## **Electronic Supplementary Information**

# FTIR microscopy for direct observation of conformational changes on immobilized $\omega$ -transaminase: effect of water activity and organic solvent on biocatalyst performance

Iulia-Ioana Rădoi<sup>1,4</sup>, Diana Eva Bedolla<sup>2,3</sup>, Lisa Vaccari<sup>2</sup>, Anamaria Todea<sup>1</sup>, Federico Zappaterra<sup>1</sup>, Alexey Volkov<sup>4</sup>, Lucia Gardossi<sup>1</sup><sup>+</sup>

<sup>1</sup> Department of Chemical and Pharmaceutical Sciences, University of Trieste, Via L. Giorgieri 1, 34127 Trieste TS, Italy

<sup>2</sup> Elettra-Sincrotrone Trieste S.C.p.A., Area Science Park, 34149 Basovizza TS, Italy

<sup>3</sup> AREA Science Park - Padriciano 99, IT-34149 Trieste Italy

<sup>4</sup> EnginZyme AB, Tomtebodavägen 6, 171 65 Solna, Sweden

+Corresponding author: Lucia Gardossi, e-amil: gardossi@units.it

Table of content

1.	EziG specifications	3
2.	Activity assay in aqueous media of all immobilized ATA-117-His-tag formulations	3
3.	Spectroscopic characterization of EziG carriers by means of FTIR microscopy	4
4.	Fluorescence microscopy	6
5.	FTIR investigation of the secondary structure of immobilized and pure ATA-117	7
6.	EziG-immobilized ATA 117: activity in organic solvents	9
	Bibliography	10

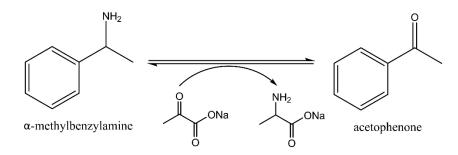
#### 1. EziG specifications

Carrier	Surface	Particle size [μm]	Pore diameter [nm]	Coating	рН range	Chelated Fe <sup>3+</sup> [µmol/g]
EziG <sup>1</sup> -Opal	hydrophilic	75 - 125	50±5	long amino alkyl chain	5 - 10	>10
EziG <sup>2</sup> -Coral	hydrophobic	75 - 125	30±5	polyvinyl benzyl chloride	5 - 10	>10
EziG <sup>3</sup> - Amber	semi- hydrophilic	75 - 125	30±5	copolymer of styrene and acrylonitrile	5 - 10	>10

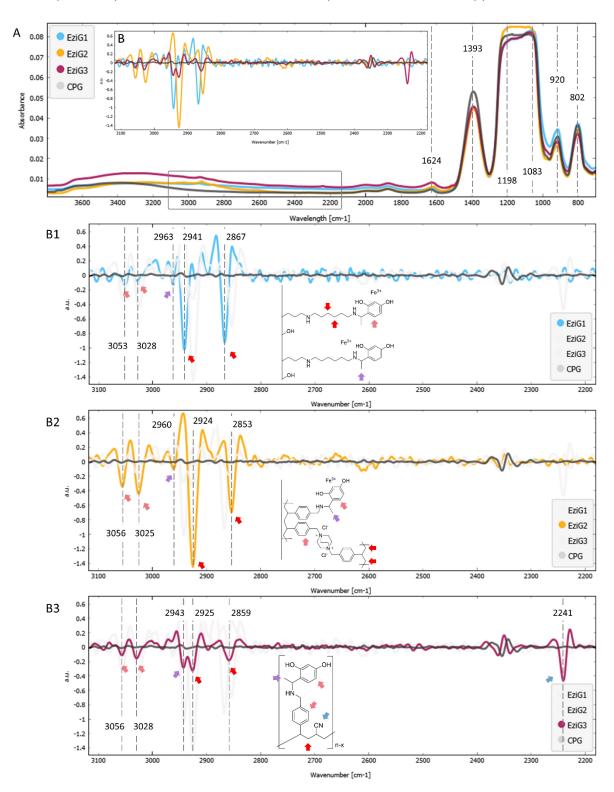
Table S1. EziG carriers' specifications: all EziG products are based on controlled porosity glass.

All information is from EziG<sup>™</sup> Product Data Sheet.

2. Activity assay in aqueous media of all immobilized ATA-117-His-tag formulations



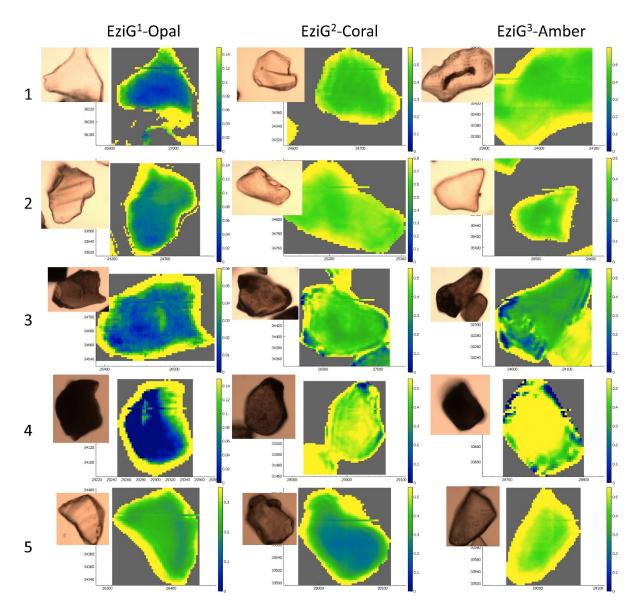
Scheme S1.Model reaction used in the spectrophotometric assay of the activity in aqueous media of all immobilized  $His_6$ -ATA-117 formulations pretreated according to the protocols of Table 1 (manuscript). The reaction was monitored by following the formation of acetophenone in a microplate reader by measuring the increase in absorbance at 245 nm.



