	ns
No peptide vs TMP2	-8,200	No	ns
No peptide vs TMP3	-3,700	No	ns
No peptide vs Hypothetical protein	-4,833	No	ns
TMP1 vs TMP2	11,23	No	ns
TMP1 vs TMP3	15,73	No	ns
TMP1 vs Hypothetical protein	14,60	No	ns
TMP2 vs TMP3	4,500	No	ns
TMP2 vs Hypothetical protein	3,367	No	ns
TMP3 vs Hypothetical protein	-1,133	No	ns

Fig.1E

Table Analyzed Fig 1E

Kruskal-Wallis test			
P value	P<0.0001		
Exact or approximate P value?	Gaussian Approximation		
P value summary	***		
Do the medians vary signif. (P < 0.05)	Yes		
Number of groups	6		
Kruskal-Wallis statistic	50,89		
Dunn's Multiple Comparison Test			
	Difference in rank sum	Significant? P < 0.05?	Summary
Ctrl vs CTX	-9.000	No	ns
Ctrl vs 13144	-51.54	Yes	***
Ctrl vs CTX+13344	-15.58	No	ns
Ctrl vs CTX+ATCC9790	-3.985	No	ns
Ctrl vs CTX+IGR11	-45.88	Yes	**
CTX vs 13144	-42.54	Yes	***
CTX vs CTX+13344	-6.585	No	ns
CTX vs CTX+ATCC9790	5.015	No	ns
CTX vs CTX+IGR11	-36.88	Yes	*
13144 vs CTX+13344	35.95	Yes	*
13144 vs CTX+ATCC9790	47.55	Yes	***
13144 vs CTX+IGR11	5.654	No	ns
CTX+13344 vs CTX+ATCC9790	11.60	No	ns
CTX+13344 vs CTX+IGR11	-30.30	No	ns
CTX+ATCC9790 vs CTX+IGR11	-41.90	Yes	*

---- Figure 2 ----

Fig.2B

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	5.73 (d.f.=5/95.9), p<0.0001	5.73 (d.f.=5/96.2), p<0.0001	26.63 (d.f.=5), p<<0.0001	28.63 (d.f.=5), p<<0.0001
Time	195.36 (d.f.=1/101.0), p<<0.0001	195.36 (d.f.=1/101.2), p<<0.0001	109.86 (d.f.=1), p<<0.0001	195.36 (d.f.=1), p<<0.0001
Treat	9.89 (d.f.=5/95.8), p<<0.0001	10.10 (d.f.=5/95.3), p<<0.0001	38.51 (d.f.=5), p<<0.0001	50.48 (d.f.=5), p<<0.0001

Selected pairwise comparisons

P-value ajustement: **no**

Largest	Smallest	Contrast	Df	Pvalue	PvalueAdj
PBS	BMDC	1.628 [-0.587;3.844]	96.48	0.1478	0.1478
PBS	Gr1pulsedBMDC	2.523 [0.142;4.903]	96.06	0.0380	0.0380
PBS	TMPpulsedBMDC	4.508 [2.452;6.564]	96.20	<0.0001	<0.0001
PBS	13144pulsedBMDC	4.044 [1.713;6.376]	96.07	0.0009	0.0009
TMP1-mut3pulsedBMDC	PBS	0.377 [-2.378;3.131]	95.95	0.7866	0.7866
BMDC	Gr1pulsedBMDC	0.894 [-1.586;3.375]	95.92	0.4759	0.4759
BMDC	TMPpulsedBMDC	2.880 [0.709;5.050]	96.01	0.0099	0.0099
BMDC	13144pulsedBMDC	2.416 [-0.018;4.849]	95.93	0.0516	0.0516
TMP1-mut3pulsedBMDC	BMDC	2.005 [-0.836;4.846]	95.86	0.1645	0.1645
Gr1pulsedBMDC	TMPpulsedBMDC	1.985 [-0.354;4.324]	95.64	0.0953	0.0953
Gr1pulsedBMDC	13144pulsedBMDC	1.522 [-1.063;4.106]	95.64	0.2454	0.2454
TMP1-mut3pulsedBMDC	Gr1pulsedBMDC	2.899 [-0.072;5.871]	95.64	0.0557	0.0557
13144pulsedBMDC	TMPpulsedBMDC	0.464 [-1.826;2.753]	95.64	0.6885	0.6885
TMP1-mut3pulsedBMDC	TMPpulsedBMDC	4.885 [2.166;7.603]	95.64	0.0006	0.0006
TMP1-mut3pulsedBMDC	13144pulsedBMDC	4.421 [1.488;7.353]	95.64	0.0035	0.0035

Fig.2C

Table Analyzed

Fig 2C

Kruskal-Wallis test

P value 0,0002

Exact or approximate P value? Gaussian Approximation

P value summary ***

Do the medians vary signif. (P < 0.05) Yes

Number of groups 6

Kruskal-Wallis statistic 23,96

Dunn's Multiple Comparison Test

	Difference in rank sum	Significant? P < 0.05?	Summary
PBS vs BMDC	4,875	No	ns
PBS vs Gr1 pulsed BMDC	18,49	No	ns
PBS vs TMP pulsed BMDC	25,84	Yes	*
PBS vs 13144 pulsed BMDC	30,21	Yes	*
PBS vs TMPmut pulsed BMDC	-3,347	No	ns
BMDC vs Gr1 pulsed BMDC	13,61	No	ns
BMDC vs TMP pulsed BMDC	20,96	No	ns
BMDC vs 13144 pulsed BMDC	25,33	No	ns
BMDC vs TMPmut pulsed BMDC	-8,222	No	ns
Gr1 pulsed BMDC vs TMP pulsed BMDC	7,354	No	ns
Gr1 pulsed BMDC vs 13144 pulsed BMDC	11,72	No	ns
Gr1 pulsed BMDC vs TMPmut pulsed BMDC	-21,83	No	ns
TMP pulsed BMDC vs 13144 pulsed BMDC	4,368	No	ns
TMP pulsed BMDC vs TMPmut pulsed BMDC	-29,19	Yes	*
13144 pulsed BMDC vs TMPmut pulsed BMDC	-33,56	Yes	*

