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Supporting Information

Double Chalcogen Bonding Recognition Arrays in Solution

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1. General remarks

1.1 Instrumentation

Thin layer chromatography (TLC) was conducted on pre-coated aluminum sheets with 0.20 mm *Merck Millipore* Silica gel 60 with fluorescent indicator F254. *Column chromatography* was carried out using *Merck Gerduran* silica gel 60 (particle size 40-63 µm). *Melting points* (mp) were measured on a *Gallenkamp* apparatus in open capillary tubes and have not been corrected. *Nuclear magnetic resonance:* (NMR) spectra were recorded on a Bruker Fourier 300 MHz spectrometer equipped with a dual $(^{13}C,~^{1}H)$ probe, a Bruker AVANCE III HD 400 MHz NMR spectrometer equipped with a Broadband multinuclear (BBFO) SmartProbe™, a

Bruker AVANCE III HD 500 MHz Spectrometer equipped with Broadband multinuclear (BBO) Prodigy CryoProbe or a Bruker AV III HDX 700 MHz NMR spectrometer (Bruker BioSpin, Rheinstetten, Germany) with a quadruple (${}^{1}H$, ${}^{13}C$, ${}^{15}N$ ${}^{19}F$) inverse helium cooled cryo probe. ¹H spectra were obtained at 300, 400, 500, 600 or 700 MHz, ${}^{13}C{}^{1}H$ } spectra were obtained at 75, 100, 125, 150 or 175 MHz NMR and ¹⁹F spectra were obtained at 376, 470 and 659 MHz. 125 Te NMR experiments were done on a Bruker AV III 600 MHz NMR spectrometer using a nitrogen cooled broad band observe cryo probe at a resonance frequency of 189.38 MHz. All spectra were obtained at r.t. Chemical shifts were reported in ppm relative to tetramethylsilane using the residual solvent signal for ¹H or the solvent signal for ¹³C as an internal reference (CDCl₃: δ_H = 7.26 ppm, δ_C = 77.16 ppm; C₆D₆: δ_H = 7.16 ppm, δ_c = 128.06 ppm). Chemical shifts for ¹⁹F and ¹²⁵Te are reported on a unified scale relative to ¹H using the Ξ value for CDCl₃.^[28] Coupling constants (*J*) were given in Hz. Resonance multiplicity was described as *s* (singlet), *d* (doublet), *t* (triplet), *dd* (doublet of doublets), *ddd* (doublet of doublets of doublets), *dm* (doublet of multiplets), *q* (quartet), *m* (multiplet) and *bs* (broad signal). Carbon spectra were acquired with ${}^{1}H$ decoupling. ${}^{77}Se$ NMR experiments were either recorded on a Bruker AV III HDX 700 NMR spectrometer (Bruker BioSpin, Rheinstetten, Germany) equipped with a broad band observe probe, or on a Bruker AV III 600 NMR spectrometer using a nitrogen cooled broad band observe cryo probe. The resonance frequency for 77Se was 133.58 MHz or 114.48 MHz, respectively. *Infrared spectra* (IR) were recorded on a Shimadzu IR Affinity 1S FTIR spectrometer in ATR mode with a diamond monocrystal. *Mass spectrometry:* (i) High-resolution ESI mass spectra (HRMS) were performed on a Waters LCT HR TOF mass spectrometer in the positive or negative ion mode. *X-ray measurements:* The X-ray intensity data of 1_{Ph}, 1_{Alk}, 1_{Anthr}, 1_{Met} and 1_{3Met} were measured on Bruker D8 Venture diffractometer equipped with multilayer monochromator, Mo and Cu K/α INCOATEC micro focus sealed tubes and Oxford cooling system. The structures were solved by Direct Methods and Intrinsic Phasing. Non-hydrogen atoms were refined with anisotropic displacement parameters. Hydrogen atoms were inserted at calculated positions and refined with riding model. The following software was used: Bruker SAINT software package^[29] using a narrow-frame algorithm for frame integration, SADABS^[30] for absorption correction, OLEX2^[31] for structure solution, refinement, molecular diagrams and graphical user-interface, Shelxle^[32] for refinement and graphical user-interface SHELXS-2015^[32] for structure solution, SHELXL-2015^[33] for refinement, Platon^[34] for symmetry check. Data collections of 1_{3F} were performed at the X-ray diffraction beamline (XRD1) of the Elettra Synchrotron (Trieste, Italy).[35] The crystals were dipped in NHV oil (Jena Bioscience, Jena, Germany) and mounted on the goniometer head with nylon loops (MiTeGen, Ithaca, USA). Complete datasets were collected at 100 K (nitrogen stream supplied through an Oxford Cryostream 700). Data were acquired using a monochromatic wavelength of 0.70 Å through the rotating crystal method on a Pilatus 2M hybrid-pixel area detector (DECTRIS Ltd., Baden-Daettwil, Switzerland). The diffraction data were indexed and integrated using XDS.^[36] The structure was solved with Olex2^[37] by using ShelXT^[38] structure solution program by Intrinsic Phasing and refined with the ShelXL^[39] refinement package using least-squares minimization. In the last cycles of refinement, non-hydrogen atoms were refined anisotropically. Hydrogen atoms were included in calculated positions, and a riding model was used for their refinement. Crystal data, data collection parameters, and structure refinement details are given in Tables S1 to S6.

1.2 Materials and methods

Chemicals were purchased from *Sigma Aldrich*, *Acros Organics*, *TCI, Apollo Scientific, ABCR, Alfa Aesar, Carbosynth* and *Fluorochem* and were used as received. Solvents were purchased from *Fluorochem, Fisher Chemical* and *Sigma Aldrich*, while deuterated solvents from *Eurisotop* and *Sigma Aldrich*. THF, Et₂O and CH₂Cl₂ were dried on a Braun MB SPS-800 solvent purification system. MeOH, CHCl₃ and acetone were purchased as reagent-grade and used without further purification. $E_{13}N$ was distilled from CaH_2 and then stored over KOH. Anhydrous dioxane and pyridine were purchased from *Sigma Aldrich*. Solutions of *i*-PrMgCl in THF were freshly prepared according to a procedure of Lin et al.^[40] and titrated with the Paquette method,[41] or directly purchased from *Sigma Aldrich*. Low temperature baths were prepared using different solvent mixtures depending on the desired temperature: 0 °C with $ice/H₂O$. Anhydrous conditions were achieved by flaming two necked flasks with a heat gun under vacuum and purging with N_2 . The inert atmosphere was maintained using Nitrogen-filled balloons equipped with a syringe and needle that was used to penetrate the silicon stoppers closing the flask's necks. Additions of liquid reagents were performed using dried plastic or glass syringes. All reactions were performed in dry conditions and under inert atmosphere unless otherwise stated.

