# SLC25A46 mutations in patients with Parkinson's Disease and optic atrophy

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#### ABSTRACT

Mutations in the gene encoding the mitochondrial carrier protein SLC25A46 are known to cause optic atrophy associated with peripheral neuropathy and congenital pontocerebellar hypoplasia.

We found novel biallelic *SLC25A46* mutations (p.H137R, p.A401Sfs\*17) in a patient with Parkinson's disease and optic atrophy. Screening of six unrelated patients with parkinsonism and optic atrophy allowed us to identify two additional mutations (p.A176V, p.K256R) in a second patient. All identified variants are predicted likely pathogenic and affect very conserved protein residues.

These findings suggest for the first time a possible link between Parkinson's Disease and SLC25A46 mutations. Replication in additional studies is needed to conclusively prove this link.

## 1. Introduction

Several monogenic forms of Parkinson's Disease (PD) are linked to genes encoding for mitochondrial proteins [1]. Mutations in *PINK1* and *Parkin*, two crucial players of the mitophagic machinery, cause recessive forms of PD, mainly characterized by early-onset PD and dystonia [2]. More complex phenotypes are linked to mutations in *OPA1*, *POLG* and *Twinkle* including: optic atrophy, ophthalmoplegia, neurosensorial hearing loss, neuropathy, cerebellar ataxia, and myopathy [3–5].

SLC25A46 encodes for a mitochondrial carrier protein located on the outer membrane that interacts with MFN2, OPA1, the mitochondrial contact site and cristae organizing system complex [6–8]. Mutations in this gene lead to a broad spectrum of neurodegenerative disorders. Congenital lethal pontocerebellar hypoplasia, optic atrophy, peripheral neuropathy, cerebellar ataxia, and spasticity are the clinical features described so far as consequences of loss of SLC25A46 function [9–11]. Nevertheless, parkinsonism has been reported neither as predominant feature, nor in association with a more complex phenotype

linked to SLC25A46 gene mutations.

Here we report, for the first time, the finding of biallelic likely pathogenetic *SLC25A46* mutations in patients affected by PD and optic atrophy.

## 2. Methods

A Caucasian family of Italian origin, including a single affected patient (Tn-1), was studied. There was no history of parkinsonism or other neurological disorders in previous generations, and there was no evidence of parental consanguinity. A screening of six unrelated patients displaying parkinsonism plus optic atrophy was performed detecting a second subject (Ps-1) with *SLC25A46* mutations.

The relevant ethical authorities approved the study and written informed consent was obtained from all participants to the publication of their images and videotapes, in both the print and online modalities. Neurological examination was performed by movement disorders specialists and included the Hoehn-Yahr scale, and Mini-Mental State Examination. Clinical details of the mutated subjects are reported in

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Table 1

Clinical details of patients carrying *SLC24A46* mutations. y, years; MRI, Magnetic Resonance Imaging; RBD, REM behaviour disorder; MMSE, Mini-Mental-State-Examination.

| Subject, gender                            | Tn-1, F  | Ps-1, M  |
|--|--|--|
| Familiarity for neurological diseases      | _  | + (mother: progressive polyneuropathy)   |
| Optic atrophy (age at onset)               | + (16y)  | + (52y)  |
| Neurosensorial hearing loss (age at onset) | <del>-</del>   | + (57y)  |
| Peripheral neuropathy                      | _  | +  |
| Pes cavus                                  | -  | +  |
| Age at PD diagnosis                        | 43y  | 63y  |
| Postural tremor (age at onset)             | + (43y)  | + (63y)  |
| Tremor at rest (age at onset)              | + (40y)  | + (63y)  |
| Bradykinesia (age at onset)                | + (43y)  | ++ (63y)   |
| Dystonia (age at onset)                    | + (foot) (46y)   | =  |
| Hoehn-Yahr Scale (age at evaluation)       | I (43y)  | III (63y)  |
| Response to Levodopa                       | ++   | +  |
| Levodopa induced dyskinesias               | +  | +++  |
| Cognitive impairment (age at evaluation)   | - (MMSE 30/30) (43y)   | ++, (MMSE 18/30) (72y)   |
| Non motor PD symptoms                      |  |  |
| Hyposmia                                   | +  | _  |
| Drooling                                   | =  | +  |
| Constipation                               | +++  | +  |
| Dysuria/Nicturia                           | +  | +  |
| Anxiety                                    | +  | _  |
| RBD  | _  | +  |
| SPECT DatSCAN                              | Significant pre-synaptic defect of the dopaminergic nigro-<br>striatal system, more evident on the left side | Pre-synaptic defect at putamen nuclei and left caudate nucleus   |
| MRI  | T2-weighted MRI hyperintense spot at the external capsule bilaterally  | T2-weighted multifocal bilateral white matter and lacunar hyperintensitie compatible with mild chronic ischemic cerebral disease |
| SLC25A46 mutations                         | p.H137R  | p.A176V  |
|  | p.A401Sfs*17   | p.K256R  |