Fig.2D

Table Analyzed	Fig 2D		
Kruskal-Wallis test			
P value		0,0003	
Exact or approximate P value?	Gaussian Approximation		
P value summary		***	
Do the medians vary signif. (P < 0.05)		Yes	
Number of groups		6	
Kruskal-Wallis statistic		23,57	
Dunn's Multiple Comparison Test	Difference in rank sum	Significant? P < 0.05?	Summary
CTX-spect vs CTX+13144	27,10	Yes	**
CTX-spect vs CTX+E.coli TMP-	10,20	No	ns
CTX-spect vs CTX+E.coli TMP+	23,43	Yes	*
CTX-spect vs CTX+E.coli TMPmut2	-0,6667	No	ns
CTX-spect vs CTX+E.coli TMPmut3	-4,667	No	ns
CTX+13144 vs CTX+E.coli TMP-	-16,90	No	ns
CTX+13144 vs CTX+E.coli TMP+	-3,667	No	ns
CTX+13144 vs CTX+E.coli TMPmut2	-27,77	No	ns
CTX+13144 vs CTX+E.coli TMPmut3	-31,77	Yes	*
CTX+E.coli TMP- vs CTX+E.coli TMP+	13,23	No	ns
CTX+E.coli TMP- vs CTX+E.coli TMPmut2	-10,87	No	ns
CTX+E.coli TMP- vs CTX+E.coli TMPmut3	-14,87	No	ns
CTX+E.coli TMP+ vs CTX+E.coli TMPmut2	-24,10	No	ns
CTX+E.coli TMP+ vs CTX+E.coli TMPmut3	-28,10	No	ns
CTX+E.coli TMPmut2 vs CTX+E.coli TMPmut3	-4,000	No	ns

Fig.2E

Table Analyzed	Fig 2E		
Kruskal-Wallis test			
P value		0,0003	
Exact or approximate P value?	Gaussian Approximation		
P value summary		***	
Do the medians vary signif. (P < 0.05)		Yes	
Number of groups		6	
Kruskal-Wallis statistic		23,68	
Dunn's Multiple Comparison Test	Difference in rank sum	Significant? P < 0.05?	Summary
CTX-spect vs CTX+13144	-19,60	Yes	*
CTX-spect vs CTX+E.coli TMP-	-8,450	No	ns
CTX-spect vs CTX+E.coli TMP+	-18,85	No	ns
CTX-spect vs CTX+E.coli TMPmut2	8,100	No	ns
CTX-spect vs CTX+E.coli TMPmut3	2,700	No	ns
CTX+13144 vs CTX+E.coli TMP-	11,15	No	ns
CTX+13144 vs CTX+E.coli TMP+	0,7500	No	ns
CTX+13144 vs CTX+E.coli TMPmut2	27,70	Yes	**
CTX+13144 vs CTX+E.coli TMPmut3	22,30	No	ns
CTX+E.coli TMP- vs CTX+E.coli TMP+	-10,40	No	ns
CTX+E.coli TMP- vs CTX+E.coli TMPmut2	16,55	No	ns
CTX+E.coli TMP- vs CTX+E.coli TMPmut3	11,15	No	ns
CTX+E.coli TMP+ vs CTX+E.coli TMPmut2	26,95	Yes	*
CTX+E.coli TMP+ vs CTX+E.coli TMPmut3	21,55	No	ns
CTX+E.coli TMPmut2 vs CTX+E.coli TMPmut3	-5,400	No	ns

---- Figure 3 ----

Fig.3B

Table Analyzed		Fig 3B		
Kruskal-Wallis test				
P value		0,0013		
Exact or approximate P value?		Gaussian Approximation		
P value summary		**		
Do the medians vary signif. (P < 0.05)		Yes		
Number of groups		4		
Kruskal-Wallis statistic		15,68		
Dunn's Multiple Comparison Test		Difference in rank sum	Significant? P < 0,05?	Summary
Ctrl vs MCA205		-8,333	No	ns
Ctrl vs TC1		-12,11	No	ns
Ctrl vs MC38		5,500	No	ns
MCA205 vs TC1		-3,778	No	ns
MCA205 vs MC38		13,83	Yes	*
TC1 vs MC38		17,61	Yes	**

Fig.3C (left panel)

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	95.65 (d.f.=2/21.6), p<<0.0001	95.94 (d.f.=2/22.2), p<<0.0001	51.22 (d.f.=2), p<<0.0001	191.88 (d.f.=2), p<<0.0001
Time	22.86 (d.f.=1/17.5), p<0.0002	22.86 (d.f.=1/16.6), p<0.0002	15.41 (d.f.=1), p<<0.0001	22.86 (d.f.=1), p<<0.0001
Treat	93.37 (d.f.=2/14.9), p<<0.0001	94.54 (d.f.=2/13.1), p<<0.0001	48.63 (d.f.=2), p<<0.0001	189.08 (d.f.=2), p<<0.0001

Selected pairwise comparisons

P-value adjustment: **no**

Largest	Smallest	Contrast	Df	Pvalue	PvalueAdj
WTclone1Ctrl	WTclone1CTX	12.073 [9.785;14.360]	22.53	<0.0001	<0.0001
WTclone1Ctrl	WTclone1EH	14.339 [12.052;16.627]	22.53	<0.0001	<0.0001
WTclone1CTX	WTclone1EH	2.266 [0.034;4.499]	20.08	0.0470	0.0470

Fig.3C (middle panel)

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	5.54 (d.f.=2/15.0), p<0.0158	5.54 (d.f.=2/15.0), p<0.0157	9.93 (d.f.=2), p<0.0070	11.09 (d.f.=2), p<0.0039
Time	48.61 (d.f.=1/17.0), p<<0.0001	48.62 (d.f.=1/16.9), p<<0.0001	24.24 (d.f.=1), p<<0.0001	48.62 (d.f.=1), p<<0.0001
Treat	9.89 (d.f.=2/14.8), p<0.0019	11.24 (d.f.=2/65.0), p<<0.0001	14.44 (d.f.=2), p<0.0007	22.48 (d.f.=2), p<<0.0001

Selected pairwise comparisons

P-value adjustment: **no**

Largest	Smallest	Contrast	Df	Pvalue	PvalueAdj
Mut2Clone1Ctrl	Mut2Clone1CTX	6.317 [2.194;10.440]	15.06	0.0052	0.0052
Mut2Clone1Ctrl	Mut2Clone1EH	4.265 [0.142;8.388]	15.06	0.0435	0.0435
Mut2Clone1EH	Mut2Clone1CTX	2.052 [-2.062;6.167]	14.89	0.3044	0.3044

