

1 **Exploring cross-taxon congruence between carabid beetles (Coleoptera: Carabidae) and**
2 **vascular plants in sites invaded by *Ailanthus altissima* versus non-invaded sites: the**
3 **explicative power of biotic and abiotic factors**

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15

16 **Abstract**

17 *Ailanthus altissima* is one of the most global widespread invasive alien species and its effect as
18 habitat transformer requires detailed investigations. In particular, its invasion in natural ecosystems
19 and its effect on local fauna should be evaluated and described. With this purpose, the identification
20 of surrogate taxa would be an important tool in order to define the impact of this invader on
21 different habitats. Here, we evaluated cross-taxon congruence to quantify the strength of plant
22 species composition in predicting multivariate patterns in carabid beetle assemblages, based on data
23 from 20 sites divided in invaded and non-invaded habitats located in the Karst area, North-east
24 Italy. We considered the habitat type (representing vegetation stages from grassland to forests) in
25 order to evaluate the impact of *A. altissima* on carabid beetles along the vegetation succession. We
26 found 28 carabid beetles and 173 plant species. Our analyses showed that plant species composition
27 had a valuable predictive accuracy, based on the interplay among environmental variables, soil
28 parameters and vegetation structure. Native vegetation and habitat type were the most important
29 factors influencing carabid beetles and plant species composition. Furthermore, 33% of the total
30 explained variation of carabid beetles assemblages (variance partition based on RDA analysis) was
31 due to the independent effect of environment. We proved cross-taxon congruence between carabid
32 beetles and plants along the successional gradient of vegetation (habitat type). In particular, we
33 attested that both communities present more species differentiation among non-invaded vegetation

34 and, in the meantime, in plots where *A. altissima* was present, anthropic and disturbed plants and
35 carabid beetle species were prevalent. As a conclusion, plants can be effectively used as a
36 surrogate taxon in the evaluation of the effect of *A. altissima* invasion in the Karst area.

37

38 **Keywords**

39 Carabidae; Co-correspondence analysis; Invasive species; Plant community; Surrogate taxon;
40 Variation partitioning.

41

42 **1. Introduction**

43 The study of non-native species (or exotics; Sax, 2002a, 2002b; Hejda et al., 2009) became very
44 popular in the last twenty years (Briggs, 2013; Díez et al., 2014; Tordoni et al. 2018), and recently
45 this field underwent an important shift in research priorities (Pyšek, 2008). In fact, the impact of
46 introduced species outside their native range by human-mediated activities or disturbances is
47 increasing in an era of globalization (Grigorescu, 2016). The heart of invasion science is the
48 realization that biological invasions are not only a biological phenomenon: the human dimension of
49 invasions is a fundamental component in the social-ecological systems, in which invasions need to
50 be understood and managed (Wilson et al., 2016). Several studies highlighted how invasive alien
51 species (hereafter IAS) may threaten human well-being (Schlaepfer et al., 2010) as well as affect
52 natural ecosystems in several ways (see Vilà et al., 2011 for a meta-analysis); for instance, it has
53 been observed that IAS may cause loss or modification of biological diversity (Hejda et al., 2009),
54 modifying also native species extinction probabilities, and habitat structure (Blackburn et al., 2014).

55 A detailed knowledge of the biology and ecology of individual invasive species is an important tool
56 in the quest for a better understanding of invasion phenomena (Pyšek, 2008) but unfortunately, for
57 the vast majority of non-native species, the consequences of invasion are not yet fully understood
58 (Jeschke et al., 2014) or realized (the so-called ‘invasion debt’, Rouget et al., 2015).

59 An important subset of IAS comprehends tree species able to form mono-specific stands in invaded
60 areas, and that are therefore major drivers of environmental change (Richardson, 2011). One of the
61 most important global colonizing tree is *Ailanthus altissima* (Mill.) Swingle (Kowarik and Saümel,
62 2007; Motard et al., 2011; Sladonja et al., 2015). *A. altissima* is a deciduous tree native to China
63 that belongs to the *Simaroubaceae* family. It arrived in Europe at the mid-XVIII century in Paris
64 (Enescu et al., 2016). Thanks to its fast expansion, it is considered as invasive in the most part of

65 Europe (Motard et al., 2011; Petruzzellis et al. 2019a,b), and it is cited as a concern for biodiversity
66 according to the Global Invasive Species Database (GISD, 2017). The first reason of this tree
67 success is the vegetative reproduction ability in forming dense clonal stands; the second reason is
68 the sexual reproduction. The species is common not only in disturbed urban and anthropic areas
69 (Vila et al., 2006; Motard et al., 2011; McAvoy et al., 2012; Motard et al., 2015; Enescu et al.,
70 2016), but can also penetrate forests (Motard et al., 2011). *A. altissima* produces allelopathic
71 compounds and has an insecticidal activity (Tsao et al., 2002; Motard et al., 2011 and 2015; Enescu
72 et al., 2016); moreover, its pollen is allergenic (Ballero et al., 2003). It has been shown that this
73 species can rapidly transform open ecosystems such as meadows or old fields into closed stands
74 (Kowarik and Böcker, 1984; Kowarik and Saümel, 2007) promoting impoverishment of biological
75 diversity through competition (Motard et al., 2011). Moreover, it spreads and displaces native
76 vegetation thanks to a huge canopy cover and due to a large amount of root suckers; its allelopathy
77 results in a significant lower floristic diversity respect to native adjacent zones (Motard et al., 2011).
78 An earlier study in forests colonized by *A. altissima* suggests that an increasing density of this plant
79 is associated also with a lower soil microbial activity, with decreasing abundance of Acari,
80 Collembola, Coleoptera, and terrestrial Gastropoda. Contrarily, increasing *A. altissima* density is
81 linked to greater abundances of Lumbricidae and coprophagous Coleoptera (Motard et al., 2015).
82 However, it is still unclear whether this impact is due to the direct action of *A. altissima* by
83 phytotoxicity of the root system, or to the transformation of soil and litter fauna (Motard et al.,
84 2011).

85 Due to the lack of information concerning the impact of *A. altissima* on terrestrial arthropods, we
86 addressed the issue of the impact of this tree on carabid beetles (Coleoptera, Carabidae)
87 assemblages in different open to forested habitats.

88 The importance of Invertebrates as bioindicators in conservation planning (Kremen et al., 1993;
89 McGeoch, 1998; Andersen et al., 2002; Hodkinson and Jackson, 2005) is well known, since they
90 are ubiquitous, a taxon-rich and dominant group of organisms throughout the world (Wilson, 1987).
91 The carabid beetles represent the fourth largest family in Coleoptera (Löwei and Sunderland, 1996).
92 Carabids are a group of Insects with terrestrial habits (Löwei and Sunderland, 1996; Koivula et al.,
93 1999; Moraes et al., 2013) mainly characterized by carnivorous species (Marinoni et al., 2001;
94 Marinoni, 2001), even though omnivorous exceptions are known for some tribes (Talarico et al.,
95 2016). Due to their adaptations, ecological requirements rather than habitat features have been
96 claimed to influence carabid communities; in fact, in this group, structural features may be suitable
97 or adapted to perform in several different habitats (Ribera et al., 2001). Most carabids are relatively

98 long-lived, and many species do not show any strong seasonal fluctuations, allowing useful
99 sampling activities, performed using pitfall traps (Desender et al., 1994; Luff, 1975; Lee and
100 Albajes, 2016), to be carried out in relatively short periods (Lindroth, 1974; Ings and Hartley,
101 1999). For these reasons, they represent an important and suitable study group in ecological
102 research, especially for what concerns standardized samplings (Lee and Albajes, 2016; Zhang et al.,
103 2017). Studies concerning carabids have addressed their diversity as indicators in distinct topics, for
104 example: effect of environmental modifications (e.g. Thiele; 1977; Niemelä et al., 1993; Lövei and
105 Sunderland, 1996; Villa-Castillo and Wagner, 2002; Kotze et al., 2011), environmental integrity
106 (Taylor and Dorann, 2001); disturbance gradients (da Silva et al., 2008), effects of alien species
107 (Martínez et al., 2009; Buchholz et al., 2015). Vascular plants can be potentially considered a good
108 proxy for invertebrates and have been widely used for this purpose due to their well-known
109 ecology, their relatively ease in identification, and sensitivity to environmental changes (e.g.
110 Sætersdal et al., 2003; Schaffers et al., 2008; Gioria et al., 2010; Maccherini et al., 2012).