## Table 1.

Genomic DNA was isolated from peripheral blood using standard protocols. After ruling out pathogenic mutations in known PD-related genes by a targeted Next Generation Sequence (NGS) panel (SNCA, LRRK2, GBA, PARKIN, PINK1, DJ-1, ATP13A2, PLA2G6, FBXO7, SYNJ1, DNAJC6, VPS13C, RAB39B, PTRHD1, POLG, OPA1, C10ORF12) and multiplex ligation-dependent probe amplification (MLPA) on both patients, exome sequencing was performed on individual Tn-1. Variants were annotated with Annovar and then filtered using the following criteria: 1) the variant being present in homozygous or compound heterozygous state 2) minor allele frequency ≤0.01% by ExAC, dbSNP, gnomAD; 3) coding non-synonymous; 4) predicted to affect splicing in silico; 5) predicted as likely pathogenetic by Mutation Taster, Polyphen2, CADD, M-CAP. Sanger sequencing was used to confirm the identified mutations and screen SLC25A46 intron-exon boundaries in all subjects available. Details on primers and PCR conditions are available upon request.

## 3. Results

## 3.1. Clinical findings in the Patient Tn-1

Patient Tn-1 was diagnosed with PD at the age of 43, with a 3-year history of left-hand rest tremor. The family history was unremarkable, with no consanguinity. Of note, visual problems appeared at age of 16, when the fundus examination revealed optic atrophy. VEP showed reduction of cortical evoked response amplitude bilaterally compatible with optic atrophy whereas electroretinogram was normal.

She complained of hyposmia, anxiety, and marked constipation from the early adulthood.

The neurologic examination at the time of diagnosis detected a mild parkinsonism with left-sided resting tremor and bradykinesia and walking difficulties with short steps (Video). No other neurological signs were detected except for a lower limb hyperreflexia and visual impairment. The response to L-Dopa and Dopamine-agonist was excellent. Neuropsychological assessment detected normal cognition.

Supplementary video related to this article can be found at https://doi.org/10.1016/j.parkreldis.2020.03.018

Brain MRI revealed T2-weighted hyperintensities in the external capsule bilaterally (Fig. 1a).

Both FDopa-PET and SPECT-with I123FP-CIT indicated a significant presynaptic defect of the dopaminergic nigrostriatal system, more evident on the left-side (Fig. 1b).

EMG/ENG was negative for sensitive or motor neuropathy.

## 3.2. Genetic results in the Patient Tn-1

Genetic analysis in the patient Tn-1 excluded mutations in mtDNA, OPA1, POLG1, PEO1, and mutations in genes related to autosomal dominant and recessive PD. After filtering variants (see Methods and Supplementary Materials) from the Exome sequencing analysis, two heterozygous variants were detected in the SLC25A46 gene. The c.1198\_1199insC, leading to a frameshift p.A401Sfs\*17, is absent by all databases. The mutation would lead either to produce an aberrant peptide followed by a premature stop codon in the functional mitochondrial carrier domain, or to affect the transcript stability through to a nonsense mediated decay mechanism (NMD). The c.410A>G, causing the missense p.H137R, is not reported by dbSNP, and is reported in only fourteen individuals in gnomAD (0,0054%), in the heterozygous state. This variant is predicted as deleterious by all in silico tools (Mutation Taster: disease causing 0.999; Polyphen2: probably damaging 0.972; CADD 21.3; M-CAP possibly pathogenic 0.027). Both variants were confirmed by Sanger sequencing. Genetic analysis in available family members confirmed these variants being on independent alleles.