2. Synthetic procedures

General comments for the preparation of the Pyrselen derivatives:

To improve the yield of **3**, *t*-BuOK needs to be activated by flame-drying it under vacuum for 20 minutes. Similarly, to freshly prepare $Li₂Se₂$, the activation of $Li⁰$ consists of pressing elemental lithium chunks with a test tube, then the resulting plates are washed with degassed petr. ether, dried under a flow of Ar and added to a solution of dry THF. For the synthesis of **1F** and **13F**, 2.5 equivalents of corresponding primary amine were used (rather than 1.2), which were added to the acyl chloride within 30 minutes at -40 °C.

2.1 Synthesis of *tert***-butyl 2-chloronicotinate 3**

To a two-necked 50 mL round-bottomed flask with a suspension of 2-chloronicotinic acid **2** $(5 g, 31.7 mmol)$ in SOCl₂ (25 mL) under anhydrous condition, dry DMF (2 drops) was added dropwise, then the reaction heated up to reflux, stirred for 4 h and the solvents distilled off. To a two-necked 50 mL round-bottomed flask with a solution of activated *t*-BuOK (3.56 g, 31.7 mmol) in dry THF (10 mL) under anhydrous condition, a solution of the resulting acyl chloride derivative in dry THF (8 mL) was added at -15 °C over 1 h. The reaction was slowly allowed to warm up to room temperature, stirred for 16 h, then poured in cold water (30 mL) and extracted with CH_2Cl_2 (3 \times 50 mL). The combined organic extracts were washed with brine (30 mL), dried over Na₂SO₄, filtered and the solvents evaporated *in vacuo*. The crude was purified by silica gel chromatography (CH₂Cl₂) to give pure 3 as an orange oil (4.1 g, 61% yield).

¹H NMR (700 MHz, CDCl₃) *δ*: 8.45 (*d*, *J*_{H,H} = 4.5 Hz, 1H, *H_a*), 8.04 (*d*, *J*_{H,H} = 7.6 Hz, 1H, *H_c*), 7.28 (*d*, $J_{H,H}$ = 7.6, 4.5 Hz, 1H, H_b), 1.59 (*s*, 9H, H_d); ¹³C NMR (175 MHz, CDCl₃) *δ*: 163.9, 151.2, 149.4, 139.7, 128.8, 122.0, 28.0; FTIR (ATR): *ν* (cm-1): 2980, 2936, 1723, 15778, 1561, 1479, 1457, 1400, 1369, 1312, 1285, 1252, 1172, 1139, 1063, 1055, 1036, 865, 847, 819, 765, 718, 647, 543, 476, 432; HRMS (ESI): *m/z* calcd for C₁₀H₁₂NOCl+Na⁺: 236.0449 [M+Na]⁺; found: 236.0445.

2.2 Synthesis of di-*tert***-butyl 2,2'-diselanediyldinicotinate 4**

To a flame-dried Schlenk tube loaded with a suspension of activated lithium (120 mg, 17.3 mmol) and 4,4´-di-*tert*-butylbiphenyl (100 mg, 0.38 mmol) in dry THF (25 mL) under anhydrous condition, a gentle vacuum was applied and the reaction sonicated for 15 minutes (colorless turned into dark green). Freshly grounded elemental selenium (4 g, 50.7 mmol) was added while a brisk flux of N_2 was passed through the flask, then the reaction sonicated at 50 °C for 4 h. To a suspension of the resulting dilithium diselenide derivative, a solution of *t*-butyl 2-chloroquinoline-3-carboxylate **3** (720 mg, 2.74 mmol) in dry THF (8 mL) was added dropwise at -15 °C. The reaction was slowly allowed to warm up to room temperature, stirred for 72 h, then quenched with MeOH (30 mL) and filtered over celite. The filtrate was concentrated under reduced pressure, then MeOH (30 mL) added, and the resulting suspension centrifugated for 15 minutes (5000 rpm). The system was filtered and the solid washed with MeOH (2 × 10 mL) to give pure **4** as an orange solid (340 mg, 33% yield).

mp: 208-210 °C; ¹H NMR (600 MHz, CDCl₃) *δ*: 8.47 (*dd*, *J*_{H,H} = 4.7, 1.8 Hz, 2H, *H_a*), 8.13 (*dd*, *J*H,H = 7.7, 1.8 Hz, 2H, *Hc*), 7.09 (*dd*, *J*H,H = 7.7, 4.7 Hz, 2H, *Hb*), 1.64 (*s*, 18H, *Hd*); 13C NMR (150 MHz, CDCl3) *δ*: 165.4, 161.1, 152.5, 138.2, 126.4, 119.4, 28.3; FTIR (ATR): *ν* (cm-1): 29707, 2931, 1682, 1567, 1552, 1392, 1366, 1302, 1256, 1235, 1171, 1116, 1062, 866, 847, 821, 804, 758, 714, 648, 515, 488, 471, 418; HRMS (ESI): *m/z* calcd for C₂₀H₂₄O₄N₂Se₂+H⁺: 517.0139 [M+H]⁺; found: 517.0146.

2.3 Synthesis of 2,2'-diselanediyldinicotinic acid 5

To a sealed vial loaded with a solution of di-*tert*-butyl 2,2'-diselanediyldinicotinate **4** (100 mg, 0.2 mmol) in CH_2Cl_2 (8 mL), CH_3SO_3H (148 mg, 0.1 mL, 1.6 mmol) was added dropwise at room temperature. The reaction was stirred for 16 h, then quenched by MeOH (10 mL).

The resulting insoluble solid was filtered and washed with MeOH $(2 \times 20 \text{ mL})$ and water (3 × 20 mL) to give pure **5** as a yellow solid (79 mg, 96% yield).

mp: 192-194 °C; ¹H NMR (600 MHz, DMSO-*d*₆) *δ*: 8.55 (*dd*, *J*_{H,H} = 4.8, 1.7 Hz, 2H, *H*_a), 8.22 (*dd*, *J*_{H,H} = 7.7, 1.7 Hz, 2H, *H_c*), 7.31 (*dd*, *J*_{H,H} = 7.7, 4.8 Hz, 2H, *H_b*); ¹³C NMR (150 MHz, DMSO-*d6*) *δ*: 167.8, 161.6, 153.0, 138.7, 125.7, 120.4; FTIR (ATR): *ν* (cm-1): 3140, 2855, 1663, 1569, 1548, 1445, 1421, 1383, 1300, 1238, 1225, 1150, 1120, 1064, 889, 818, 758, 705, 644, 553, 523, 476, 450, 437, 420; HRMS (ESI): m/z calcd for C₁₂H₈O₄N₂Se₂+H⁺: 404.8890 [M+H]⁺; found: 404.8873.