Fig.3C (right panel)

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	2.77 (d.f.=2/15.0), p<0.0949	2.77 (d.f.=2/14.9), p<0.0952	5.64 (d.f.=2), p<0.0596	5.53 (d.f.=2), p<0.0629
Time	27.10 (d.f.=1/17.0), p<<0.0001	27.10 (d.f.=1/16.8), p<<0.0001	17.11 (d.f.=1), p<<0.0001	27.10 (d.f.=1), p<<0.0001
Treat	4.75 (d.f.=2/15.0), p<0.0252	5.38 (d.f.=2/102.0), p<0.0060	7.42 (d.f.=2), p<0.0245	10.76 (d.f.=2), p<0.0046

Selected pairwise comparisons

P-value adjustment: **no**

Largest	Smallest	Contrast	Df	Pvalue	PvalueAdj
Mut3Clone1Ctrl	Mut3Clone1CTX	4.589 [0.245;8.934]	15.02	0.0398	0.0398
Mut3Clone1Ctrl	Mut3Clone1EH	3.499 [-0.846;7.843]	15.00	0.1066	0.1066
Mut3Clone1EH	Mut3Clone1CTX	1.091 [-3.252;5.434]	14.98	0.6003	0.6003

Fig.3D

Table Analyzed Fig 3D – WT Polyclonal

Column A CTX
vs
Column B CTX + 13144

Mann Whitney test

P value 0,0119

Exact or approximate P value? Gaussian Approximation

P value summary *

Are medians signif. different? (P < 0.05) Yes

One- or two-tailed P value? Two-tailed

Sum of ranks in column A,B 40 , 15

Mann-Whitney U 0,0000

Table Analyzed Fig 3D – WT Clone 1

Column C CTX
vs
Column D CTX + 13144

Mann Whitney test

P value 0,0247

Exact or approximate P value? Gaussian Approximation

P value summary *

Are medians signif. different? (P < 0.05) Yes

One- or two-tailed P value? Two-tailed

Sum of ranks in column C,D 53.50 , 24.50

Mann-Whitney U 3,500

Table Analyzed Fig 3D – WT Clone 2

Column E CTX
vs
Column F CTX + 13144

Mann Whitney test

P value 0,0823

Exact or approximate P value? Exact

P value summary ns

Are medians signif. different? (P < 0.05) No

One- or two-tailed P value? Two-tailed

Sum of ranks in column E,F 46 , 20

Mann-Whitney U 5,000

Table Analyzed Fig 3D – mut2 Clone 1

Column M CTX
vs
Column N CTX + 13144

Mann Whitney test

P value 0,3939

Exact or approximate P value?	Exact
P value summary	ns
Are medians signif. different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column M,N	33 , 45
Mann-Whitney U	12,00

Table Analyzed **Fig 3D – mut2 Clone 2**

Column O	CTX
vs	vs
Column P	CTX + 13144

Mann Whitney test	
P value	0,3095
Exact or approximate P value?	Exact
P value summary	ns
Are medians signif. different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column O,P	32 , 46
Mann-Whitney U	11,00

Table Analyzed **Fig 3D – mut3 Clone 1**

Column G	CTX
vs	vs
Column H	CTX + 13144

Mann Whitney test	
P value	0,6884
Exact or approximate P value?	Gaussian Approximation
P value summary	ns
Are medians signif. different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column G,H	36 , 42
Mann-Whitney U	15,00

Table Analyzed **Fig 3D – mut3 Clone 2**

Column I	CTX
vs	vs
Column J	CTX + 13144

Mann Whitney test	
P value	0,9360
Exact or approximate P value?	Gaussian Approximation
P value summary	ns
Are medians signif. different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column I,J	40 , 38
Mann-Whitney U	17,00

Table Analyzed **Fig 3D – mut3 Clone 3**

Column K	CTX
vs	vs
Column L	CTX + 13144

Mann Whitney test	
P value	0,3358
Exact or approximate P value?	Gaussian Approximation
P value summary	ns
Are medians signif. different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column K,L	45.50 , 32.50
Mann-Whitney U	11,50

Fig.3E (left panel)

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	6.35 (d.f.=2/26.3), p<0.0056	6.35 (d.f.=2/26.1), p<0.0057	11.50 (d.f.=2), p<0.0032	12.69 (d.f.=2), p<0.0018
Time	80.88 (d.f.=1/28.0), p<<0.0001	80.88 (d.f.=1/27.8), p<<0.0001	39.41 (d.f.=1), p<<0.0001	80.88 (d.f.=1), p<<0.0001
Treat	0.88 (d.f.=2/26.2), p<0.4248	0.95 (d.f.=2/144.3), p<0.3882	1.93 (d.f.=2), p<0.3801	1.91 (d.f.=2), p<0.3858

Selected pairwise comparisons

P-value adjustment: **no**

Largest	Smallest	Contrast	Df	Pvalue	PvalueAdj
TC1WTPBS	TC1WTCTX	2.291 [-1.256;5.838]	26.98	0.1962	0.1962
TC1WTPBS	TC1WT13144	5.866 [2.389;9.344]	27.09	0.0018	0.0018
TC1WTCTX	TC1WT13144	3.575 [0.352;6.799]	25.22	0.0311	0.0311

Fig.3E (middle panel)

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	1.54 (d.f.=2/27.9), p<0.2324	1.54 (d.f.=2/28.1), p<0.2322	3.21 (d.f.=2), p<0.2006	3.08 (d.f.=2), p<0.2147
Time	126.95 (d.f.=1/30.0), p<<0.0001	126.99 (d.f.=1/30.1), p<<0.0001	51.38 (d.f.=1), p<<0.0001	126.99 (d.f.=1), p<<0.0001
Treat	0.19 (d.f.=2/27.9), p<0.8319	0.20 (d.f.=2/120.8), p<0.8200	0.34 (d.f.=2), p<0.8449	0.40 (d.f.=2), p<0.8197