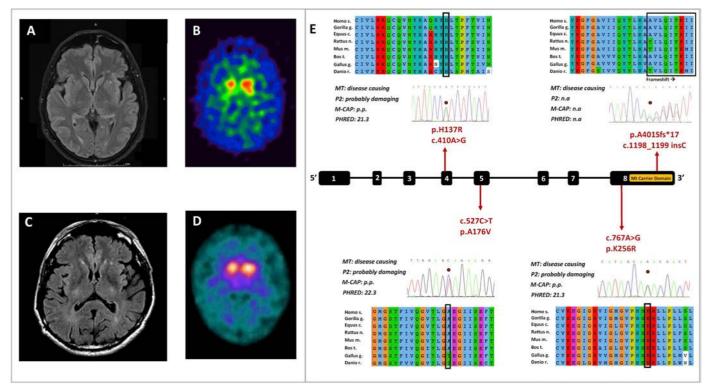


Fig. 1. A-C) FLAIR T2-weighted MRI showing bilatzeral external capsule hyperintensity in patient Tn-1 (A) and white matter and lacunar hyperintensities in patient Ps-1 (C).

B-D) SPECT with 123I-FP-CIT showing significant presynaptic defect of nigrostriatal system, more evident on the left side in patient Tn-1 (B) and a reduced radioligand uptake in the putamen bilaterally and in the left caudate in patient Ps-1 (D).

#### 3.3. Genetic screening

After a *SLC25A46* gene screening of six patients displaying parkinsonism plus optic atrophy, one additional patient was detected presenting two *SLC25A46* variants.

Patient Ps-1 was diagnosed with PD at the age of 63, preceded by a 5-year history of REM behavior disorder, hyposmia, and depression. He had a history of gait difficulties, lower limb muscle hypotrophy and pes cavus from the childhood with a recent genetic diagnosis of CMT1A (17p11.2 duplication). Visual problems appeared in the adulthood and a diagnosis of optic atrophy was established at the age of 52. In addition, he was affected by progressive sensorineural hearing loss starting from the age of 57.

No other family members were affected by parkinsonism and optic atrophy. His mother presented a progressive polyneuropathy from the early adulthood. The course of the disease was characterized by a good response to L-Dopa and IMAO-B, peak-dose dyskinesias, postural instability, cognitive decline and visual hallucinations.

Brain MRI displayed T2-weighted multifocal bilateral white-matter and lacunar hyperintensities compatible with a mild chronic ischemic cerebral disease (Fig. 1c) and mild diffuse cerebral atrophy. 18F-FDG PET showed a bilateral temporoparietal metabolism reduction, and SPECT with 123I-FP-CIT revealed reduced radioligand uptake in the putamen bilaterally and in the left caudate (Fig. 1d). Florbetapir PET was negative.

Mutations in mtDNA and genes related to autosomal dominant and recessive PD were excluded in subject Ps-1. Sanger sequencing of all *SLC25A46* coding sequence and intron-exon boundaries detected two heterozygous variants. The c.527C>T causes the missense substitution p.A176V, which is absent in all databases and predicted as be deleterious by all in silico tools (Mutation Taster: disease causing, 0.999; Polyphen2 probably damaging 0.972; CADD 22.3; M-CAP possibly pathogenic 0,027). The c.767A>G, leading to the missense mutation

p.K256R is reported as rare (0,21% in gnomAD) and predicted to be deleterious by in silico tools (Mutation Taster: disease causing, 0.999; Polyphen2: probably damaging 0.972; CADD 24.5; M-CAP possibly pathogenic 0.225). No family members were available for segregation testing of the variants nor was possible to isolate RNA.

#### 4. Discussion

Mutations in *SLC25A46* have been associated with a very broad spectrum of neurological disorders. Initially, *SLC25A46* mutations were found in patients with optic atrophy and peripheral neuropathy, and later also in subjects with pontocerebellar hypoplasia, cerebellar and myoclonic ataxia, and limb spasticity [9–11].