111 The diversity of different taxa is influenced by their spatial concordance and by the strength of this
112 association, the so-called cross-taxon congruence, which depends on the studied taxonomic groups,
113 on the scale of analysis (Toranza and Arim, 2010) and on the type of data used (Santi et al., 2016).
114 Cross-taxon congruence analysis can be a suitable tool to elucidate spatio-temporal correlation in
115 patterns of species richness and/or diversity of organisms using surrogate taxa as potential
116 biodiversity indicators (e.g. Margules and Pressey, 2000; Sætersdal et al., 2003). In this way,
117 environmental parameters could be effectively used and potentially also be directly managed in the
118 context of biodiversity conservation to make biodiversity monitoring and conservation planning
119 more efficient (Su et al., 2004; Oertli et al., 2005; Gioria et al., 2011; Barbato et al., 2019). Thus,
120 surrogate taxa could be identified based on the direct biotic interactions such as trophic
121 relationships between target and surrogate taxa (Castagneyrol et al., 2012; Westgate et al., 2014).

122 Generally, cross-taxon congruence is well-connected with similarities in the response patterns of
123 taxa to changes in environmental gradients, biotic interactions or even in their biogeographical
124 history (Pearson and Carroll, 1999; Su et al., 2004; Duan et al., 2016). Despite some criticisms (e.g.
125 Westgate et al., 2014), the use of surrogate taxa to predict community patterns remains a very useful
126 tool especially in case of limited resources or if knowledge gaps make difficult a complete species
127 inventory. It should be also considered that cross-taxon congruence can be promoted by longer
128 disturbance gradient, increasing species characteristic of low or high disturbance levels (Rooney et
129 al., 2014). Nonetheless, the relative contributions of environmental abiotic drivers and biotic
130 interactions in cross-taxon congruence still remains widely unknown (Gioria et al., 2011).

131 In this study, we aimed at evaluating the impact of *A. altissima* invasion on carabid beetles species
132 composition and assessing the role of vascular plants as a possible surrogate group for the
133 distributional and diversity patterns of carabid beetles in North Adriatic Karst (hereafter NAK).
134 Furthermore, the joint and independent effects of spatial location, abiotic variables, non-invaded
135 and invaded vegetation structure, have been also tested to understand their explanatory role in
136 shaping the observed congruence patterns. Specifically, we aimed at: (1) assessing cross-taxon
137 congruence between carabid beetle and vascular plant compositions along a successional gradient of
138 vegetation (from grassland to forest) in sites invaded and non-invaded by *A. altissima*; (2)
139 quantifying and comparing the ability of vegetation structure and environmental variables to predict
140 carabid beetle species composition in both non-invaded and invaded habitats. Additionally, we
141 supposed that cross-taxon congruence between carabid beetles and plants differ among non-invaded
142 and invaded sites; particularly, in invaded sites we expected more homogenous communities all
143 along the successional gradient of vegetation. More, we hypothesized that *A. altissima* favors more
144 synanthropic and eurieious carabid beetle and plant species.

145

146 **2. Materials & methods**

147 *2.1. Study area*

148 Fieldwork was carried out in the northeastern part of Italy, in North Adriatic Karst (NAK). The
149 study area (total extension of 4.9 km²) ranges between 58-114 m a.s.l. and lies in the municipalities
150 of Ronchi dei Legionari and Doberdò del Lago (Fig. 1). The NAK is a well-known area for its
151 geographical and geomorphological characteristics, located between the Adriatic Sea and the Alps.
152 The study area encompasses the westernmost part of the Italian Karst, that consists of the limestone
153 plateau (100-500 m a.s.l.), characterized by the typical geomorphological karst phenomena (karst
154 poljes, dolines, caves, etc.), and Red Mediterranean soils (Kaligarič et al., 2006; Poldini, 2009).

155 The Karst is known and traditionally recognized as a treeless, stony grassland landscape (with
156 exceptions of many flysch-bedrock patches), where the strong and cold Bora wind affects
157 vegetation causing desiccation and soil erosion. The climate is sub-Mediterranean, transitional
158 between Mediterranean and continental pre-Alpine types, with rainy cool winters and long and
159 relatively dry summers (Kaligarič et al., 2006; Poldini, 2009).

160 Currently, Karst landscape is generally dominated by mixed deciduous termophilous woodlands of
161 *Quercus pubescens*, *Ostrya carpinifolia* and *Fraxinus ornus*.

162 In the study area, we have identified one successional gradient of vegetation typical of the
163 calcareous soils of the Karst plateau: in phytosociological term, the vegetation series belong to the

164 series leading to the thermophilous mixed oak woods dominated by *Quercus pubescens*
165 (*Aristolochio luteae-Quercetum pubescentis* (Horvat 1959) Poldini 2008)). Along this dynamic
166 series of vegetation, we have selected non-invaded and invaded by *A. altissima* stages.

167 For methodological simplicity, each successional stage of the Karst vegetation series was defined as
168 an habitat due to homogeneous vegetation characteristics. In total 4 different habitats were
169 identified and sampled, of which four were not invaded (coded as 1, 2, 3 and 4) and three were
170 invaded by *A. altissima* (coded as 5, 6 and 7). Within each habitat, three randomly selected plots (of
171 almost 500 m² each) were sampled where both carabids and vegetation were investigated following
172 the scheme described below. It follows a brief description of the habitats along with the definition
173 of sampled plot codes.

174 Non-invaded communities (4 habitats):

175 (1) *Karst grassland*: characterized by *Centaureo cristatae-Chrysopogonetum grylli* Ferlan and
176 Giacomini 1955. Thermoxerophilous dry grassland (*A. altissima* absent). Here three plots were
177 sampled and coded as KG1, KG2, KG3.

178 (2) *Karst bushes (shrubland)*: this habitat is characterized by thermoxerophilous shrubs (*A.*
179 *altissima* absent) belonging to *Frangulo rupestris-Cotinetum coggygriae* Poldini and Vidali 2002
180 association. Three plots coded as KB1, KB2, KB3 were sampled.

181 (3) *Karst Wood in doline (dolines)*: characterized by mixed deciduous *Quercus pubescens* woodland
182 in shallow dolines (native species abundance > 90%, *A. altissima* < 1%). Three plots coded as
183 KW1, KW2, KW3 were sampled.