Selected pairwise comparisons

P-value adjustment: **no**

Largest	Smallest	Contrast	Df	Pvalue	PvalueAdj
TC1mut3PBS	TC1mut3CTX	0.833 [-1.969;3.635]	28.54	0.5478	0.5478
TC1mut3PBS	TC1mut313144	2.369 [-0.432;5.170]	28.48	0.0942	0.0942
TC1mut3CTX	TC1mut313144	1.536 [-1.294;4.367]	26.83	0.2751	0.2751

Fig.3E (right panel)

Table Analyzed Fig 3E right – TC1 WT

Mann Whitney test	
P value	0,0229
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,B	142,0 , 89,00
Mann-Whitney U	23,00
Difference between medians	
Median of column A	195,0
Median of column B	91,00
Difference: Actual	-104,0
Difference: Hodges-Lehmann	-65,00

Table Analyzed Fig 3E right - TC1 mut3

Mann Whitney test	
P value	0,4467
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column C,D	115,5 , 94,50
Mann-Whitney U	39,50

Difference between medians	
Median of column C	187,5
Median of column D	189,0
Difference: Actual	1,500
Difference: Hodges-Lehmann	-38,50

---- Figure 4 ----

Fig.4A (left panel)

Table Analyzed		Fig 4A		
Kruskal-Wallis test				
P value		P<0.0001		
Exact or approximate P value?		Gaussian Approximation		
P value summary		***		
Do the medians vary signif. (P < 0.05)		Yes		
Number of groups		3		
Kruskal-Wallis statistic		23,42		
Dunn's Multiple Comparison Test				
	Difference in rank sum	Significant? P < 0.05?	Summary	
Ctrl vs CTX	-12,12	No	ns	
Ctrl vs CTX + 13144	-24,48	Yes	***	
CTX vs CTX + 13144	-12,36	Yes	*	

Fig.4A (middle panel)

Table Analyzed		Fig 4A		
Kruskal-Wallis test				
P value		0,0015		
Exact or approximate P value?		Gaussian Approximation		
P value summary		**		
Do the medians vary signif. (P < 0.05)		Yes		
Number of groups		3		
Kruskal-Wallis statistic		13,02		
Dunn's Multiple Comparison Test				
	Difference in rank sum	Significant? P < 0.05?	Summary	
Ctrl vs CTX	-11,17	No	ns	
Ctrl vs CTX + 13144	-18,27	Yes	***	
CTX vs CTX + 13144	-7,100	No	ns	

Fig.4A (right panel)

Table Analyzed		Fig 4A		
Kruskal-Wallis test				
P value		0,0007		
Exact or approximate P value?		Gaussian Approximation		
P value summary		***		
Do the medians vary signif. (P < 0.05)		Yes		
Number of groups		3		
Kruskal-Wallis statistic		14,51		
Dunn's Multiple Comparison Test				
	Difference in rank sum	Significant? P < 0.05?	Summary	
Ctrl vs CTX	-6,652	No	ns	
Ctrl vs CTX + 13144	-18,73	Yes	***	
CTX vs CTX + 13144	-12,08	Yes	*	

Fig.4C

Table Analyzed		Figure 4F – left panel		
Kruskal-Wallis test				
P value		0,0019		
Exact or approximate P value?		Gaussian Approximation		
P value summary		**		
Do the medians vary signif. (P < 0.05)		Yes		
Number of groups		5		
Kruskal-Wallis statistic		17,04		

Table Analyzed		Figure 4C		
Column B		TMP1		
vs.		vs,		
Column D		SIINFELK		
Mann Whitney test				

P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column B,D	26,00 , 10,00
Mann-Whitney U	0,0
Difference between medians	
Median of column B	105,5
Median of column D	14,00
Difference: Actual	91,50
Difference: Hodges-Lehmann	90,00

Table Analyzed	Figure 4C
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Column C	PSMB4
vs.	vs,
Column D	SIINFEKL
Mann Whitney test	
P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column C,D	26,00 , 10,00
Mann-Whitney U	0,0
Difference between medians	
Median of column C	35,00
Median of column D	14,00
Difference: Actual	21,00
Difference: Hodges-Lehmann	21,00

Table Analyzed	Figure 4C
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Column D	SIINFEKL
vs.	vs,
Column E	13144
Mann Whitney test	
P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column D,E	10,00 , 26,00
Mann-Whitney U	0,0
Difference between medians	
Median of column D	14,00
Median of column E	38,50
Difference: Actual	-24,50
Difference: Hodges-Lehmann	-24,50

---- Figure S2 ----

Fig.S2A

Table Analyzed

Fig S2A - DC

Kruskal-Wallis test				
P value		0,0464		
Exact or approximate P value?	Gaussian Approximation			
P value summary		*		
Do the medians vary signif. (P < 0.05)		Yes		
Number of groups		4		
Kruskal-Wallis statistic		7,981		
Dunn's Multiple Comparison Test				
	Difference in rank sum		Significant? P < 0.05?	Summary
Naïves vs CTX	-8,138		No	ns
Naïves vs CTX + 708	-13,60		No	ns
Naïves vs CTX + 13144	-19,65		Yes	*
CTX vs CTX + 708	-5,462		No	ns
CTX vs CTX + 13144	-11,51		No	ns
CTX + 708 vs CTX + 13144	-6,050		No	ns

Table Analyzed

Fig S2A – DC + Eh13144

Kruskal-Wallis test				
P value		P<0.0001		
Exact or approximate P value?	Gaussian Approximation			
P value summary		***		
Do the medians vary signif. (P < 0.05)		Yes		
Number of groups		4		
Kruskal-Wallis statistic		34,98		
Dunn's Multiple Comparison Test				
	Difference in rank sum		Significant? P < 0.05?	Summary
Naïves vs CTX	-6,575		No	ns
Naïves vs CTX + 708	-30,68		Yes	***
Naïves vs CTX + 13144	-36,15		Yes	***
CTX vs CTX + 708	-24,10		Yes	**
CTX vs CTX + 13144	-29,58		Yes	***
CTX + 708 vs CTX + 13144	-5,475		No	ns

Table Analyzed

Fig S2A –DC + Eh708

Kruskal-Wallis test				
P value		0,0148		
Exact or approximate P value?	Gaussian Approximation			
P value summary		*		
Do the medians vary signif. (P < 0.05)		Yes		
Number of groups		4		
Kruskal-Wallis statistic		10,49		
Dunn's Multiple Comparison Test				
	Difference in rank sum		Significant? P < 0.05?	Summary
Naïves vs CTX	-2,200		No	ns
Naïves vs CTX + 708	-13,58		No	ns
Naïves vs CTX + 13144	-20,63		Yes	*
CTX vs CTX + 708	-11,38		No	ns
CTX vs CTX + 13144	-18,43		No	ns
CTX + 708 vs CTX + 13144	-7,050		No	ns