The SLC25A46 protein was identified for the first time as a mitochondrial solute carrier widely expressed in the central nervous system [12]. Recently, SLC25A46 possible involvement in mitochondrial dynamics is suggested through the direct interaction with the components of the mitochondrial fusion machinery OPA1 and MFN1/2 [7]. Interestingly, the phenotype linked to *OPA1* mutations, which is mainly characterized by optic atrophy, may lead in some cases to a complex neurodegenerative syndrome including parkinsonism, ataxia, neuropathy and dementia [13].

Similarly, other mitochondrial gene mutations may lead to syndromes with complex neurological manifestation, including ataxia, ophthalmoplegia, optic atrophy, parkinsonism and myopathy (e.g. *POLG*, *Twinkle*) [14,15].

The reason of a such significant variability in *SLC25A46* mutations phenotypic expression is still unclear. No defined genotype-phenotype correlation has been determined yet, except for a recent report suggesting a negative correlation between SLC25A46 protein stability and phenotype severity [16].

The finding that in neuron-specific knockdown Drosophila at synapse level an accumulation of reactive oxygen species and reduction of ATP have been observed may suggest a pathogenetic mechanism related to oxidative stress, which is largely associated with the specific loss of dopaminergic neurons in PD [17]. Among the cases with *SLC25A46* mutations reported so far, the parkinsonism has never been observed. Of note, parkinsonism appeared in both patients only in adulthood comparing with the patients described with *SLC25A46*-related optic atrophy or pontocerebellar hypoplasia, that appeared early in life. This could explain why parkinsonism has not been observed in the cases described so far.

Both patients described here present a diagnosis of PD, with a broad age of onset and course of the disease. Indeed, patient Tn-1 shows an early-onset PD with slow progression, foot dystonia and anxiety, which resemble the phenotype of *Parkin/PINK1*-mutated PD patients, whereas patient Ps-1 presents a PD with a classical age of onset, complicated then by motor fluctuations and cognitive impairment.

A possible hypothesis explaining such a difference between the two subjects could be related to the different severity of the *SLC25A46* mutations identified. The c.1198\_1199insC in patient Tn-1 may lead to a juvenile onset parkinsonism due to the expected deleterious consequence on *SLC25A46* protein, by replacing the mitochondrial carrier domain sequence with an aberrant peptide or affecting the transcript through NMD.

Both subjects manifested optic atrophy, in line with most of the *SLC25A46* cases described so far.

Genotype-phenotype correlation in Ps-1 is made more difficult by the presence of the CMT1A related duplication which can account for some clinical features such as familial pes cavus and lower-limb sensorimotor polyneuropathy. Nevertheless, CMT1A has never been associated with optic atrophy or parkinsonism. Moreover, patient Ps-1 suffered from progressive sensorineural hearing loss. Despite auditory impairment has been described in CMT type 1 and 2, the neurosensorial hearing loss detected in Ps-1 appeared at later stage of the disease, suggesting a different etiology that observed in CMT patients [18]. The combination of mutations in two different genes in patient Ps-1 configures a double-trouble condition and may explain the complexity of his phenotype.

Neuroimaging features of these patients do not resemble those of previously reported *SLC25A46*-mutated cases, mainly characterized by cerebellar atrophy/hypoplasia. Indeed, patient Tn-1 showed a slight hyperintensity in the external capsule and subject Ps-1 displayed white matter and lacunar hyperintensities.

Two out of the four mutations detected lie on transmembrane protein domains, possibly affecting SLC25A46 membrane localization. The frameshift mutation lies on the mitochondrial carrier domain likely determining an impairment in protein carrier function.

In conclusion, these results likely expand the neurodegenerative presentations linked with recessive *SLC25A46* mutations now featuring parkinsonism in addition to optic atrophy and hearing loss. Despite the promising results linking the new identified *SLC25A46* variants and parkinsonism, the exome sequencing approach on a single subject and *SLC25A46* screening in a small sample set of patients with optic atrophy and late-onset PD is not conclusive and need to be replicated in a larger sample set. After independent confirmations, *SLC25A46* gene should be definitively considered in the screening of genetic forms of PD. Understanding how mutations in *SLC25A46* gene result in parkinsonism will shed further light on the molecular mechanisms of PD.