2.4 Synthesis of 2-phenyl-[1,2]selenazolo[5,4-β]pyridin-3(2H)-one 1Ph

To a two-necked 25 mL round-bottomed flask with a suspension of 2,2'-diselanediyldinicotinic acid 5 (80 mg, 0.2 mmol) in SOCl₂ (0.6 mL) under anhydrous condition, dry DMF (2 drops) was added dropwise, then the reaction heated up to reflux and stirred for 6 h. The reaction was cooled down to room temperature and a second portion of SOCl₂ (0.6 mL) and dry DMF (2 drops) added. The solution was heated up to reflux and stirred for 16 h, then the solvents distilled off. To a solution of the resulting acyl chloride derivative in dry CH_2Cl_2 (5 mL), a solution of aniline (20 mg, 0.02 mL, 0.22 mmol) and NEt₃ (44 mg, 0.06 mL, 0.44 mmol) in dry CH_2Cl_2 (0.4 mL) was added dropwise at -15 °C. The reaction was slowly allowed to warm up to room temperature and stirred for 16 hours, then poured in cold water (30 mL) and extracted with CH_2Cl_2 (3 × 30 mL). The combined organic extracts were washed with brine (20 mL), dried over Na₂SO₄, filtered and the solvents evaporated *in vacuo*. The crude was purified by silica gel chromatography (CHCl3/EtOAc 3:1) to give pure **1Ph** as a white solid (25 mg, 45% yield).

mp: 180-182 °C; ¹H NMR (600 MHz, CDCl₃) *δ*: 8.84 (*d*, *J*_{H,H} = 4.5 Hz, 1H, *H_a*), 8.39 (*dd*, *J*H,H = 7.7, 1.7 Hz, 1H, *Hc*), 7.65 (*dm*, *J*H,H = 8.5 Hz, 2H, *Hd*), 7.49 (*d*, *J*H,H = 7.7, 4.5 Hz, 1H, *Hb*), 7.46 (*dd*, *J*_{H,H} = 8.5, 7.5 Hz, 2H, *H*_e), 7.31 (*t*, *J*_{H,H} = 7.5 Hz, 1H, *H_f*); ¹³C NMR (150 MHz, CDCl₃) *δ*: 163.8, 161.7, 152.7, 138.8, 137.6, 129.6, 127.1, 125.7, 124.5, 122.1; 77Se NMR (114 MHz, CDCl3) *δ*: 953.8 (*s*); FTIR (ATR): *ν* (cm-1): 3070, 2921, 2852, 1655, 1638, 1584, 1486, 1452, 1391, 1336, 1262, 1205, 1181, 1141, 1108, 1085, 943, 815, 751, 692, 672, 600, 528, 481; HRMS (ESI): m/z calcd for C₁₂H₈ON₂Se+H⁺: 276.9875 [M+H]⁺; found: 276.9873. Single crystals suitable for X-ray diffraction analysis were grown from slow evaporation of solvent from a CHCl₃/toluene 1:1 solution.

2.5 Synthesis of 2-pentyl-[1,2]selenazolo[5,4-β]pyridin-3(2H)-one 1*Alk*

To a two-necked 25 mL round-bottomed flask with a suspension of 2,2'-diselanediyldinicotinic acid 5 (80 mg, 0.2 mmol) in SOCl₂ (0.6 mL) under anhydrous condition, dry DMF (2 drops) was added dropwise, then the reaction heated up to reflux and stirred for 6 h. The reaction was cooled down to room temperature and a second portion of SOCl2 (0.6 mL) and dry DMF (2 drops) added. The solution was heated up to reflux and stirred for 16 h, then the solvents distilled off. To a solution of the resulting acyl chloride derivative in dry CH_2Cl_2 (5 mL), a solution of amylamine (19 mg, 0.03 mL, 0.22 mmol) and NEt₃ (44 mg, 0.06 mL, 0.44 mmol) in dry CH_2Cl_2 (0.4 mL) was added dropwise at -15 °C. The reaction was slowly allowed to warm up to room temperature and stirred for 16 h, then poured in cold water (30 mL) and extracted with CH_2Cl_2 (3 × 30 mL). The combined organic extracts were washed with brine (20 mL), dried over Na₂SO₄, filtered and the solvents evaporated *in vacuo*. The crude was purified by silica gel chromatography (CHCl₃/EtOAc 4:1) to give pure 1_{Alk} as a white solid (38 mg, 72% yield).

mp: 138-140 °C; ¹H NMR (600 MHz, CDCl₃) *δ*: 8.74 (*dd*, *J*_{H,H} = 4.9, 1.7 Hz, 1H, *H_a*), 8.25 (dd, J_{H,H} = 7.8, 1.7 Hz, 1H, H_c), 7.39 (dd, J_{H,H} = 7.8, 4.9 Hz, 1H, H_b), 3.87 (*t*, J_{H,H} = 7.2 Hz, 2H, *H_d*), 1.78 – 1.73 (*m*, 2H, *H_e*), 1.42 – 1.36 (*m*, 4H, *H_{f,g}*), 0.91 (*t*, *J*_{H,H} = 7.2 Hz, 3H, *H_h*); ¹³C NMR (150 MHz, CDCl₃) δ: 165.2, 161.8, 152.7, 136.5, 123.5, 121.5, 44.6, 30.2, 28.8, 22.3, 13.9; 77Se NMR (114 MHz, CDCl3) *δ*: 897.6 (*s*); FTIR (ATR): *ν* (cm-1): 2950, 2923, 2854, 1641, 1624, 1581, 1562, 1465, 1385, 1308, 1251, 1217, 1167, 1105, 1082, 1051, 820, 755, 698, 666, 519, 490, 443; HRMS (ESI): m/z calcd for C₁₁H₁₄ON₂Se+Na⁺: 293.0164 [M+Na]⁺; found: 293.0167. Single crystals suitable for X-ray diffraction analysis were grown from slow evaporation of solvent from a toluene solution.

2.6 Synthesis of 2-(anthracen-9-yl)-[1,2]selenazolo[5,4-β]pyridin-3(2H)-one 1Anthr

To a two-necked 25 mL round-bottomed flask with a suspension of 2,2'-diselanediyldinicotinic acid 5 (80 mg, 0.2 mmol) in SOCl₂ (0.6 mL) under anhydrous condition, dry DMF (2 drops) was added dropwise, then the reaction heated up to reflux and stirred for 6 h. The reaction was cooled down to room temperature and a second portion of

SOCI₂ (0.6 mL) and dry DMF (2 drops) added. The solution was heated up to reflux and stirred for 16 h, then the solvents were distilled off. To a solution of the resulting acyl chloride derivative in dry CH₂Cl₂ (5 mL), a solution of anthracen-9-amine (43 mg, 0.22 mmol) and NEt₃ (44 mg, 0.06 mL, 0.44 mmol) in dry CH_2Cl_2 (0.4 mL) was added dropwise at -15 °C. The reaction was stirred at 50 °C for 16 h, then poured in cold water (30 mL) and extracted with CH_2Cl_2 (3 × 30 mL). The combined organic extracts were washed with brine (20 mL), dried over Na2SO4, filtered and the solvents evaporated *in vacuo*. The crude was purified by silica gel chromatography (CHCl₃/EtOAc 3:1) to give pure 1_{Anthr} as a pale-yellow solid (30 mg, 41% yield).