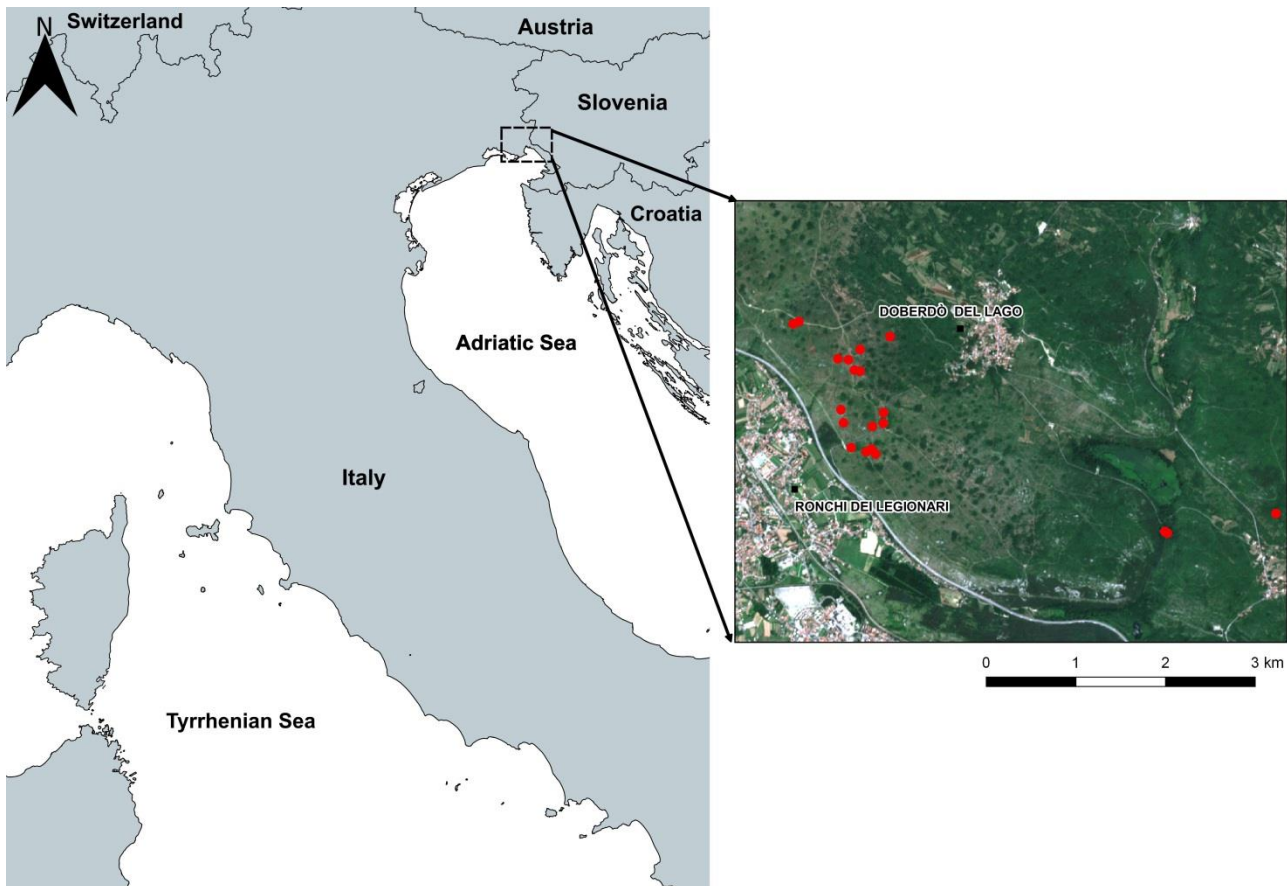
184 (4) *Karst Quercus woodland*: this habitat is characterized by *Aristolochio luteae-Quercetum*
185 *pubescentis* (Horvat 1959) Poldini 2008. Mature woodland of *Q. pubescens* (*A. altissima* absent).
186 Three plots coded as KQW1, KQW2, KQW3 were sampled.

187 Invaded communities dominated by *A. altissima* (3 habitats):

188 (5) *A. altissima* on karst grasslands: this habitat represents the first step of *A. altissima*'s
189 colonization on dry calcareous grassland (*A. altissima* abundance > 60%, native species abundance
190 < 20%). At the beginning of the study three plots were individuated, but one was destroyed by fire.
191 Therefore, two plots coded as AG1, AG2 were sampled.

192 (6) *Bushland with A. altissima*: characterized by the phases of invasion of *A. altissima* in karst
193 shrublands and pre-woods (*A. altissima* abundance > 55%, native species abundance < 35%). Three
194 plots coded as AB1, AB2, AB3 were sampled.

195 (7) *A. altissima* Wood: represented by dense and monospecific *A. altissima* woods in shallow
196 dolines (*A. altissima* abundance > 90%, native species abundance < 5%). Three plots coded as
197 AW1, AW2, AW3 were sampled.



199

200 Fig. 1. Study area with respect to Italian peninsula (left panel); the right panel shows a particular of
 201 the study area highlighting the sampled plots and the two municipalities near the study areas
 202 (Ronchi dei Legionari and Doberdò del Lago; Friuli Venezia Giulia region) (aerial photo from
 203 IRDAT FVG, <http://irdat.regione.fvg>).

204

205 2.2. Carabid beetles sampling

206 In each plot, three pitfall traps were placed 10 m apart in a straight line at the center of the area (N-
 207 W direction). The pitfall method is considered an inexpensive and labour efficient way to collect
 208 carabids for statistical analysis (Capar, 2003). Following Brandmayr et al. (2005), Zhang et al.
 209 (2017) traps were made by plastic vessels with an upper diameter of 9 cm and a depth of 11 cm
 210 provided with two small holes to avoid water filling and “aquaplaning effect” of beetles. Traps were
 211 filled up to two thirds of their depth with wine-vinegar saturated by sodium chloride as further
 212 preservation measure. Vinegar has been demonstrated to be a good collecting fluid and compared to
 213 pure water it allows a collection of larger numbers of species and individuals by avoiding
 214 degradation of anatomical structures (Mazzei et al., 2015). Carabid beetles were collected from 10th
 215 May 2014 until 12th January 2017 with a monthly control and emptying of pitfall-traps all year

216 round. The activity density of each species was calculated as DAa (annual activity density)
217 expressed as individuals/trap in the standard period of 10 days, the captures of all species as ADat
218 (total annual activity density) as in Brandmayr et al. (2005).

219 2.3. *Vegetation data collection*

220 In the central part of the 20 plots, an area of 10 m x 10 m was sampled. Within it, vascular plant
221 occurrences and abundances were recorded. Abundance was evaluated as percentage cover,
222 expressed using a seven-point scale according to Braun-Blanquet (1964). Sampling was carried out
223 in May-June 2014 and August-September 2014 in order to get a complete composition of plant
224 communities.

225

226 2.4. *Vegetation structure analysis*

227 In each of the 20 plots, we randomly defined a quadrat of 100 m² where we measured the following
228 metrics only for woody species higher than 1 m: *mean height* (H_mean), *mean diameter* (D_mean),
229 *mean total number of trees* (N), *density* and *total percentage cover* (Cop) of woody indigenous tree
230 species (I) and for *A. altissima* (A). The diameter was calculated at breast height (DBH) using a
231 metal tree caliper and the resulting measure was the arithmetic mean of the two readings, according
232 with Clark et al. (2000). The estimate of the tree height was performed using a manual hypsometer,
233 a precise and fast instrument (Rennie, 1979) using geometrical and trigonometric principles to
234 determine tree height.