3. Spectroscopic characterization of EziG carriers by means of FTIR microscopy

Figure S1. A: FTIR spectra (3800–700 cm<sup>-1</sup> region) of EziG<sup>1</sup>-Opal (blue), EziG<sup>2</sup>-Coral (orange), EziG<sup>3</sup>-Amber (dark red), and uncoated CPG (gray); black dashed lines indicate the FTIR spectral vibrations of<sup>1–3</sup>: H-O-H bending at 1624 cm<sup>-1</sup>, B-O-B at 1393 cm<sup>-1</sup>, asymmetric Si-O stretching at 1198 and 1083 cm<sup>-1</sup>, Si-O-B stretching at 920 cm<sup>-1</sup> and Si-O-Si cm<sup>-1</sup> stretching at 802 cm<sup>-1</sup>. B: Second derivative FTIR spectra (vector normalized in the spectra region 3120–2180 cm<sup>-1</sup>)

of the region where the most diagnostic differences are seen. In B1 (EziG<sup>1</sup>-Opal), B2 (EziG<sup>2</sup>-Coral) and B3 (EziG<sup>3</sup>-Amber) panels the main functional groups in the coating polymer composition ae highlighted. Each of the spectra contains an insertion of the possible formula of the polymer coating <sup>4,5</sup>: the peaks at 3053, 3056 cm<sup>-1</sup> and 3028, 3025 cm<sup>-1</sup> were attributed to the aromatic C-H stretches; peaks at 2963, 2960 and 2943 cm<sup>-1</sup> were associated with C-H stretches in CH<sub>3</sub>; peaks at 2941, 2925, 2924 cm<sup>-1</sup> and 2087, 2859, 2853 cm<sup>-1</sup> from CH<sub>2</sub> asymmetric and symmetric stretches; the peak at 2241 cm<sup>-1</sup> that can be observed only for EziG<sup>3</sup>-Amber is attributed to C-N stretching.



Scheme S2. The optical images of the carriers with the overlap of the chemical infrared images generated by the ratio of the integration in the specific region of Amide I and Amide II bands (~1710-1480 cm<sup>-1</sup>) and the band identifying the glass carrier (~1500-1300 cm<sup>-1</sup>). The colour scale bar ranges from higher biocatalyst concentration (yellow areas) to not-detectable concentrations (blue areas).

### 4. Fluorescence microscopy

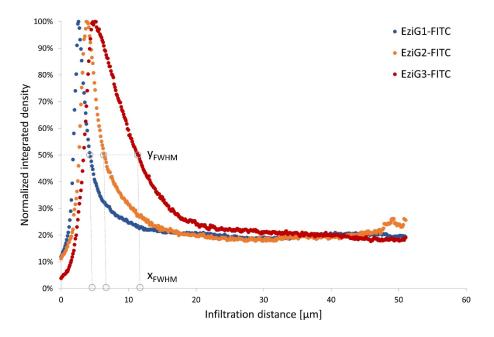
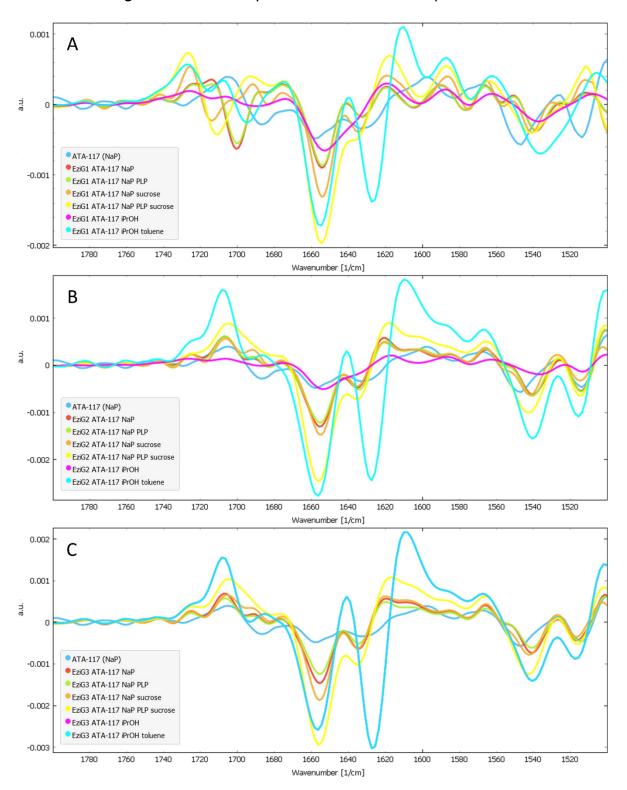


Figure S3. Radial cross-section fluorescence profile of EziG carriers shown in Figure 4.