mp: > 230 °C; ¹H NMR (600 MHz, CDCl₃) *δ*: 8.89 (*dd*, *J*_{H,H} = 4.9, 1.7 Hz, 1H, *H_a*), 8.63 (*s*, 1H, *Hh*), 8.46 (*dd*, *J*H,H = 7.8, 1.7 Hz, 1H, *Hc*), 8.18 (*d*, *J*H,H = 8.6 Hz, 2H, *Hd*), 7.89 (*d*, *J*H,H = 8.6 Hz, 2H, *Hg*), 7.57 – 7.49 (*m*, 5H, *Hb,e,f*); 13C NMR (150 MHz, CDCl3) *δ*: 165.6, 163.2, 153.8, 137.6, 131.9, 129.7, 129.1, 128.9, 128.6, 127.4, 125.8, 123.0, 122.1, 121.8; FTIR (ATR): *ν* (cm-1): 2979, 2363, 1639, 1579, 1560, 1463, 1409, 1391, 1366, 1233, 1083, 912, 843, 820, 782, 757, 731, 697, 671, 596, 551, 512, 476, 446, 435; HRMS (LD): *m/z* calcd for C₂₀H₁₂ON₂Se⁺: 376.0110 [M]⁺; found: 376.0103. Single crystals suitable for X-ray diffraction analysis were grown from slow evaporation of solvent from a CHCl₃/toluene solution.

2.7 Synthesis of 2-(4-fluorophenyl)-[1,2]selenazolo[5,4-β]pyridin-3(2H)-one 1F

To a two-necked 25 mL round-bottomed flask with a suspension of 2,2'-diselanediyldinicotinic acid 5 (80 mg, 0.2 mmol) in SOCl₂ (0.6 mL) under anhydrous condition, dry DMF (2 drops) was added dropwise, then the reaction heated up to reflux and stirred for 6 h. The reaction was cooled down to room temperature and a second portion of SOCl₂ (0.6 mL) and dry DMF (2 drops) added. The solution was heated up to reflux and stirred for 16 h, then the solvents were distilled off. To a solution of the resulting acyl chloride derivative in dry CH_2Cl_2 (5 mL), a solution of 4-fluoroaniline (61 mg, 52 µL, 0.55 mmol) and NEt₃ (67 mg, 0.08 mL, 0.6 mmol) in dry CH_2Cl_2 (0.4 mL) was added dropwise at -40 °C. The reaction was stirred at room temperature for 8 h, then poured in cold water (30 mL) and extracted with CH_2Cl_2 (3 × 30 mL). The combined organic extracts were washed with brine (20 mL), dried over Na2SO4, filtered and the solvents evaporated *in vacuo*. The crude was purified by silica gel chromatography (CHCl₃/EtOAc 5:1) to give pure 1_F as a pale-yellow solid (13 mg, 23% yield).

mp: 158-160 °C; ¹ H NMR (400 MHz, CDCl3) *δ*: 8.83 (*d*, *J*H,H = 4.5 Hz, 1H, *Ha*), 8.36 (*d*, *J*H,H = 7.7 Hz, 1H, *Hc*), 7.59 (*dd*, *J*H,H = 8.4, 4.8 Hz, 2H, *Hd*), 7.48 (*dd*, *J*H,H = 7.7, 4.5 Hz, 1H, *H_b*), 7.15 (*d*, *J*_{H,H} = 8.8, 8.4 Hz, 2H, *H_e*); ¹⁹F NMR (376 MHz, CDCl₃) *δ*: -113.80 (s, 1F); 13C NMR (100 MHz, CDCl3) *δ*: 161.6, 153.1, 137.3, 127.7, 127.6, 123.6, 122.0, 116.4, 116.2, 110.3 (d, J_{C,F} = 250.1 Hz); FTIR (ATR): *ν* (cm⁻¹): 3052, 2989, 1687, 1644, 1523, 1491, 1463, 1343, 1290, 1223, 1199, 1156, 1111, 1063, 943, 889, 823, 778, 699, 616; HRMS (ESI): *m/z* calcd for C₁₂H₇ON₂FSe+Na⁺: 316.9600 [M+Na]⁺; found: 316.9599.

2.8 Synthesis of 2-(4-methoxyphenyl)-[1,2]selenazolo[5,4-β]pyridin-3(2H)-one 1Met

To a two-necked 25 mL round-bottomed flask with a suspension of 2,2'-diselanediyldinicotinic acid 5 (80 mg, 0.2 mmol) in SOCl₂ (0.6 mL) under anhydrous condition, dry DMF (2 drops) was added dropwise, then the reaction heated up to reflux and stirred for 6 h. The reaction was cooled down to room temperature and a second portion of $SOCl₂$ (0.6 mL) and dry DMF (2 drops) added. The solution was heated up to reflux and stirred for 16 h, then the solvents were distilled off. To a solution of the resulting acyl chloride derivative in dry CH_2Cl_2 (5 mL), a solution of 4-methoxyaniline (28 mg, 0.22 mmol) and NE t_3 (44 mg, 0.06 mL, 0.44 mmol) in dry CH_2Cl_2 (0.4 mL) was added dropwise at -15 °C. The reaction was stirred at room temperature for 16 h, then poured in cold water (30 mL) and extracted with CH_2Cl_2 (3 × 30 mL). The combined organic extracts were washed with brine (20 mL), dried over $Na₂SO₄$, filtered and the solvents evaporated *in vacuo*. The crude was purified by silica gel chromatography (CHCl3/EtOAc 4:1) to give pure **1Met** as a white solid (52 mg, 85% yield).

mp: 133-136 °C; ¹H NMR (400 MHz, CDCl₃) *δ*: 8.81 (*dd, J*_{H,H} = 5.0, 1.6 Hz, 1H, *H_a*), 8.34 (*dd*, *J*H,H = 7.9, 1.6 Hz, 1H, *Hc*), 7.51 (*d*, *J*H,H = 8.8 Hz, 2H, *Hd*), 7.45 (*dd*, *J*H,H = 7.9, 5.0 Hz, 1H, *Hb*), 6.97 (*d*, *J*H,H = 8.8 Hz, 2H, *He*), 3.85 (*s*, 3H, *Hf*); 13C NMR (100 MHz, CDCl3) *δ*: 163.9, 161.8, 158.5, 152.7, 137.2, 131.1, 127.4, 123.9, 121.8, 114.6, 55.6; FTIR (ATR): *ν* (cm-1): 3051, 2989, 2913, 1623, 1599, 1467, 1421, 1387, 1350, 1321, 1284, 1222, 1190, 1136, 1110, 993, 867, 801, 745, 721, 654, 572; HRMS (ESI): m/z calcd for C₁₃H₁₀O₂N₂Se+H⁺: 306.9981 [M+H]⁺; found: 306.9982. Single crystals suitable for X-ray diffraction analysis were grown from slow evaporation of solvent from a CHCl₃/toluene solution.