235 2.5. *Soil parameters*

236 A soil pit 10 cm x 10 cm x 10 cm was dug near the center of the 20 plots. To characterize soil
237 properties, we considered the following variables: *soil rock fragments*, *soil fine earth fraction*, *soil*
238 *fine earth fraction*, *soil humidity*, *soil bulk density*, *soil porosity*, *soil mean weight diameter* (Table
239 A.1 in Appendix A in Supplementary material). One sample was taken from the upper part of the
240 mineral soil (0-7 cm in KG, AG, KB, 0-10 cm in KQW, 0-8 cm in AW, AB and KW) on each plot
241 in October 2017. The soil bulk density is the mass to volume ratio of an oven-dried soil. It was
242 determined in the 0-10 cm surface soil layer of all investigated sites, following the methods
243 described by Grossman and Reinsch (2002). In non-gravelly soil surface strata, it was determined
244 with the core method by using a steel cylinder of 10 cm height and 0.724 l volume. The cylinder
245 was inserted in the soil with the help of a steel hammer with nylon heads and dug out with a shovel.
246 The soil sample was finally extracted from the cylinder and put in a plastic bag. When the content

247 of rock fragments was high, an alternative excavation method was adopted. A plastic film with a
248 central hole of 15 cm diameter was placed on a roughly plain soil surface. A soil sample was dug
249 out of the hole with a knife, put on the plastic film and transferred in a plastic bag. Afterwards, the
250 excavated hole was covered with a second plastic film and filled with water to the surface with the
251 help of 250 ml graduated cylinder. The volume of the hole was given by volume of water added to
252 the excavated hole. In both cases, samples were transported in the laboratory, oven-dried at 105°C
253 and weighted. The bulk density was calculated as the mass to volume ratio of oven-dried cores and
254 expressed as kg L⁻¹. Total soil porosity was obtained by the equations:

$$255 \qquad \qquad \qquad \text{Total porosity} = 1 - \rho_a / \rho_s$$

256 where ρ_a is the bulk density, and ρ_s the particle density of the soil, which usually averages 2.65 Kg
257 l⁻¹. Soil porosity was obtained from soil bulk density; soil mean weight diameter refers to the mean
258 dimension of soil aggregates; soil fine earth fraction is the solid fraction less than 2 mm.

259

260 *2.6. Environmental variables*

261 We derived environmental variables (Latitude, Longitude, Slope, Elevation, Northness, Eastness,
262 depth of dolines) from the Digital Elevation Models (DEMs, resolution 10 m) using ArcGIS 10.2.1
263 (ESRI, 2014). Karstification level was defined following Forti (1982).

264 Plot environmental characteristics (ordered by successional gradient of vegetation from wood to
265 grassland) are reported in Table A.2 in Appendix A in Supplementary material.

266 *2.7. Statistical analysis*

267 Cross taxon congruence between plants and carabid beetles and their relationships with the other
268 predictors were assessed using different statistical methods. Firstly, Mantel and partial Mantel test
269 were calculated using Spearman correlation coefficients and 999 permutations. This analysis aims
270 at assessing the significance of the relationship between pairwise distance of species composition
271 (both carabids and plants) and those of predictor variables (Legendre and Legendre, 2012; Frenett et
272 al., 2013; Westgate et al., 2014; Larios et al., 2017). Before performing Mantel and partial Mantel
273 test, distance matrices for each group of variables (environmental parameters, soil parameters,
274 vegetation structure; Fig. A.1 in Appendix A in Supplementary material) were calculated as
275 follows: 1) for environmental variables, soil variables and vegetation structure, a Euclidean distance
276 matrix was calculated; 2) for carabid beetles and plant communities the Bray-Curtis distance was
277 used (Legendre and Legendre, 1998). Carabid abundance data and plant community data were

278 square root transformed prior to analysis to meet assumptions about normality. Later, Co-
279 correspondence analysis (Co-CA, see ter Braak and Schaffers, 2004; Schaffers et al., 2008 for a full
280 description of this method), was used to quantify the strength of plant community data in predicting
281 the carabid beetle species composition. Here, we used the predictive version of Co-CA, which
282 combines the maximization of weighted covariance between weighted averages of species scores
283 and partial least squares methodology (PLS; Martens and Naes, 1992). A leave-one-out cross-
284 validatory fit percentage was estimated to select the minimal adequate predictive models. Schaffers
285 et al. (2008) pointed out that, due to its predictive nature, any cross-validatory fit > 0 implicitly
286 validates the model, indicating that prediction is better than that obtained under the null model. A
287 permutation test for predictive co-correspondence analysis models to assess the significance of each
288 Co-CA ordination axis was applied (999 permutations); in addition, abiotic and biotic vectors were
289 fitted onto ordination axes assessing the importance and significance of each vector by means of
290 ‘*envfit.coca*’ function.

291 In order to further assess the relationships between beetles and successional stages of vegetation, we
292 used indicator species analysis (Dufrêne and Legendre, 1997) coupled with combinations of site
293 groups according to De Cáceres and Legendre (2009). The same analysis was also applied to detect
294 differences between non-invaded vs. invaded stages of vegetation.

295 Lastly, the pure and shared effect of spatial factors, soil and environment parameters, vegetation
296 structure on plants and on carabid communities was evaluated using a variation partitioning
297 approach that allows the partitioning of the total variation (calculated as partial redundancy
298 analysis) to be broken down into the contributions of each variable group (Borcard et al., 1992;
299 Peres-Neto and Legendre, 2010).

300 Mantel and partial Mantel tests along with variation partitioning were computed using R package
301 *vegan* (Oksanen et al., 2018); Co-CA and related analysis with R package *cocorresp* (Simpson,
302 2009), indicator species analysis with R package *indicspecies* (De Cáceres and Legendre, 2009). All
303 statistical analyses were performed using R 3.5.1 (R Core Team 2018).

304

305 **3. Results**

306 In total, 3119 individuals were collected belonging to 28 carabid species. Non-invaded stands
307 (KQW2) showed higher abundance of carabids whereas values of invaded stands the lowest ones
308 (AG2; see Table A.2 in Appendix A in Supplementary material). Overall, the most common species

309 was *Carabus coriaceus coriaceus* and the rarest were *Carabus granulatus interstitialis*, *Harpalus*
 310 *serripes*, *Ophonus azureus*, *Brachinus explodens*. Invaded wood was the habitat type where the
 311 highest number of species was detected (15 species in AW3, 11 species in AW1); in contrast, non-
 312 invaded karst bushes showed lower values of carabid species richness (2 species in KB3) (Table
 313 A.2 in Appendix A in Supplementary material). Out of the 18 species sampled in the non-invaded
 314 and invaded wood habitats, *Laemostenus cavicola* and *Carabus granulatus interstitialis* were found
 315 only in the invaded plots, *Abax parallelepipedus subpunctatus*, *Calosoma sycophanta*, *Notiophilus*
 316 *rufipes* were found exclusively in non-invaded woods. In total, 10 species were associated only with
 317 *A. altissima*: *Licinus hoffmanseggii*, *Abax carinatus sulcatus*, *Carabus catenulatus catenulatus*,
 318 *Harpalus atratus*, *Leistus rufomarginatus*, *Harpalus rubripes*, *Ophonus azureus*, *Calathus*
 319 *melanocephalus*, *Brachinus explodens*, *Pseudoophonus rufipes*.

320 Considering plant species, we detected 173 species in total (mean number of species per plot was
 321 28.55 ± 9.35). The most abundant tree species was *Fraxinus ornus* (85% of the plots), the most
 322 abundant shrub species was *Cotinus coggygria* (70% of the plots), and the most abundant
 323 herbaceous species were *Brachypodium rupestre* (55% of the plots), *Sesleria autumnalis* (40% of
 324 the plots), *Carex humilis* (40% of the plots). As expected, non-invaded grasslands showed the
 325 highest values of plant species richness whereas non-invaded shrubs the poorest ones. 35 species
 326 were associated only with *A. altissima*, the most abundant were: *Duchesnea indica* (10% of the
 327 plots), *Elymus repens* (10% of the plots), *Erigeron annuus* (10% of the plots), *Aristolochia*
 328 *clematitis* (5% of the plots), *Fragaria viridis* (5% of the plots), *Mercurialis perennis* (5% of the
 329 plots), *Allium vineale* (10% of the plots), *Lathyrus latifolius* (5% of the plots), *Populus nigra* (5% of
 330 the plots). 72 species were present only in non-invaded plots: *Sesleria autumnalis* (40% of the
 331 plots), *Quercus pubescens* (30% of the plots), *Cornus mas* (25% of the plots), *Chrysopogon gryllus*
 332 (15% of the plots), *Stipa eriocalis s.l.* (15% of the plots) *Quercus cerris* (10% of the plots).