 $FWHM = R - X_{FWHM}$ 



5. FTIR investigation of the secondary structure of immobilized and pure ATA-117

Figure S4. Savitzky-Golay's second derivative spectra for Amide I and II – comparison between different washes. The comparison was presented between native ATA-117 (resuspended in NaP buffer), immobilized ATA-117, but also the buffer used for the purification and resuspension of the enzyme. The spectra were grouped as follow: A. EziG<sup>1</sup>-Opal, B. EziG<sup>2</sup>-Coral, C. EziG<sup>3</sup>-Amber (samples were dried under vacuum at 100 mbar, 25°C for 24h).

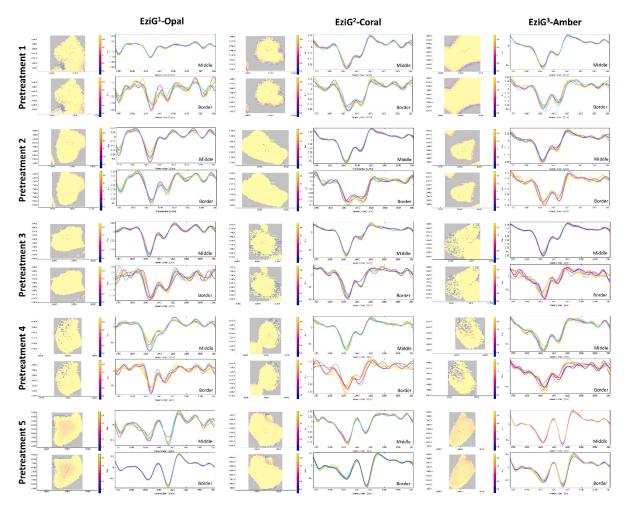
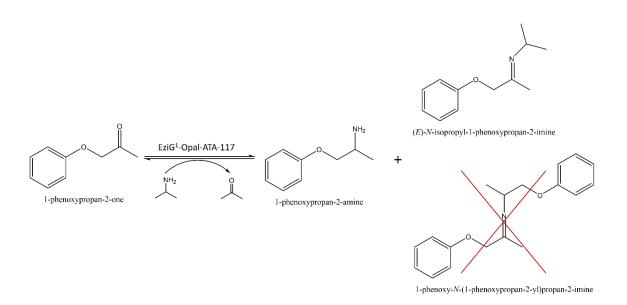


Figure S5. Savitzky-Golay's second derivative spectra for Amide I and II – comparison betweendifferent washes looking to representative spots of FRIR images at the border and at thecenter of the carrier. The integral from baseline of Amide I and Amide II and for the glasscarrierwerecalculatedandratioed.

#### 6. EziG-immobilized ATA 117: activity in toluene



Scheme S3. Model reaction used to evaluate the effect of the water on the performance of the immobilized ATA-117 on EziG<sup>1</sup>-Opal; reaction was performed in toluene. The formation of the imine between the ketone substrate and the formed 1-phenoxypropan-2-amine was not observed.

Table S2. Values of water activity measured after 24h of equilibration of some reference mixtures. All samples were incubated in 4 mL vial at 25°C for 24h. The achievement of the equilibrium was confirmed by the constant values of  $a_w$  observed across the time (see materials and methods).

Solvent	Pretreatment of E							
Solvent	Rinse with	Drying	a <sub>w</sub>					
Control mixtures								
dry toluene	no	no	0.22±0.01					
dry toluene	iPrOH followed by dry toluene	under vacuum 100mbar, 25°C, 24h	0.22±0.01					
dry toluene	iPrOH followed by dry toluene	no	0.81±0.01					
water-saturated toluene	no	no	0.60±0.01					

#### Bibliography

- 1 L. Pang, R. Zhao, J. Chen, J. Ding, X. Chen, W. Chai, X. Cui, X. Li, D. Wang and H. Pan, *Bioact Mater*, 2022, **12**, 1–15.
- 2 A. M. E<sup>®</sup>mov and V. G. Pogareva, *Water-related IR absorption spectra for some phosphate and silicate glasses*, .
- 3 S. Y. Marzouk, R. Seoudi, D. A. Said and M. S. Mabrouk, *Opt Mater (Amst)*, 2013, **35**, 2077–2084.
- M. R. Jung, F. D. Horgen, S. v. Orski, V. Rodriguez C., K. L. Beers, G. H. Balazs, T. T. Jones, T. M. Work, K. C. Brignac, S. J. Royer, K. D. Hyrenbach, B. A. Jensen and J. M. Lynch, *Mar Pollut Bull*, 2018, **127**, 704–716.
- 5 S. Krimm, C. Y. Liang and G. B. B. M. Sutherland, *J Chem Phys*, 1956, **25**, 549–562.