Table Analyzed

Fig S2A – DC + Eh13344

Kruskal-Wallis test				
P value		0,0987		
Exact or approximate P value?	Gaussian Approximation			
P value summary		ns		
Do the medians vary signif. (P < 0.05)		No		
Number of groups		4		
Kruskal-Wallis statistic		6,282		

Dunn's Multiple Comparison Test	Difference in rank sum	Significant? P < 0.05?	Summary
Naïves vs CTX	-10,75	No	ns
Naïves vs CTX + 708	-11,48	No	ns
Naïves vs CTX + 13144	-18,18	No	ns
CTX vs CTX + 708	-0,7250	No	ns
CTX vs CTX + 13144	-7,425	No	ns
CTX + 708 vs CTX + 13144	-6,700	No	ns

Table Analyzed Fig S2A - DC + L.plant

Kruskal-Wallis test	
P value	0,0443
Exact or approximate P value?	Gaussian Approximation
P value summary	*
Do the medians vary signif. (P < 0.05)	Yes
Number of groups	4
Kruskal-Wallis statistic	8,082

Dunn's Multiple Comparison Test	Difference in rank sum	Significant? P < 0.05?	Summary
Naïves vs CTX	-6,200	No	ns
Naïves vs CTX + 708	13,90	No	ns
Naïves vs CTX + 13144	-0,3000	No	ns
CTX vs CTX + 708	20,10	Yes	*
CTX vs CTX + 13144	5,900	No	ns
CTX + 708 vs CTX + 13144	-14,20	No	ns

Fig.S2C (left panel)

Table Analyzed Fig S2C – left panel

Kruskal-Wallis test	
P value	0,0032
Exact or approximate P value?	Approximate
P value summary	**
Do the medians vary signif. (P < 0.05)	Yes

Dunn's multiple comparisons test	Mean rank diff,	Significant?	Summary
Ctrl vs. CTX	-2,200	No	ns
Ctrl vs. 13144	-12,50	Yes	**
CTX vs. 13144	-10,30	Yes	*

Fig.S2C (right panel)

Table Analyzed Fig S2C – D11

Kruskal-Wallis test	
P value	0,1939
Exact or approximate P value?	Exact
P value summary	ns
Do the medians vary signif. (P < 0.05)	No

Dunn's multiple comparisons test	Mean rank diff,	Significant?	Summary
Column A vs. Column B	-4,600	No	ns
Column A vs. Column C	-0,2000	No	ns
Column B vs. Column C	4,400	No	ns

Table Analyzed Fig S2C – D18

Kruskal-Wallis test	
P value	0,4163
Exact or approximate P value?	Exact
P value summary	ns
Do the medians vary signif. (P < 0.05)	No

Dunn's multiple comparisons test	Mean rank diff,	Significant?	Summary
Column E vs. Column F	-0,6000	No	ns
Column E vs. Column G	-3,600	No	ns
Column F vs. Column G	-3,000	No	ns

Table Analyzed		Fig S2C – D25		
Kruskal-Wallis test				
P value	0,0130			
Exact or approximate P value?	Exact			
P value summary	*			
Do the medians vary signif. (P < 0.05)	Yes			
Dunn's multiple comparisons test				
	Mean rank diff,	Significant?	Summary	
Column I vs. Column J	4,200	No	ns	
Column I vs. Column K	-3,600	No	ns	
Column J vs. Column K	-7,800	Yes	*	

Fig.S2D

Table Analyzed		Fig S2D – D11		
Kruskal-Wallis test				
P value	0,8261			
Exact or approximate P value?	Gaussian Approximation			
P value summary	ns			
Do the medians vary signif. (P < 0.05)	No			
Number of groups	3			
Kruskal-Wallis statistic	0,3820			
Dunn's Multiple Comparison Test				
	Difference in rank sum	Significant? P < 0.05?	Summary	
Ctrl vs CTX	0,2000	No	ns	
Ctrl vs 13144	-1,400	No	ns	
CTX vs 13144	-1,600	No	ns	

Table Analyzed		Fig S2D – D18		
Kruskal-Wallis test				
P value	0,0131			
Exact or approximate P value?	Gaussian Approximation			
P value summary	*			
Do the medians vary signif. (P < 0.05)	Yes			
Number of groups	3			
Kruskal-Wallis statistic	8,676			
Dunn's Multiple Comparison Test				
	Difference in rank sum	Significant? P < 0.05?	Summary	
Ctrl vs CTX	1,700	No	ns	
Ctrl vs 13144	-6,200	No	ns	
CTX vs 13144	-7,900	Yes	*	

Table Analyzed		Fig S2D – D25		
Kruskal-Wallis test				
P value	0,0657			
Exact or approximate P value?	Gaussian Approximation			
P value summary	ns			
Do the medians vary signif. (P < 0.05)	No			
Number of groups	3			
Kruskal-Wallis statistic	5,445			
Dunn's Multiple Comparison Test				
	Difference in rank sum	Significant? P < 0.05?	Summary	
Ctrl vs CTX	0,4000	No	ns	
Ctrl vs 13144	-5,500	No	ns	
CTX vs 13144	-5,900	No	ns	

Fig.S2F

Table Analyzed		Fig S2F – D10		
Kruskal-Wallis test				
P value	0,0177			
Exact or approximate P value?	Exact			
P value summary	*			
Do the medians vary signif. (P < 0.05)	Yes			

Number of groups 3
 Kruskal-Wallis statistic 7,246

Dunn's multiple comparisons test	Mean rank diff,	Significant?	Summary
Ctrl vs. CTX	5,200	No	ns
Ctrl vs. CTX+13144	7,400	Yes	*
CTX vs. CTX+13144	2,200	No	ns

Fig.S2G

Table Analyzed	Fig S2G – D14
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Kruskal-Wallis test	
P value	0,0298
Exact or approximate P value?	Exact
P value summary	*
Do the medians vary signif. (P < 0.05)	Yes
Number of groups	3
Kruskal-Wallis statistic	6,503