333 The correlation between the plants and carabid beetles dissimilarity matrices and each group of the
 334 environment variables (e.g. soil variables, vegetation structures, habitat types distance matrices)
 335 were significant (Table 1). Even though most of the Mantel tests showed significant correlations,
 336 partial Mantel revealed that most of the cross-taxon congruence is controlled by abiotic and biotic
 337 predictors; interestingly, the correlation remained significant only after controlling for *A. altissima*
 338 presence and structure. A slight trend was detected, albeit not significant, when controlling for soil
 339 variables.

Test	Distance matrices	Spearman's ρ	p-value
	Plant ~ All Environment and Vegetation	0.51	<0.001

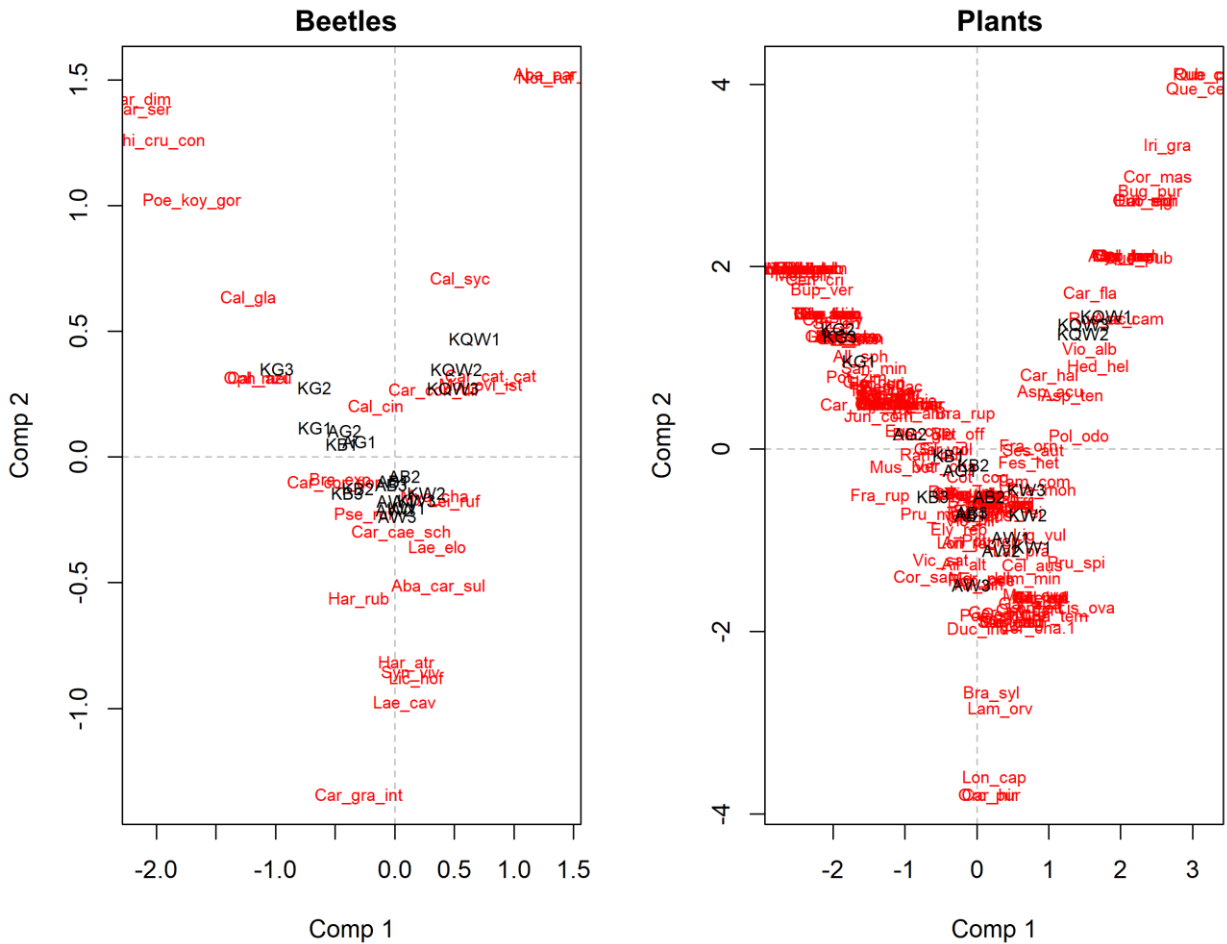
Mantel Test	Plants ~ Environmental Variables	0.24	0.008
	Plants ~ Soil Variables	0.22	0.0123
	Plants ~ Ailanthus presence and structure	0.22	0.027
	Plants ~ Native Plant Structure	0.42	<0.001
	Plants ~ Habitat Type	0.45	<0.001
	Beetles ~ All Environment and Vegetation	0.43	<0.001
	Beetles ~ Environmental Variables	0.40	<0.001
	Beetles ~ Soil Variables	0.43	<0.001
	Beetles ~ Ailanthus presence and structure	-0.09	0.768
	Beetles ~ Native Plant Structure	0.35	<0.001
	Beetles ~ Habitat Type	0.34	0.001
	Plants vs. Beetles	0.23	0.019
Partial Mantel Test	Plants vs. Beetles ~ All Environment and Vegetation	0.00	0.468
	Plants vs. Beetles ~ Environmental Variables	0.14	0.085
	Plants vs. Beetles ~ Soil Variables	0.14	0.074
	Plants vs. Beetles ~ Ailanthus presence and structure	0.25	0.010
	Plants vs. Beetles ~ Native Plant Structure	0.09	0.178
	Plants vs. Beetles ~ Habitat Type	0.08	0.193

340 **Table 1.** The Spearman correlation coefficients of Mantel test and Partial Mantel tests between all environmental
341 distance matrices and the dissimilarity matrix of carabid beetles and plants (~ means: controlling for). In bold the
342 significant values.

343

344 The ordination diagram of Co-CA analysis shows a gradient based on vegetation structure
345 (grassland, shrubland, woodland in dolines) that could be identified along the first axis (Fig. 2). The
346 biplot allowed the identification of thermophilous and xerotolerant carabid beetle species, typical of
347 open habitats such as *Harpalus serripes*, *Harpalus dimidiatus*, *Philorhizus crucifer confusus*,
348 *Poecilus koyi goricianus*, grouped on the bottom left of the diagram, and associated with the
349 characteristic grassland plant species as *Bupleurum veronense* and *Chrysopogon gryllus*. Non-
350 invaded mature stands dominated by *Quercus pubescens* (KQW) were all grouped together on the
351 bottom right, and were associated to *Abax parallelepipedus subpunctatus*, *Notiophilus rufipes*, and
352 with higher abundances of *Calosoma sycophanta*, *Molops ovipennis istrianus*, *Carabus catenulatus*
353 *catenulatus*. These species were consistently associated with plant species typical of Karst mature
354 forests (such as *Quercus pubescens*, *Quercus cerris*, *Asparagus tenuifolius*). In the upper part of the
355 biplot, species connected with non-invaded and invaded stands in dolines occur: humid forest
356 species, such as *Licinus hoffmanseggii* and *Abax carinatus sulcatus*, troglophilous and crevice
357 dweller species *Laemostenus cavicola* and *Laemostenus elongatus*, species usually linked with
358 swampy forests as *Carabus granulatus interstitialis*, and woodland thermophilous species as
359 *Synuchus vivalis* and *Harpalus atratus*. In association with these carabid species there were

360 *Lonicera caprifolium*, *Lamium orvala* and *Brachypodium sylvaticum*, which are particularly
 361 abundant in doline wooded habitats.