2.9 Synthesis of 2-(3,4,5-trifluorophenyl)-[1,2]selenazolo[5,4-β]pyridin-3(2H) one 13F

To a two-necked 25 mL round-bottomed flask with a suspension of 2,2'-diselanediyldinicotinic acid 5 (80 mg, 0.2 mmol) in SOCl₂ (0.6 mL) under anhydrous condition, dry DMF (2 drops) was added dropwise, then the reaction heated up to reflux and stirred for 6 h. The reaction was cooled down to room temperature and a second portion of SOCl₂ (0.6 mL) and dry DMF (2 drops) added. The solution was heated up to reflux and stirred for 16 h, then the solvents were distilled off. To a solution of the resulting acyl chloride derivative in dry CH_2Cl_2 (5 mL), a solution of 3,4,5-trifluoroaniline (81 mg, 0.55 mmol) and NEt₃ (67 mg, 0.08 mL, 0.6 mmol) in dry CH_2Cl_2 (0.4 mL) was added dropwise at -40 °C. The reaction was stirred at room temperature for 10 h, then poured in cold water (30 mL) and extracted with CH_2Cl_2 (3 × 30 mL). The combined organic extracts were washed with brine (20 mL), dried over Na2SO4, filtered and the solvents evaporated *in vacuo*. The crude was purified by silica gel chromatography (CHCl3/EtOAc 3:1) to give pure **13F** as a white solid (14 mg, 21% yield).

mp: 176-178 °C; ¹ H NMR (400 MHz, CDCl3) *δ*: 8.83 – 8.82 (*m*, 1H, *Ha*), 8.34 – 8.32 (*m*, 1H, *Hc*), 7.52 – 7.49 (*m*, 1H, *Hb*), 7.39 – 7.36 (*m*, 2H, *Hd*); 19F NMR (659 MHz, DMSO-*d6*) *δ*: -134.28 (*d*, $J_{F,F}$ = 22.08 Hz, 2F), -164.45 (*t*, $J_{F,F}$ = 22.08 Hz, 1 F); ¹³C analysis is missing due to the poor solubility of the targeted molecule in several solvents screened; FTIR (ATR): *ν* (cm⁻¹): 3098, 2923, 2887, 1710, 1643, 1552, 1491, 1413, 1392, 1337, 1289, 1238, 1154, 1113, 1062, 983, 907, 826, 796, 701, 592, 478, 452; HRMS (ESI): m/z calcd for C₁₂H₅ON₂F₃Se+H⁺: 330.9592 [*M*+H] + ; found: 330.9594.

pyridin-3(2H)-one 13Met

To a two-necked 25 mL round-bottomed flask with a suspension of 2,2'-diselanediyldinicotinic acid 5 (80 mg, 0.2 mmol) in SOCl₂ (0.6 mL) under anhydrous condition, dry DMF (2 drops) was added dropwise, then the reaction heated up to reflux and stirred for 6 h. The reaction was cooled down to room temperature and a second portion of SOCl₂ (0.6 mL) and dry DMF (2 drops) added. The solution was heated up to reflux and stirred for 16 h, then the solvents were distilled off. To a solution of the resulting acyl chloride derivative in dry CH_2Cl_2 (5 mL), a solution of 3,4,5-trimethoxyaniline (40 mg, 0.22 mmol) and $NEt₃$ (44 mg, 0.06 mL, 0.44 mmol) in dry CH₂Cl₂ (0.4 mL) was added dropwise at -15 °C. The reaction was stirred at room temperature for 16 h, then poured in cold water (30 mL) and extracted with CH_2Cl_2 (3 \times 30 mL). The combined organic extracts were washed with brine (20 mL), dried over $Na₂SO₄$, filtered and the solvents evaporated *in vacuo*. The crude was purified by silica gel chromatography (CHCl3/EtOAc 4:1) to give pure **13Met** as a white solid (64 mg, 88% yield).

mp: 141-143 °C; ¹ H NMR (400 MHz, CDCl3) *δ*: 8.83 (*d*, *J*H,H = 4.7 Hz, 1H, *Ha*), 8.36 (*d*, *J*H,H = 7.8 Hz, 1H, *Hc*), 7.49 (*dd*, *J*H,H = 7.8, 4.7 Hz, 1H, *Hb*), 6.85 (*s*, 2H, *Hd*), 3.89 (*s*, 6H, *He*), 3.87 (*s*, 3H, *Hf*); 13C NMR (100 MHz, CDCl3) *δ*: 163.9, 161.8, 153.6, 152.9, 137.3, 137.2, 134.0, 123.9, 122.0, 103.7, 60.9, 56.3; FTIR (ATR): *ν* (cm-1): 3030, 2984, 2916, 1621, 1589, 1515, 1438, 1367, 1329, 1251, 1222, 1167, 1098, 1003, 965, 891, 744, 639, 588; HRMS (ESI): *m/z* calcd for $C_{15}H_{14}O_4N_2Se+H^+$: 367.0192 $[M+H]^+$; found: 367.0197. Single crystals suitable for X-ray diffraction analysis were grown from slow evaporation of solvent from a CHCl₃/toluene solution.

3. NMR-HRMS Spectroscopic characterization (1 H, 13C, 19F, 77Se, HRMS)

3.1 Characterization of 3

Figure S2: 175 MHz ¹³C NMR in CDCl₃ of molecule 3.

Figure S3: HRMS-ESI mass spectrum of molecule 3 in the positive ion mode.

3.2 Characterization of 4

Figure S4: 600 MHz 1H NMR in CDCl3 of molecule 4.

Figure S6: HRMS-ESI mass spectrum of molecule 4 in the positive ion mode. The peak at 1053 corresponds to the dimeric species.

Figure S9: HRMS-ESI mass spectrum of molecule 5 in the positive ion mode.

3.4 Characterization of 1Ph

Figure S10: 600 MHz 1H NMR in CDCl3 of molecule 1Ph.

Figure S12: 114 MHz 77Se NMR in CDCl3 of molecule 1Ph.

Figure S13: HRMS-ESI mass spectrum of molecule 1Ph in the positive ion mode. The peak at 574 corresponds to the dimeric species.

3.5 Characterization of 1Alk

Figure S14: 600 MHz 1H NMR in CDCl3 of molecule 1Alk.

 1_{Alk} $(114 MHz, CDCl₃)$

MW

940 930 920 910 900 890 880 870 860 850 840 830 960 980 970 950 820 810 ppm

Figure S17: HRMS-ESI mass spectrum of molecule $1_{A/k}$ *in the positive ion mode. The peak at 563 corresponds to the dimeric species.*

3.6 Characterization of 1Anthr

Figure S18: 600 MHz 1H NMR in CDCl3 of molecule 1Anthr.

Figure S20: HRMS-LD mass spectrum of molecule 1_{Anth} *in the positive ion mode.*

Figure S22: 376 MHz 19F NMR in CDCl3 of molecule 1F.