Dunn's multiple comparisons test	Mean rank diff,	Significant?	Summary
Ctrl vs. CTX	3,600	No	ns
Ctrl vs. CTX+13144	7,200	Yes	*
CTX vs. CTX+13144	3,600	No	ns

Fig.S2H

Table Analyzed	Fig S2H – D17
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ANOVA summary	
F	12,86
P value	0,0010
P value summary	**
Are differences among means statistically significant? (P < 0.05)	Yes
R square	0,6819

Dunnett's multiple comparisons test	Mean Diff,	95% CI of diff,	Significant?	Summary
Ctrl vs. CTX+13144	1,740	0,8359 to 2,644	Yes	***
CTX vs. CTX+13144	1,368	0,4639 to 2,272	Yes	**

---- Figure S4 ----

Fig.S4B

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	5.18 (d.f.=5/64.0), p<0.0005	5.18 (d.f.=5/64.0), p<0.0005	23.80 (d.f.=5), p<0.0002	25.92 (d.f.=5), p<<0.0001
Time	413.68 (d.f.=1/69.0), p<<0.0001	413.68 (d.f.=1/69.0), p<<0.0001	136.16 (d.f.=1), p<<0.0001	413.68 (d.f.=1), p<<0.0001
Treat	1.67 (d.f.=5/64.0), p<0.1545	1.72 (d.f.=5/64.0), p<0.1421	8.35 (d.f.=5), p<0.1382	8.61 (d.f.=5), p<0.1255

Selected pairwise comparisons

P-value adjustement: **no**

Largest	Smallest	Contrast	Df	Pvalue	PvalueAdj
CTX+S	13144	2.973 [-1.302;4.644]	64.01	0.0007	0.0007
CTX+S	E.coli-EGFP	1.220 [-0.451;2.892]	64.01	0.1495	0.1495
CTX+S	E.coli-TMP	2.961 [-1.289;4.632]	64.01	0.0008	0.0008
E.coli-TMPmut2	CTX+S	0.373 [-1.990;2.737]	64.00	0.7535	0.7535
E.coli-TMPmut3	CTX+S	0.455 [-1.909;2.818]	64.00	0.7019	0.7019
E.coli-EGFP	13144	1.753 [0.081;3.424]	63.99	0.0401	0.0401
E.coli-TMP	13144	0.012 [-1.659;1.684]	63.99	0.9882	0.9882
E.coli-TMPmut2	13144	3.346 [0.983;5.710]	63.99	0.0062	0.0062
E.coli-TMPmut3	13144	3.428 [1.064;5.791]	63.99	0.0051	0.0051
E.coli-EGFP	E.coli-TMP	1.740 [0.069;3.411]	63.99	0.0415	0.0415
E.coli-TMPmut2	E.coli-EGFP	1.594 [-0.770;3.957]	63.99	0.1827	0.1827
E.coli-TMPmut3	E.coli-EGFP	1.675 [-0.688;4.039]	63.99	0.1616	0.1616
E.coli-TMPmut2	E.coli-TMP	3.334 [0.970;5.697]	63.99	0.0064	0.0064
E.coli-TMPmut3	E.coli-TMP	3.415 [1.052;5.779]	63.99	0.0053	0.0053
E.coli-TMPmut3	E.coli-TMPmut2	0.082 [-2.813;2.976]	63.99	0.9552	0.9552

---- Figure S6 ----

Fig.S6A

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	78.75 (d.f.=2/27.1), p<<0.0001	78.99 (d.f.=2/26.8), p<<0.0001	59.80 (d.f.=2), p<<0.0001	157.98 (d.f.=2), p<<0.0001
Time	12.62 (d.f.=1/29.0), p<0.0013	12.62 (d.f.=1/28.2), p<0.0014	10.91 (d.f.=1), p<0.0010	12.62 (d.f.=1), p<0.0004
Treat	20.60 (d.f.=2/27.0), p<<0.0001	22.09 (d.f.=2/27.6), p<<0.0001	6.14 (d.f.=2), p<0.0464	44.19 (d.f.=2), p<<0.0001

Selected pairwise comparisons

P-value adjustment: **no**

Largest	Smallest	Contrast	Df	Pvalue	PvalueAdj
Ctrl	CTX	16.277 [12.721;19.834]	28.52	<0.0001	<0.0001
Ctrl	CTX+13144	20.995 [17.439;24.552]	28.52	<0.0001	<0.0001
CTX	CTX+13144	4.718 [1.277;8.159]	24.84	0.0092	0.0092

Fig.S6B

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	11.94 (d.f.=2/25.9), p<0.0002	11.97 (d.f.=2/26.0), p<0.0002	19.34 (d.f.=2), p<<0.0001	23.94 (d.f.=2), p<<0.0001
Time	31.62 (d.f.=1/27.9), p<<0.0001	31.64 (d.f.=1/27.2), p<<0.0001	22.08 (d.f.=1), p<<0.0001	31.64 (d.f.=1), p<<0.0001
Treat	15.25 (d.f.=2/26.0), p<<0.0001	16.42 (d.f.=2/24.4), p<<0.0001	13.97 (d.f.=2), p<0.0009	32.83 (d.f.=2), p<<0.0001

Selected pairwise comparisons

P-value adjustment: **no**

Largest	Smallest	Contrast	Df	Pvalue	PvalueAdj
Ctrl	CTX	6.951 [3.078;10.824]	26.33	0.0010	0.0010
Ctrl	CTX+13144	8.849 [4.976;12.722]	26.33	<0.0001	<0.0001
CTX	CTX+13144	1.898 [-1.814;5.611]	25.19	0.3025	0.3025

Fig.S6C

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	54.75 (d.f.=1/46.0), p<<0.0001	54.76 (d.f.=1/44.0), p<<0.0001	37.54 (d.f.=1), p<<0.0001	54.76 (d.f.=1), p<<0.0001
Time	129.98 (d.f.=1/47.0), p<<0.0001	129.99 (d.f.=1/46.1), p<<0.0001	63.50 (d.f.=1), p<<0.0001	129.99 (d.f.=1), p<<0.0001
Treat	0.35 (d.f.=1/44.7), p<0.5588	0.36 (d.f.=1/216.6), p<0.5473	0.15 (d.f.=1), p<0.6999	0.36 (d.f.=1), p<0.5467