362
 363 Fig. 2. Predictive Co-CA biplot of carabid beetle species composition (beetles, left) and plant species
 364 composition (plants, right) (A). In each plot, species are positioned according to their loadings with respect
 365 to normalized site scores derived from the plant composition data. The axes were rescaled to the same ranges
 366 so that sites occupy the same position in both plots.

367 It is very important to underline that the variables that produce an effect on congruence across taxa
 368 were the structure of non-invaded vegetation and habitat type, while invaded vegetation did not
 369 show an influence on carabid beetles and plants.

370 The prediction accuracy of plant species composition on beetle composition was above zero (cross-
 371 validatory fit percentages, Table 2), indicating that the predictions of carabid beetle species
 372 composition based on that variables were better than those expected under the null model (no
 373 relationship). Plotting the cross-validatory fit percentage for the compared predictive datasets
 374 against the number of axes showed the maximum prediction level obtained at three axes. In the

375 model we therefore retained only the first axis. In Fig. A.2 in Appendix A in Supplementary
 376 material are displayed the results of the Predictive Co-Ca. These showed a clear clustering of
 377 habitat types in some groups: firstly, woods growing in dolines (both non-invaded and invaded) are
 378 characterized by higher karstification values, higher soil porosity and soil fine earth fraction;
 379 whereas *Karst Quercus woodland* are correlated with higher values of soil rock fragments and soil
 380 bulk density. All plots with the highest density of *A. altissima* are clustered together. Most of the
 381 environmental variables were significantly correlated with Co-Ca axes (Table 3). It is important to
 382 underline that we found a strong correlation of carabid beetles with soil porosity ($P < 0.01$),
 383 karstification ($P < 0.01$), non-invaded vegetation structure canopy cover ($P < 0.01$). Plants were
 384 strongly correlated with soil porosity ($P < 0.01$) and soil rock fragments ($P < 0.01$), karstification (P
 385 < 0.001), native canopy cover ($P < 0.001$), native plants diameter ($P < 0.01$) and native plants
 386 height ($P < 0.01$), diameter ($P < 0.01$) and height ($P < 0.01$).

387

	Predictive Power (plants on beetles)	Cumulated Predictive Power	Cross-validatory fit (%)	p-value
Axis 1	18.642	18.642	9.739	0.001
Axis 2	10.844	29.486	16.796	0.472
Axis 3	10.222	39.709	17.862	0.625
Axis 4	6.890	46.599	14.745	0.837

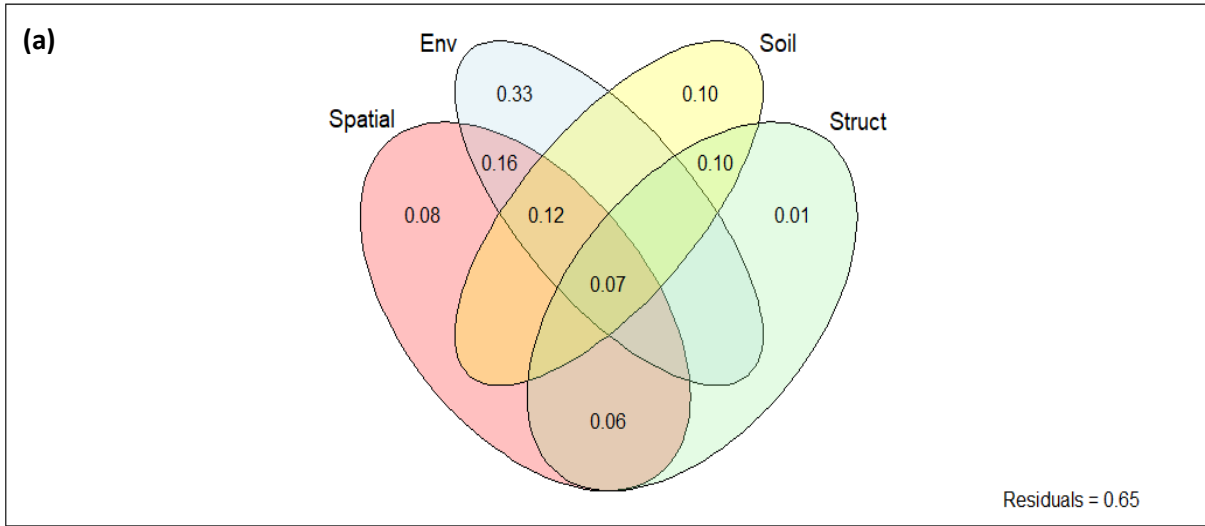
388 **Table 2.** Predictive power of plants on carabid beetles composition

389

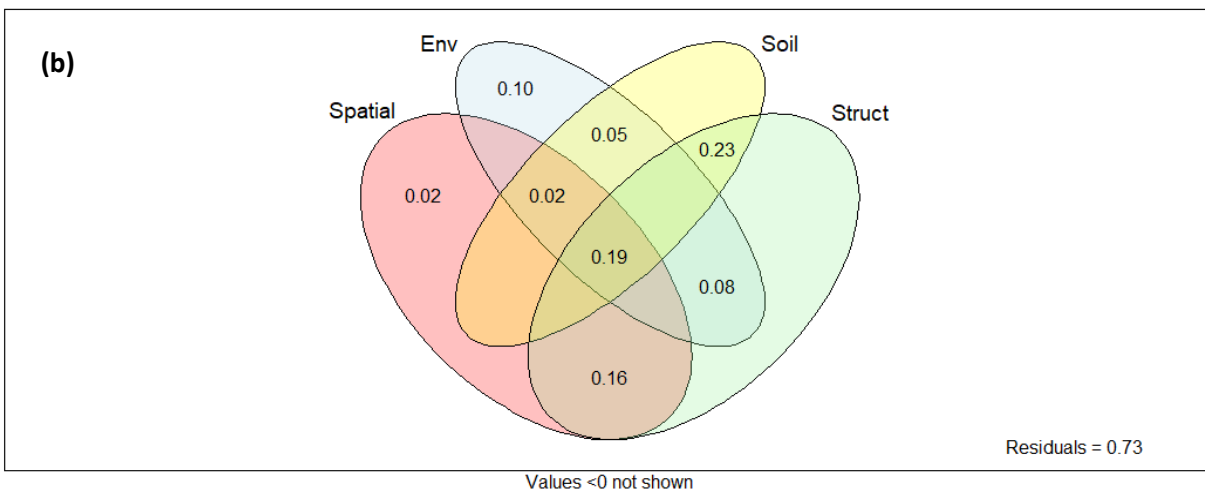
390 Considering indicator species analysis, we found a low number of carabid beetle indicator species
 391 connected with certain successional vegetation stages. Furthermore, even though in non-invaded
 392 stages no species were detected as indicators, one species (*Calathus cinctus*) resulted as closely
 393 associated with invaded communities (Tab. A.3 in Appendix A).