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#### Declaration of competing interest

The authors report no conflict of interest related to this work.

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#### Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.parkreldis.2020.03.018.

#### References

- [1] Y.C. Wong, K. Luk, K. Purtell, S. Burke Nanni, A.J. Stoessl, L.-E. Trudeau, Z. Yue, D. Krainc, W. Oertel, J.A. Obeso, L.A. Volpicelli-Daley, Neuronal vulnerability in Parkinson disease: should the focus be on axons and synaptic terminals? Mov. Disord. (2019), https://doi.org/10.1002/mds.27823.
- [2] A.M. Pickrell, R.J. Youle, The roles of PINK1, parkin, and mitochondrial fidelity in Parkinson's disease, Neuron 85 (2015) 257–273, https://doi.org/10.1016/j.neuron. 2014 12 007
- [3] P. Yu-Wai-Man, M. Votruba, F. Burté, C. La Morgia, P. Barboni, V. Carelli, A neurodegenerative perspective on mitochondrial optic neuropathies, Acta Neuropathol. 132 (2016) 789–806, https://doi.org/10.1007/s00401-016-1625-2.
- [4] J. Wan, J. Steffen, M. Yourshaw, H. Mamsa, E. Andersen, S. Rudnik-Schöneborn, K. Pope, K.B. Howell, C.A. McLean, A.J. Komberg, J. Joseph, P.J. Lockhart, K. Zerres, M.M. Ryan, S.F. Nelson, C.M. Koehler, J.C. Jen, Loss of function of SLC25A46 causes lethal congenital pontocerebellar hypoplasia, Brain 139 (2016) 2877–2890, https://doi.org/10.1093/brain/aww212.
- [5] T. van Dijk, S. Rudnik-Schöneborn, J. Senderek, G. Hajmousa, H. Mei, M. Dusl, E. Aronica, P. Barth, F. Baas, Pontocerebellar hypoplasia with spinal muscular atrophy (PCH1): identification of SLC25A46 mutations in the original Dutch PCH1 family, Brain 140 (2017) e46, https://doi.org/10.1093/brain/awx147.
- [6] A.J. Abrams, R.B. Hufnagel, A. Rebelo, C. Zanna, N. Patel, M.A. Gonzalez, L.J. Campeanu, L.B. Griffin, S. Groenewald, A.V. Strickland, F. Tao, F. Speziani, L. Abreu, R. Schüle, L. Caporali, C. La Morgia, A. Maresca, R. Liguori, R. Lodi, Z.M. Ahmed, K.L. Sund, X. Wang, L.A. Krueger, Y. Peng, C.E. Prada, C.A. Prows, E.K. Schorry, A. Antonellis, H.H. Zimmerman, O.A. Abdul-Rahman, Y. Yang, S.M. Downes, J. Prince, F. Fontanesi, A. Barrientos, A.H. Németh, V. Carelli, T. Huang, S. Zuchner, J.E. Dallman, Mutations in SLC25A46, encoding a UGO1-like protein, cause an optic atrophy spectrum disorder, Nat. Genet. 47 (2015) 926–932, https://doi.org/10.1038/ng.3354.
- [7] A. Janer, J. Prudent, V. Paupe, S. Fahiminiya, J. Majewski, N. Sgarioto, C. Des Rosiers, A. Forest, Z.-Y. Lin, A.-C. Gingras, G. Mitchell, H.M. McBride, E.A. Shoubridge, SLC25A46 is required for mitochondrial lipid homeostasis and cristae maintenance and is responsible for Leigh syndrome, EMBO Mol. Med. 8 (2016) 1019–1038, https://doi.org/10.15252/emmm.201506159.
- [8] J. Steffen, A.A. Vashisht, J. Wan, J.C. Jen, S.M. Claypool, J.A. Wohlschlegel, C.M. Koehler, Rapid degradation of mutant SLC25A46 by the ubiquitin-proteasome system results in MFN1/2-mediated hyperfusion of mitochondria, Mol. Biol. Cell 28 (2017) 600–612, https://doi.org/10.1091/mbc.E16-07-0545.
- [9] G. Charlesworth, B. Balint, N.E. Mencacci, L. Carr, N.W. Wood, K.P. Bhatia, SLC25A46 mutations underlie progressive myoclonic ataxia with optic atrophy and neuropathy, Mov. Disord. 31 (2016) 1249–1251, https://doi.org/10.1002/mds. 26716
- [10] R.A. Sulaiman, N. Patel, H. Alsharif, S.T. Arold, F.S. Alkuraya, A novel mutation in SLC25A46 causes optic atrophy and progressive limb spasticity, with no cerebellar atrophy or axonal neuropathy: letter to the Editor, Clin. Genet. 92 (2017) 230–231, https://doi.org/10.1111/cge.12963.
- [11] M.B. Hammer, J. Ding, F. Mochel, G. Eleuch-Fayache, P. Charles, M. Coutelier, J.R. Gibbs, S.K. Arepalli, S.B. Chong, D.G. Hernandez, E. Majounie, S. Clipman, Y. Bouhlal, H. Nehdi, A. Brice, F. Hentati, G. Stevanin, R. Amouri, A. Durr, A.B. Singleton, SLC25A46 mutations associated with autosomal recessive cerebellar ataxia in North African Families, Neurodegener. Dis. 17 (2017) 208–212, https://doi.org/10.1159/000464445.
- [12] T. Haitina, J. Lindblom, T. Renström, R. Fredriksson, Fourteen novel human members of mitochondrial solute carrier family 25 (SLC25) widely expressed in the central nervous system, Genomics 88 (2006) 779–790, https://doi.org/10.1016/j. ygeno.2006.06.016.
- [13] V. Carelli, O. Musumeci, L. Caporali, C. Zanna, C. La Morgia, V. Del Dotto, A.M. Porcelli, M. Rugolo, M.L. Valentino, L. Iommarini, A. Maresca, P. Barboni, M. Carbonelli, C. Trombetta, E.M. Valente, S. Patergnani, C. Giorgi, P. Pinton, G. Rizzo, C. Tonon, R. Lodi, P. Avoni, R. Liguori, A. Baruzzi, A. Toscano, M. Zeviani, Syndromic parkinsonism and dementia associated with OPA1 missense mutations, Ann. Neurol. 78 (2015) 21–38, https://doi.org/10.1002/ana.24410.
- [14] S. Rahman, W.C. Copeland, POLG-related disorders and their neurological