Selected pairwise comparisons

P-value adjustment: **no**

Comp to MCA205WT	Contrast	Df	Pvalue	PvalueAdj
MCA205mut2	-7.321 [-9.313;-5.329]	45.98	<0.0001	<0.0001

Fig.S6D

Longitudinal analysis

	F test (KR)	F test (S)	LR test	Wald test
Time x Treat	53.16 (d.f.=1/23.0), p<<0.0001	53.16 (d.f.=1/24.3), p<<0.0001	30.07 (d.f.=1), p<<0.0001	53.16 (d.f.=1), p<<0.0001
Time	43.59 (d.f.=1/24.0), p<<0.0001	43.59 (d.f.=1/24.1), p<<0.0001	25.87 (d.f.=1), p<<0.0001	43.59 (d.f.=1), p<<0.0001
Treat	25.21 (d.f.=1/23.0), p<<0.0001	27.40 (d.f.=1/66.8), p<<0.0001	8.16 (d.f.=1), p<0.0043	27.40 (d.f.=1), p<<0.0001

Selected pairwise comparisons

P-value adjustment: **no**

Comp to TC1WT	Contrast	Df	Pvalue	PvalueAdj
TC1mut2	-10.327 [-13.257;-7.397]	23.00	<0.0001	<0.0001

Fig.S6E

Table Analyzed Fig S6E – MCA205 WT

Mann Whitney test	
P value	0,0065
Exact or approximate P value?	Exact
P value summary	**
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,B	55,50 , 22,50
Mann-Whitney U	1,500
Difference between medians	
Median of column A	80,00
Median of column B	54,00
Difference: Actual	-26,00
Difference: Hodges-Lehmann	-26,50

Table Analyzed Fig S6E – MCA205 mut3

Mann Whitney test	
P value	0,9805
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column C,D	39,50 , 38,50
Mann-Whitney U	17,50
Difference between medians	
Median of column C	65,00
Median of column D	58,00
Difference: Actual	-7,000
Difference: Hodges-Lehmann	-2,000

Fig.S6G (upper panel)

Table Analyzed Figure 6G – upper panel

Kruskal-Wallis test	
P value	0,0014
Exact or approximate P value?	Approximate
P value summary	**
Do the medians vary signif. (P < 0.05)	Yes
Number of groups	5
Kruskal-Wallis statistic	17,65
Data summary	
Number of treatments (columns)	5

Table Analyzed	Figure 6G – upper panel
Column G	Ctrl
vs.	vs,
Column H	TMP1
Mann Whitney test	
P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column G,H	10,00 , 26,00
Mann-Whitney U	0,0
Difference between medians	
Median of column G	1,000
Median of column H	5,481
Difference: Actual	-4,481
Difference: Hodges-Lehmann	-4,481

Table Analyzed	Figure 6G – upper panel
Column G	Ctrl
vs.	vs,
Column I	PSMB4
Mann Whitney test	
P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column G,I	10,00 , 26,00
Mann-Whitney U	0,0
Difference between medians	
Median of column G	1,000
Median of column I	1,818
Difference: Actual	-0,8182
Difference: Hodges-Lehmann	-0,8182

Table Analyzed	Figure 6G – upper panel
Column G	Ctrl
vs.	vs,
Column K	13144
Mann Whitney test	
P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column G,K	10,00 , 26,00
Mann-Whitney U	0,0
Difference between medians	
Median of column G	1,000
Median of column K	2,000
Difference: Actual	-1,000
Difference: Hodges-Lehmann	-1,000

Table Analyzed	Figure 6G – upper panel
Column H	TMP1
vs.	vs,
Column J	SIINFEKL
Mann Whitney test	
P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column H,J	26,00 , 10,00

Mann-Whitney U	0,0
Difference between medians	
Median of column H	5,481
Median of column J	0,7273
Difference: Actual	4,753
Difference: Hodges-Lehmann	4,675

Table Analyzed	Figure 6G – upper panel
Column I	PSMB4
vs.	vs,
Column J	SIINFEKL
Mann Whitney test	
P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column I,J	26,00 , 10,00
Mann-Whitney U	0,0
Difference between medians	
Median of column I	1,818
Median of column J	0,7273
Difference: Actual	1,091
Difference: Hodges-Lehmann	1,091

Table Analyzed	Figure 6G – upper panel
Column J	SIINFEKL
vs.	vs,
Column K	13144
Mann Whitney test	
P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column J,K	10,00 , 26,00
Mann-Whitney U	0,0
Difference between medians	
Median of column J	0,7273
Median of column K	2,000
Difference: Actual	-1,273
Difference: Hodges-Lehmann	-1,273

Fig.S6G (lower panel)

Table Analyzed	Fig S6G - lower panel
Kruskal-Wallis test	
P value	0,0005
Exact or approximate P value?	Exact
P value summary	***
Do the medians vary signif. (P < 0.05)	Yes
Number of groups	3
Kruskal-Wallis statistic	9,269
Data summary	
Number of treatments (columns)	3
Number of values (total)	12

Table Analyzed	Fig S6G - lower panel
Column B	PSMB4
vs.	vs,
Column A	SIINFEKL
Mann Whitney test	
P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes

One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,B	10,00 , 26,00
Mann-Whitney U	0,0
Difference between medians	
Median of column A	154,0
Median of column B	246,5
Difference: Actual	92,50
Difference: Hodges-Lehmann	104,0

Table Analyzed	Fig S6G - lower panel
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Column C	TMP1
vs.	vs,
Column A	SIINFEKL
Mann Whitney test	
P value	0,0286
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,C	10,00 , 26,00
Mann-Whitney U	0,0
Difference between medians	
Median of column A	154,0
Median of column C	176,0
Difference: Actual	22,00
Difference: Hodges-Lehmann	23,00

Table Analyzed	Fig S6G - lower panel
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Column C	TMP1
vs.	vs,
Column B	PSMB4
Mann Whitney test	
P value	0,0571
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column B,C	25,00 , 11,00
Mann-Whitney U	1,000
Difference between medians	
Median of column B	246,5
Median of column C	176,0
Difference: Actual	-70,50
Difference: Hodges-Lehmann	-66,00