394 The contribution of space, environment, soil parameters and vegetation structure differed between
 395 carabid beetle and plant community composition (Fig. 3). The relative contribution of spatial factors
 396 was similar between these two groups, and environmental relative contribute was the most
 397 important factor in both groups, even though beetles resulted to be more influenced (Fig. 3a,b). The
 398 relative contribution of the vegetation structure and soil factors was not coincident among the two
 399 groups. The covariation of vegetation structure with soil parameters accounted for much of the
 400 variation in plant communities. Spatial and vegetation structure factors together determined the
 401 abundance and distribution of plant pattern almost like spatial, vegetation, environment and soil
 402 together. On one hand, soil parameters and vegetation structure together and on the other hand
 403 spatial, environment and soil structures together were the most important factors in defining

404 patterns of carabid beetles. Considering also *A. altissima* vegetation structure as factor (Fig. A.1 a,b
 405 in Appendix A in Supplementary material), its presence explained carabid beetle distribution, but
 406 less than spatial and environment relative factors. Non-invaded vegetation considered as factor did
 407 not influence carabids species pattern, since most of the variations was explained by shared
 408 variation between environmental and spatial factors.



409



410

411 Fig. 3. Result of the variance partitioning analysis, showing the contribution of Spatial factors, Environmental variables
 412 (Env), Soil and Vegetation Structure factors (Struct) to the variation in carabid beetles (a) and plants (b).

413

414

	Beetle		Plant	
	R ²	p-value	R ²	p-value
Soil rock fragments	0.39	0.019	0.45	0.010

Soil fine earth fraction	0.28	0.065	0.35	0.037
Soil humidity	0.27	0.077	0.29	0.066
Soil bulk density	0.35	0.033	0.36	0.030
Soil porosity	0.47	0.005	0.48	0.006
Soil mean weight diameter	0.33	0.037	0.35	0.036
Environmental Variables	R²	p-value	R²	p-value
Latitude	0.63	0.000	0.52	0.003
Longitude	0.84	0.001	0.86	0.000
Slope	0.52	0.005	0.61	0.002
Elevation	0.44	0.008	0.35	0.030
Northness	0.14	0.303	0.14	0.308
Eastness	0.01	0.904	0.03	0.790
Doline depth	0.40	0.022	0.30	0.065
Karstification	0.58	0.002	0.64	0.001
Vegetation Structure	R²	p-value	R²	p-value
AH_mean	0.40	0.019	0.54	0.005
AD_mean	0.37	0.026	0.49	0.008
A_density	0.32	0.047	0.42	0.017
N_A	0.32	0.047	0.42	0.017
Cop_A	0.25	0.096	0.40	0.020
IH_mean	0.54	0.002	0.50	0.003
ID_mean	0.49	0.005	0.47	0.006
I_density	0.42	0.017	0.34	0.043
N_I	0.42	0.017	0.34	0.043
Cop_I	0.49	0.005	0.61	0.001
	R²	p-value	R²	p-value
Environmental variables with soil	0.58	0.001	0.43	0.008

415 **Table 3.** Environmental vectors estimated on Co-Ca ordination using *envfit.coca* function (999 permutations) and
416 related squared correlation coefficient. In bold the significant values, for abbreviations see Section 2.4
417

418 Variance partitioning showed that 33% of the total explained variation for the ordination of carabid
419 beetles assemblages was due to the independent effect of environment, whereas the pure effects of
420 spatial variables, vegetation structure and soil parameters explained 8%, 10% and 1% of this total
421 variation, respectively. If we analyze independently the effect of invaded vegetation structure, it

422 explained the 18% of the total variance, whereas the non-invaded vegetation structure was not
423 informative.

424

425 **4. Discussion**

426 In this study, we assessed cross-taxon congruence between carabid beetles and vascular plants
427 among invaded vs. non-invaded stages along a successional gradient of vegetation (from grassland
428 to woodland), estimating also the explicative power of biotic and abiotic factors on both
429 communities. Since both plants and carabid beetles respond similarly to environmental gradient,
430 plants can effectively be used as a surrogate taxon in the study area. Our findings are in agreement
431 with Brunbjerg et al. (2018), who proved that plant species, considered along environmental
432 gradients, are a useful surrogate for biodiversity in general.

433 In this context, we also proved that there is a differentiation in species communities along non-
434 invaded vegetation stages, while this differentiation is not significant along the invaded vegetation
435 stages. This suggests that both carabid beetle and plant communities are strongly influenced by *A.*
436 *altissima*, which may promote a homogenization of the communities, fostering a subset of species
437 typical of disturbed and anthropized area.

438 Carabid assemblages are known to be affected by microhabitat variation and biotic interactions at
439 smaller scales (e.g. “patchiness”) (Niemelä et al., 1992; Antvogel and Bonn, 2001; Thomas et al.,
440 2001; Brose, 2003; Barbaro et al., 2007; Schreiner and Irmeler, 2009) and their communities have
441 been directly related to different forest stages (Riley et Browne, 2011), ground vegetation and litter
442 (Niemelä et al. 1993; Niemelä and Spence, 1994; Koivula, 2001; Rainio and Niemelä, 2003).

443 Even if many studies related carabid fauna to biotic and abiotic conditions (Rainio and Niemelä,
444 2003; Koivula, 2011) and to environmental variables such as temperature, rainfall, vegetation cover,
445 soil moisture (Ings and Hartley, 1999; Jukes et al., 2001; Perner and Malt 2003; Kotze et al., 2011;
446 Moraes et al., 2013; Lee and Albajes, 2016), it is still not completely clear which of these variables
447 best correlate to their observed patterns of richness and abundance (Niemelä, 1996) and which are
448 more informative at a certain scale and/or condition (e.g. preserved or altered environments, Moraes
449 et al., 2013). In agreement with Ings and Hartley (1999), who reported that vegetation and soil
450 factors influenced the prey of the carabids rather than Carabidae themselves, we found a strong
451 effect of soil features on carabid beetle communities. The most important soil factors for both
452 carabid beetles and plant species in our study area were the soil porosity and soil rock fragments,

453 characterizing the typical Karst red soil. Karstic woodlands, especially in doline (both non-invaded
454 and invaded), appeared to be linked to greater soil porosity whereas soil rock fragments characterize
455 grasslands. Carabid beetles living in different types of grassland are directly associated with soil
456 factors such as soil organic matter content and soil moisture (Luff et al., 1989; Gardner, 1991;
457 Gardner et al., 1997) and there are many studies showing how carabid fauna is useful to
458 discriminate with fine resolution habitats such as grassland (e.g. Tanabe et al., 2007). In our study,
459 we found species related to grassland, those typical of open habitats, that were *Harpalus dimidiatus*,
460 *Philorhizus crucifer confusus*, *Poecilus koyi goricianus*. The indicator species analysis confirmed
461 *Poecilus koyi goricianus* as indicator species of grassland dominated by *A. altissima* (see Tab. A.3
462 in Appendix A). On the other hand, we found also many groups of carabid beetles belonging to
463 woodland habitats such as *Synuchus vivalis*, *Harpalus atratus*, *Leistus rufomarginatus* which are
464 thermotolerant and heliophilous species. Others are known to be typical species of more humid and
465 fresh woods (*Licinus hoffmanseggii*, *Abax carinatus sulcatus*) or to be typical species of
466 mesophilous karst woods (*Carabus catenulatus catenulatus*, *Molops ovipennis istrianus*)
467 (Brandmayr et al., 1983, 2005).

468 In general, even if we found many species which are connected exclusively to grassland habitats,
469 such as *Harpalus dimidiatus*, *Harpalus serripes*, *Ophonus azureus*, it is interesting to note that
470 when *A. altissima* invaded grasslands, it promoted *Calathus cinctus*, a typical species of secondary
471 ecosystems and degraded grasslands. The indicator species analysis confirms this pattern, with *C.*
472 *cinctus* and *Synuchus vivalis* that are the only species strictly connected with invaded habitats. This
473 result is particularly significant considering that *A. altissima* seems to be connected with an increase
474 of distribution abundance also of another species, *Harpalus atratus*, a species connected to
475 anthropic and disturbed areas (Brandmayr et al., 2005).