Figure S24: HRMS-ESI mass spectrum of molecule 1_F *in the positive ion mode. The peak at 610 corresponds to the dimeric species.*

Figure S26: 100 MHz 13C NMR in CDCl3 of molecule 1Met.

Figure S27: HRMS-ESI mass spectrum of molecule 1_{Met} *in the positive ion mode. The peak at 635 corresponds to the dimeric species.*

3.9 Characterization of 13F

Figure S28: 400 MHz 1H NMR in CDCl3 of molecule 13F.

Figure S30: HRMS-ESI mass spectrum of molecule 13F in the positive ion mode. The peak at 778 corresponds to the dimeric species.

3.10 Characterization of 13Met

Figure S32: 150 MHz 13C NMR in CDCl3 of molecule 13Met.

Figure S33: HRMS-ESI mass spectrum of molecule 13Met in the positive ion mode.

4. Crystallographic data

Table S1. Crystal data and structure refinement for 1_{Ph} (2333906).

Table S2. Crystal data and structure refinement for **1Alk** (2333763). Empirical formula C₁₁H₁₄N2OSe

Formula weight 269.20 Crystal system Monoclinic Space group C 2/c

Volume 2639.0(15) \hat{A}^3 $Z \hspace{2.5cm} 8$ Density (calculated) 1.355 mg/m³ Absorption coefficient 3.683 mm⁻¹ F(000) 1088

Temperature 100(2) K Wavelength 1.54184 Å Theta range for data collection 7.032 to 143.012°. Reflections collected 18255 Independent reflections 2238 [R(int) = 0.0476] Completeness to theta = 25.242° 94.8 %

Absorption correction multi-scan Data / restraints / parameters 2433 / 0 / 137 Goodness-of-fit on F^2 1.191 Extinction coefficient n/a Largest diff. peak and hole 0.776 and -0.698 e·Å $^{-3}$

Unit cell dimensions $a = 25.268(7)$ Å $\alpha = 90^{\circ}$. b = 4.5670(12) Å $\beta = 95.76(3)^\circ$. c = 22.985(9) Å $y = 90^\circ$. ∩ Sé 1_{Alk} Crystal size $0.100 \times 0.057 \times 0.020$ mm³ **Data collection** Index ranges -18<=18<=18<=28, -5<=k<=4, -26<=l<=28 **Refinement** Refinement method Full-matrix least-squares on $F²$ Final R indices [I>2sigma(I)] R1 = 0.0476, wR2 = 0.1113 R indices (all data) R1 = 0.0519, wR2 = 0.1145

Table S3. Crystal data and structure refinement for **1Anthr** (2333913). Empirical formula C₂₀H₁₂N₂OSe Formula weight 375.53 Crystal system Triclinic

Space group example and the P-1

Z 2 Density (calculated) 1.262 mg/m³ Absorption coefficient 1.907 mm⁻¹ F(000) 376 Crystal size $0.080 \times 0.080 \times 0.060$ mm³

- Temperature 100(2) K Wavelength 0.71073 Å Theta range for data collection 4.872 to 60.292°. Reflections collected 43351 Independent reflections 5049 [R(int) = 0.0257] Completeness to theta = 25.242° 99.7 %
- Absorption correction Refinement method Data / restraints / parameters Goodness-of-fit on $F²$. Final R indices $[1>2$ sigma $(1)]$ R indices (all data) Extinction coefficient Largest diff. peak and hole
- Unit cell dimensions $a = 9.1044(2)$ Å $\alpha = 70.8921(11)^\circ$. Volume 987.60(5) \AA^3
	-

Data collection

Index ranges -12<=12, -12<=12, -13<=k<=13, -17<=l<=17

Refinement

Table S4. Crystal data and structure refinement for 1_{Met} (2333910).

Volume $1146.1(2)$ \AA^3 Z 4 Density (calculated) 1.769 mg/m³ Absorption coefficient 3.270 mm⁻¹ F(000) 608 Crystal size $0.030 \times 0.133 \times 0.320$ mm³ Temperature 100(2) K Wavelength 0.71073 Å Theta range for data collection 1.774 to 31.373°.

Reflections collected 25886

Completeness to theta = 25.242° 97.8 %

Absorption correction multi-scan Max. and min. transmission 0.942 and 0.731 Data / restraints / parameters 3680 / 0 / 164 Goodness-of-fit on $F²$ 1.324 Extinction coefficient n/a Largest diff. peak and hole 0.575 and -0.819 e·Å 3

b = 13.8376(14) Å $\beta = 94.648(8)^\circ$. c = 20.631(2) Å $y = 90^\circ$. \circ 1_{Met} **Data collection** Index ranges -5<=h<=5, -20<=k<=18, -29<=l<=29 Independent reflections 3652 [R(int) = 0.0364] **Refinement** Refinement method $F²$ Full-matrix least-squares on $F²$ Final R indices [I>2sigma(I)] R1 = 0.0806, wR2 = 0.2052

R indices (all data) R1 = 0.1070, wR2 = 0.2052

Table S5. Crystal data and structure refinement for 1_{3F} (2223676).

Volume $542.6(2)$ \AA^3 Z 2 Density (calculated) 2.015 mg/m³ Absorption coefficient 3.338 mm⁻¹ F(000) 320 1_{3F} Crystal size $0.020 \times 0.050 \times 0.100$ mm³ **Data collection** Temperature 100(2) K Wavelength 0.700 Å Theta range for data collection 0.981 to 25.949°. Index ranges -4<=h<=4, -14<=k<=14, -15<=l<=15 Reflections collected 2742 Completeness to theta = 25.242° 98.1 % **Refinement** Absorption correction multi-scan Max. and min. transmission 0.936 and 0.731 Refinement method $F²$ Full-matrix least-squares on $F²$ Data / restraints / parameters 2182 / 0 / 173 Goodness-of-fit on $F²$ 1.113 Final R indices [I>2sigma(I)] R1 = 0.1056, wR2 = 0.2555 R indices (all data) R1 = 0.1519, wR2 = 0.2793 Extinction coefficient n/a Largest diff. peak and hole 1.860 and -1.566 e·Å -3

Table S6. Crystal data and structure refinement for **13Met** (2333762).

33

1_{3Met}

Density (calculated) 1.732 mg/m³ Absorption coefficient 2.701 mm⁻¹ F(000) 736

Temperature 100(2) K Wavelength 0.71073 Å Theta range for data collection 2.26 to 27.50°. Reflections collected 4572 Completeness to theta = 25.242° 96.4 %

Absorption correction multi-scan Max. and min. transmission 0.922 and 0.724 Data / restraints / parameters 3302 / 0 / 203 Goodness-of-fit on F² 1.117 Extinction coefficient n/a Largest diff. peak and hole 0.925 and -1.277 e·Å 3

Crystal size $0.010 \times 0.067 \times 0.180$ mm³ **Data collection** Index ranges -6<=h<=6, -20<=k<=21, -33<=l<=34 **Refinement** Refinement method $F²$ Full-matrix least-squares on $F²$ Final R indices [I>2sigma(I)] R1 = 0.0347, wR2 = 0.0212 R indices (all data) R1 = 0.06500, wR2 = 0.0843

Figure S34: Molecular packing of 1Ph, held together by a) Se∙∙∙N ChBs (pointed out with dashed blue lines) and HBs (highlighted with dotted black lines); b) crystal packing along b axis, expanded through Se∙∙∙π vdW interactions. Crystallization solvent: THF. Space group: Pca21.