- manifestations, Nat. Rev. Neurol. 15 (2019) 40-52, https://doi.org/10.1038/ s41582-018-0101-0.
- [15] H. Morino, S.B. Pierce, Y. Matsuda, T. Walsh, R. Ohsawa, M. Newby, K. Hiraki-Kamon, M. Kuramochi, M.K. Lee, R.E. Klevit, A. Martin, H. Maruyama, M.-C. King, H. Kawakami, Mutations in Twinkle primase-helicase cause Perrault syndrome with neurologic features, Neurology 83 (2014) 2054–2061, https://doi.org/10.1212/WNL.000000000001036.
- [16] A.J. Abrams, F. Fontanesi, N.B.L. Tan, E. Buglo, I.J. Campeanu, A.P. Rebelo, A.J. Kornberg, D.G. Phelan, Z. Stark, S. Zuchner, Insights into the genotype-
- $phenotype\ correlation\ and\ molecular\ function\ of\ SLC25A46,\ Hum.\ Mutat.\ 39$
- (2018) 1995–2007, https://doi.org/10.1002/humu.23639.
  [17] K. Suda, I. Ueoka, Y. Azuma, Y. Muraoka, H. Yoshida, M. Yamaguchi, Novel Drosophila model for mitochondrial diseases by targeting of a solute carrier protein SLC25A46, Brain Res. 1689 (2018) 30–44, https://doi.org/10.1016/j.brainres. 2018.03.028.
- [18] G. Rance, M.M. Ryan, K. Bayliss, K. Gill, C. O'Sullivan, M. Whitechurch, Auditory function in children with Charcot-Marie-Tooth disease, Brain 135 (2012) 1412– 1422, https://doi.org/10.1093/brain/aws085.