---- Figure S11 ----

Fig.S11A

Table Analyzed	Fig S11A – Peptide 1
Column A	No peptide
vs.	vs,
Column G	1
Mann Whitney test	
P value	0,1385
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,G	124,0 , 176,0
Mann-Whitney U	46,00
Difference between medians	
Median of column A	5,500
Median of column G	13,50
Difference: Actual	-8,000
Difference: Hodges-Lehmann	-6,000

Table Analyzed	Fig S11A – Peptide 2
Column A	No peptide
vs.	vs,
Column C	2
Mann Whitney test	
P value	0,0294
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,C	112,5 , 187,5
Mann-Whitney U	34,50
Difference between medians	
Median of column A	5,500
Median of column C	15,50
Difference: Actual	-10,00
Difference: Hodges-Lehmann	-8,500

Table Analyzed	Fig S11A – Peptide 3
Column A	No peptide
vs.	vs,
Column D	3
Mann Whitney test	
P value	0,0229
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,D	111,0 , 189,0
Mann-Whitney U	33,00
Difference between medians	
Median of column A	5,500
Median of column D	18,00
Difference: Actual	-12,50
Difference: Hodges-Lehmann	-10,50

Table Analyzed	Fig S11A – Peptide 4
Column A	No peptide
vs.	vs,
Column E	4
Mann Whitney test	
P value	0,1081

Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,E	122,0 , 178,0
Mann-Whitney U	44,00
Difference between medians	
Median of column A	5,500
Median of column E	9,500
Difference: Actual	-4,000
Difference: Hodges-Lehmann	-4,000

Table Analyzed	Fig S11A – Peptide 5
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Column A	No peptide
vs.	vs,
Column F	5
Mann Whitney test	
P value	0,2350
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,F	129,0 , 171,0
Mann-Whitney U	51,00
Difference between medians	
Median of column A	5,500
Median of column F	10,50
Difference: Actual	-5,000
Difference: Hodges-Lehmann	-4,000

Table Analyzed	Fig S11A – Peptide 6
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Column A	No peptide
vs.	vs,
Column G	6
Mann Whitney test	
P value	0,1385
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,G	124,0 , 176,0
Mann-Whitney U	46,00
Difference between medians	
Median of column A	5,500
Median of column G	13,50
Difference: Actual	-8,000
Difference: Hodges-Lehmann	-6,000

Table Analyzed	Fig S11A – Peptide 7
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Column A	No peptide
vs.	vs,
Column H	7
Mann Whitney test	
P value	0,2016
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,H	127,5 , 172,5
Mann-Whitney U	49,50
Difference between medians	
Median of column A	5,500
Median of column H	10,50
Difference: Actual	-5,000
Difference: Hodges-Lehmann	-4,000

Table Analyzed	Fig S11A – Peptide 8
Column A	No peptide
vs.	vs,
Column I	8
Mann Whitney test	
P value	0,0246
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,I	111,5 , 188,5
Mann-Whitney U	33,50
Difference between medians	
Median of column A	5,500
Median of column I	12,00
Difference: Actual	-6,500
Difference: Hodges-Lehmann	-6,000

Table Analyzed	Fig S11A – Peptide 9
Column A	No peptide
vs.	vs,
Column J	9
Mann Whitney test	
P value	0,0281
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,J	112,0 , 188,0
Mann-Whitney U	34,00
Difference between medians	
Median of column A	5,500
Median of column J	17,50
Difference: Actual	-12,00
Difference: Hodges-Lehmann	-10,50

Table Analyzed	Fig S11A – Peptide 10
Column A	No peptide
vs.	vs,
Column K	10
Mann Whitney test	
P value	0,0270
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,K	112,0 , 188,0
Mann-Whitney U	34,00
Difference between medians	
Median of column A	5,500
Median of column K	17,50
Difference: Actual	-12,00
Difference: Hodges-Lehmann	-9,000

Table Analyzed	Fig S11A – Peptide 11
Column A	No peptide
vs.	vs,
Column L	11
Mann Whitney test	
P value	0,1466
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,L	124,5 , 175,5
Mann-Whitney U	46,50

Difference between medians	
Median of column A	5,500
Median of column L	10,50
Difference: Actual	-5,000
Difference: Hodges-Lehmann	-5,000

Table Analyzed	Fig S11A – Peptide 12
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Column A	No peptide
vs.	vs,
Column M	12
Mann Whitney test	
P value	0,1918
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,M	127,0 , 173,0
Mann-Whitney U	49,00
Difference between medians	
Median of column A	5,500
Median of column M	11,50
Difference: Actual	-6,000
Difference: Hodges-Lehmann	-4,000

Table Analyzed	Fig S11A – Peptide 13
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Column A	No peptide
vs.	vs,
Column N	13
Mann Whitney test	
P value	0,0497
Exact or approximate P value?	Exact
P value summary	*
Significantly different? (P < 0.05)	Yes
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,N	116,0 , 184,0
Mann-Whitney U	38,00
Difference between medians	
Median of column A	5,500
Median of column N	10,00
Difference: Actual	-4,500
Difference: Hodges-Lehmann	-5,500

Table Analyzed	Fig S11A – Peptide 14
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Column A	No peptide
vs.	vs,
Column O	14
Mann Whitney test	
P value	0,2123
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,O	128,0 , 172,0
Mann-Whitney U	50,00
Difference between medians	
Median of column A	5,500
Median of column O	8,500
Difference: Actual	-3,000
Difference: Hodges-Lehmann	-3,000

Table Analyzed	Fig S11A – Peptide 15
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Column A	No peptide
vs.	vs,
Column P	15
Mann Whitney test	
P value	0,0909

Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,P	120,5 , 179,5
Mann-Whitney U	42,50
Difference between medians	
Median of column A	5,500
Median of column P	12,50
Difference: Actual	-7,000
Difference: Hodges-Lehmann	-5,000

Table Analyzed	Fig S11A – Peptide 16
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Column A	No peptide
vs.	vs,
Column Q	16
Mann Whitney test	
P value	0,2124
Exact or approximate P value?	Exact
P value summary	ns
Significantly different? (P < 0.05)	No
One- or two-tailed P value?	Two-tailed
Sum of ranks in column A,Q	128,0 , 172,0
Mann-Whitney U	50,00
Difference between medians	
Median of column A	5,500
Median of column Q	11,50
Difference: Actual	-6,000
Difference: Hodges-Lehmann	-3,000