Figure S35: Molecular packing of a) 1Met, forming a 2D non-covalent polymer by means of double Se∙∙∙N ChBs (pointed out with dashed blue lines) and HBs (highlighted with dotted black lines); crystal structure of b) 13Met, developed through multiple HBs (highlighted with dotted black lines). Crystallization solvent: CHCl3 for 1Met, THF for 13Met. Space group: Pca21 for 1Met, P21/n for 13Met.

Figure S36: Double linear polymeric chain composing the molecular packing of 1_{Alk}, driven by concurrent Se…N ChBs (highlighted with dashed blue lines) and C∙∙∙C contacts (outlined with dotted black lines). Crystallization solvent: CHCl3/EtOAc. Space group: C2/c.

Figure S37: a) Self-assembly of 1Anthr in doubly Se∙∙∙N chalcogen-bonded dimers and b) molecular packing developed through π∙∙∙π stacking between the anthracene rings. Crystallization solvent: CHCl3/toluene. Space group: P-1.

5. 1H and 77Se NMR measurements

5.1 Diluition experiments

Dimerization of compounds 1_i was investigated by dilution experiments in CDCl₃ at 298 K. The chemical shift data of the 77 Se resonance were fitted with Dynafit software packages^[42] using a 1:1 dimerization model and the obtained dimerization constants (K_d) are reported in the Figures below. The chemical shift data of the three pyridyl protons (H_a, H_b, end H_c) were fitted with Dynafit using a global fit approach in which the data of the three protons are fitted contemporaneously to the 1:1 dimerization model. The global fit approach increases the accuracy of the fitting results and it is particularly useful in this case where the chemical shift variations upon dilution are really small. The Figures below reports the obtained dimerization constants (K_d) and the fitting curve relative to proton Ha which is the one that shows the highest chemical shift variation upon dilution. In general, the dimerization constants obtained from the fitting of the ⁷⁷Se resonance and those from the pyridyl protons are in good agreement, with the latter being slightly higher in some cases. However, due to the larger chemical shift variation, the readout errors on the ⁷⁷Se signals are less prone to affect the fitting results, allowing the collection of a more consistent and wide set of results. Accordingly, this set of data is reported in the manuscript. In any case, the same trend in the K_d values is obtained from the ¹H titrations.

Figure S38: Absorption spectra of 1Ph upon decreasing concentration, recorded in benzene at 298 K.

Figure S39: 600 MHz 1H NMR overlapped spectra of 1Ph diluted solutions in CDCl3 at 298 K.

Figure S40: 115 MHz ⁷⁷Se NMR overlapped spectra of 1_{Ph} diluted solutions in CDCI₃ at 298 K.

Figure S41: Experimental chemical shift from a) 1H NMR and b) 77Se NMR dilution experiments in CDCl3 vs concentration of 1Ph, fitted to a 1:1 dimerization equilibrium.

Figure S42: 600 MHz 1H NMR overlapped spectra of 1F diluted solutions in CDCl3 at 298 K.

Figure S43: 600 MHz ¹H NMR zoomed overlapped spectra of 1_F diluted solutions in CDCl₃ at 298 K.

Figure S44: 115 MHz ⁷⁷Se NMR overlapped spectra of 1_F diluted solutions in CDCl₃ at 298 K.

Figure S45: Experimental chemical shift from a) 1H NMR and b) 77Se NMR dilution experiments in CDCl3 vs concentration of 1F, fitted to a 1:1 dimerization equilibrium. The four points at higher concentrations were not included in the fitting process because they largely deviate from the binding isotherm. This is likely due to the low solubility of 1F in CDCl3.

Figure S46: 600 MHz ¹H NMR overlapped spectra of 1_{Met} diluted solutions in CDCl₃ at 298 K.

Figure S48: 115 MHz ⁷⁷Se NMR overlapped spectra of 1_{Met} diluted solutions in CDCI₃ at 298 K.

Figure S49: Experimental chemical shift from a) 1H NMR and b) 77Se NMR dilution experiments in CDCl3 vs concentration of 1Met, fitted to a 1:1 dimerization equilibrium. In the case of the 77Se NMR dilution experiment, the three points at higher concentrations were not included in the fitting process because they largely deviate from the binding isotherm. This is not observed in the case of the 1H NMR experiments and, therefore, it is likely due to scattering in the measurements of the 77Se chemical shifts.

Figure S50: 600 MHz ¹H NMR overlapped spectra of $1_{3\text{Met}}$ diluted solutions in CDCI₃ at 298 K.

Figure S51: 600 MHz ¹H NMR zoomed overlapped spectra of 1_{3Met} diluted solutions in CDCl₃ at 298 K.

Figure S52: 115 MHz ⁷⁷Se NMR overlapped spectra of 1_{3Met} diluted solutions in CDCI₃ at 298 K.

Figure S53: Experimental chemical shift from a) 1H NMR and b) 77Se NMR dilution experiments in CDCl3 vs concentration of 13Met, fitted to a 1:1 dimerization equilibrium.

Figure S54: 600 MHz 1H NMR overlapped spectra of 1Alk diluted solutions in CDCl3 at 298 K.

Figure S56: Experimental chemical shift from a) 1H NMR and b) 77Se NMR dilution experiments in CDCl3 vs concentration of 1Alk, fitted to a 1:1 dimerization equilibrium.

Figure S57: 600 MHz ¹H NMR overlapped spectra of 1_{Ph} *diluted solutions in CD₂Cl₂ at 298 K.*

Figure S58: 600 MHz ¹H NMR zoomed overlapped spectra of 1_{Ph} diluted solutions in CD₂Cl₂ at 298 K.

Figure S59: 115 MHz ⁷⁷Se NMR overlapped spectra of 1_{Ph} diluted solutions in CD₂Cl₂ at 298 K.

Figure S60: Experimental chemical shift from a) ¹H NMR and b) ⁷⁷Se NMR dilution experiments in CD₂Cl₂ <i>vs concentration of 1Ph at 298 K, fitted to a 1:1 dimerization equilibrium.

Note for the reader: For the screening of 1_{Ph} in CD₂Cl₂/toluene 1:1, to avoid the overlapping of the signals of the solvent with those of the molecule in the 1 H NMR, only ⁷⁷Se NMR measurements were performed.

Figure S61: 115 MHz ⁷⁷Se NMR overlapped spectra of 1_{Ph} diluted solutions in CD₂Cl₂/toluene at 298 K.

Figure S62: Experimental chemical shift from a) ⁷⁷Se NMR dilution experiments in CD₂Cl₂/toluene vs *concentration of 1Ph at 298 K, fitted to a 1:1 dimerization equilibrium; b) plotted 77Se NMR recorded in CDCl3, CD2Cl2 and CD2Cl2/toluene solutions as function of concentration.*

While the K_d values obtained in CDCl₃ and in CD₂Cl₂ are very similar (51 ± 4 M⁻¹ and 45 ± 4 M⁻¹, respectively) the K_d measured in the CD₂Cl₂/toluene is slightly lower (23 ± 4 M⁻¹). However, this is likely due to some deviation of the experimental points due to the limited solubility of 1_{Ph} in this solvent mixture. As a matter of fact, simply excluding the three points measured at the highest concentrations of 1_{Ph} a K_d value of 51 \pm 16 M⁻¹ is obtained. Moreover, plotting all the ⁷⁷Se chemical shift recorded from dilution experiments in the different solvents reveals their almost complete overlap, globally excluding a solvent effect in the ChB-driven dimerization of 1_{Ph} (Fig. S62b).

5.2 Variable Temperature (VT) experiments

Figure S64: 115 MHz variable temperature (VT) ⁷⁷Se NMR overlapped spectra of 20 mM 1_{Ph} solutions in CDCl₃.

Figure S65: 600 MHz variable temperature (VT) ¹H NMR zoomed overlapped spectra of 20 mM 1_{Alk} solutions in *CDCl3.*

Figure S66: 115 MHz variable temperature (VT) ⁷⁷Se NMR overlapped spectra of 20 mM 1_{AJK} solutions in CDCl₃.

Figure S67: van't Hoff plot for the dimerization equilibrium of 20 mM solutions of a) 1_{Ph} *and b)* 1_{Alk} *in CDCl₃, determined in relation to 77Se NMR spectra. Fitting parameters: r2= 0.9993 for 1Ph; r2= 0.9989 for 1Alk.*

Table S7: Thermodynamic parameters obtained from ⁷⁷Se NMR investigations of 1_{Ph} and 1_{Alk}.

	From VT experiments				From dilutions	
		ΔH (KJ·mol ⁻¹) ΔS (J·K ⁻¹ ·mol ⁻¹) ΔG (KJ·mol ⁻¹) K (M ⁻¹)			ΔG (KJ·mol ⁻¹)	$K (M^{-1})$
1_{Ph}	-35.18	-93.78	-7.22	18	-9.88	51
1_{Alk}	-27.45	-81.46	-3.16	36	-3.89	48

6. Computational studies

6.1. Density Functional Theory (DFT) calculations

The DFT calculations reported in this work have been performed via the mixed Gaussian and plane waves (GPW) method implemented in the CP2K package.^[43]

The choice of the exchange-correlation (XC) functional, which is always an important consideration when investigating molecular interactions, is especially controversial when it comes to the description of chalcogenide bonds. For instance, the work of *Bickelhaupt*^[44] recommends the usage of hybrid functionals, particularly B3LYP, as opposed to the addition of dispersion corrections to non-hybrid functionals – which might lead to overestimate the strength of chalcogen bonds. On the other hand, *Goerigk* argues against the usage of B3LYP and recommends instead dispersion-corrected functionals such as the PW6B95-D3.[45] To address this conundrum, we have simply chosen to explore different XC functionals, namely PBE;^[46] PBE-D3,^[47] which feature a dispersion correction; vdW-DF69,^[48] which is a fully selfconsistent, nonlocal XC functional and $HSE06$,^[49] which is a hybrid functional. Goedecker-type pseudopotentials^[50] with four, one, six, five and six valence electrons for C, H, O, N and Se respectively have been employed. The Kohn-Sham orbitals were expanded in a triple-zeta valence plus two sets of polarisation functions (TZV2P) Gaussian-type basis set. The plane wave cutoff for the finest level of the multi-grid^[43] has been set to 400 Ry to

efficiently solve the Poisson equation within periodic boundary conditions using the Quickstep scheme.^[43] Brillouin zone integration was restricted to the supercell Gamma point. We have found that a cubic simulation box (edge = 40 Å), introduces \sim 20 Å of vacuum between the periodic replica of the dimers in each direction, which is sufficient to converge the total energy to 2 meV/atom.

6.2 ALMO-EDA calculations

The Absolutely Localized Molecular Orbitals Energy Decomposition Analysis (ALMO-EDA) is a computational framework that can be used to investigate the different contributions to the total interaction energies we have discussed in the previous section. This framework has been extensively reviewed elsewhere^[51] and it is readily available within the CP2K package. For the purposes of this work, it suffices to say that within the ALMO-EDA formalism the total interaction energy E_{Tot} between two molecules can be written as:

$$
E_{Tot} = E_{Frz} + E_{Pol} + E_{Cov}.
$$

The frozen density term (E_{Frz}) is defined as the energy change that corresponds to bringing infinitely separated molecules into the dimer geometry without any relaxation of the molecular orbitals (MOs) on the monomers. The polarization energy (E_{pol}) is defined as the energy lowering due to the *intra*molecular relaxation of each molecule's ALMOs within the field of all other molecules in the system. The remaining portion of the total interaction energy is the electron delocalization or charge-transfer energy term (E_{cov}) , which is calculated as the energy difference between the state formed from the polarized ALMOs and the state constructed from the fully optimized delocalized MOs. This framework also gives access to the actual charge transfer (CT) contribution, which is computed as the degree of electron relaxation from the polarized state to the delocalized state. By contrast, population analysis methods include not only the "true" CT, but also the separate effect of partitioning the charge distribution of the polarized pre-CT state. Thus, the key advantage of the ALMO approach is that it shows the electron transfer associated with the energy lowering due to dative interactions exclusively.

Crucial to the correct assessment of the interaction energy is an estimate of the BSSE, which is not introduced when calculating the frozen density and the polarization energy contributions (i.e., E_{Frz} and E_{pol}) because constrained ALMO optimization prevents electrons on one molecule from borrowing the atomic orbitals (AOs) of other molecules to compensate for the incompleteness of their own AOs. However, the BSSE does enter the charge transfer terms (i.e., E_{Cov}) since both the BSSE and charge transfer result from the same physical phenomenon of delocalization of fragment MOs. Therefore, these terms are inseparable from each other when finite Gaussian basis sets are used to describe fragments at finite spatial separation.

Figure S68: a) Molecular structures of the three Pyrselen derivatives we have considered for our DFT calculations; b) Interaction energies for the different dimers, computed via different XC functionals (see text); c) Charge transfer (CT) for the different dimers, as a function of the intermolecular distance (units of milli electrons, me); d) Contributions to the total interaction energy (Tot) for the different dimers as a function of the intermolecular distance; Frz, Pol and Cov refer to the frozen density, polarization, and covalent contribution terms, respectively (see text); the (BSSE) results include the basis set superposition error (see ESI) as well.

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