476 Furthermore, invaded habitats showed the highest carabid beetle species number. Several studies
477 underlined that heterogeneous environments have usually higher diversity and abundance of carabid
478 beetle species (Butterfield, 1997; Magura et al., 2003; Hartley et al., 2007; Fujita et al. 2008).
479 Actually, when carabid fauna is compared between undisturbed and disturbed environments, the
480 species number is often observed to increase in the latter ones (Brandmayr et al., 2005; Latty et al.,
481 2006; Uehara-Prado et al., 2009). This can be a consequence of generalist species emigrating from
482 open areas, following the appropriate conditions created by disturbance (Latty et al., 2006). Another
483 possible interpretation of the higher species numbers observed could be seen in the higher soil
484 humidity of the *Ailanthus* wood that favors the concentration of wet soil dwellers, like *Carabus*
485 *granulatus*. There is a clear evidence from the literature that carabid assemblage composition

486 changes under structural changes of the habitat (Scott and Anderson, 2003), and that habitat type
487 represents the most important factor explaining their distribution patterns at the regional and
488 landscape scales (Dufrêne, 1992; Penev, 1996; Aviron et al., 2005; Eyre et al., 2005; Hartley et al.,
489 2007).

490 Concerning the impact of invasive species on ecosystems, it is interesting to cite the effects of
491 *Robinia pseudoacacia* on 18 arthropod taxa in woodlands (Buchholz et al., 2015); it emerges that
492 the invasion of this tree can induce species turnover in ground-dwelling arthropods, but it is not
493 connected necessarily with reducing in arthropod species abundances or diversity.

494 Carabid beetles living in different types of grassland are directly associated with soil factors such as
495 soil organic matter content and soil moisture (Luff et al., 1989; Gardner, 1991; Gardner et al., 1997)
496 and there are many studies showing how carabid fauna is useful to discriminate with fine resolution
497 habitats such as grassland (e.g. Tanabe et al., 2007).

498 The highly complex of environmental, soil and vegetation structure variables which explains the
499 cross-taxon congruence of carabid beetle and plant communities, confirms the various biotic and
500 abiotic factors (Judas et al., 2002; Samin et al., 2011) affecting carabid beetles in their distribution,
501 both within and between habitats (Thiele, 1977, 1979; Lövei and Sunderland, 1996). Our study area
502 is intentionally localized, so that we did not expect to observe a great influence of environmental
503 and, in particular, climatic parameters (Duan et al., 2016), that usually become more important in
504 large study areas due to more pronounced gradients (Qian and Kissling, 2010; Toranza and Arim,
505 2010). An in-depth analysis among the environmental variables showed that plant and carabid
506 beetle assemblages responded to almost the same variables, such as slope, elevation, karstification.
507 Elevation is commonly considered as a whole array of environmental factors (Duan et al., 2016),
508 and its crucial role as explanatory environmental factor for the species. The resulting congruence in
509 cross-taxon is furthermore an evidence that elevation is strongly correlated not only with
510 intermediate and large study areas, but also with the small ones. Karstification is another important
511 factor (more in plant than in carabid beetle distribution) that must be taken into account: a strong
512 relationship degree was already known for the two sphodrine species, *Laemostenus elongatus* and
513 *Laemostenus cavicola*, that in our study area were present in relation only to doline forest plots
514 (Brandmayr et al., 1980).

515 The combination of soil and vegetation structure together expressed the 10% of the total variation,
516 and this is in accordance with the main known factors driving the distribution of carabid beetles,
517 that are always related to micro- and mesoscale landscape (Barbaro et al., 2007), and that take into
518 account microclimate, vegetation structure, prey density, predation, competition or localized

519 oviposition sites (Niemelä et al., 1992; Antvogel and Bonn, 2001; Thomaset al., 2001; Magura,
520 2002; Brose, 2003). However, all the environmental variables combined with soil parameters were
521 really significant for carabid beetle's distribution. The effect of vegetation structure, in particular
522 those of the canopy of the native species, was already reported to be an important factor
523 conditioning carabid beetle's assemblages (Gardner, 1991; Sanderson et al., 1995; Brandmayr et al.,
524 2005; Tanabe et al., 2007) thanks to the greater shading and water uptake caused by larger trees,
525 which in turn affects incoming solar radiation, light availability, soil moisture, humidity and the
526 composition of the flora (Ings and Hartley, 1999). Shady humid conditions are associated with
527 many species of carabid beetles especially in the early stages of their development (Thiele, 1977),
528 whereas other species are associated with more dry and open conditions. A good example of these
529 indirect effects can be observed in the distributions of *Leistus rufomarginatus*, *Carabus catenulatus*
530 *catenulatus*, *Carabus caelatus schreiberi*, *Molops ovipennis istrianus*, *Abax parallelepipedus*
531 *subpunctatus*, *Calosoma sycophanta*: all these species were more present in the habitats of more
532 developed, older woods, suggesting a preference for mature woodland (Lindroth, 1985). Ings and
533 Hartley (1999) confirmed these observations and suggested that species are responding to the shady
534 and humid microclimate found under large and old trees, rather than to the presence of mature trees
535 *per se*. Furthermore, in contrast with Tanabe et al. (2007), it is worth nothing that the different wood
536 habitats were grouped separately in the Co-correspondence analysis (Fig. 2).

537 We found different species communities: thermotolerant and heliophilous species (such as *Synuchus*
538 *vivalis*, *Harpalus atratus*, *Leistus rufomarginatus*), species typical of more humid and fresh wood
539 (*Licinus hoffmanseggii*, *Abax carinatus sulcatus*), species typical of mesophilous karst wood
540 (*Carabus catenulatus catenulatus*, *Molops ovipennis istrianus*) (Brandmayr et al., 2005). Their
541 habitat affinity in the Triestine Karst has been carefully described analyzing the main bioclimate
542 gradient from deciduous woodlands to Mediterranean wood types in Brandmayr et al. (1983).

543

544 Summing up, the most important variables showing a determinant effect on congruence across taxa
545 were the structure of non-invaded vegetation and habitat type, considering the successional gradient
546 of vegetation from grassland to wood. On the contrary, if we consider sites dominated by *A.*
547 *altissima*, in correspondence of habitat type change, the structure of invaded vegetation and carabid
548 assemblages remain almost the same. This fact confirms the known concept that carabid beetles
549 show a positive relationship with environmental parameters such as vegetation cover (Thiele, 1977;
550 Gardner, 1991; Gardner et al., 1997; Sanderson et al., 1995; Ings and Hartley, 1999; Eyre et al.,
551 2001; Brandmayr et al., 2005; Desender et al., 2013; Alignier and Aviron, 2017), despite the
552 strength of this connection is stronger in non-invaded habitats compared to the ones invaded by *A.*
553 *altissima*.

554

555 **5. Conclusions**

556 Our results suggest that spatial distribution patterns of carabid beetles in the study area were mainly
557 determined by environmental parameters rather than vegetation or spatial structure. Interestingly,
558 we observed that more diversified communities of carabid beetles and plants occurred in non-
559 invaded habitats, while trivialized communities characterized invaded areas. Additionally, the
560 typical carabid species connected with *A. altissima* stands were those mainly common in
561 anthropized and degraded environments. Finally, plants and carabid beetles respond to
562 environmental variables in the same way and, in particular, plants respond to the same underlying
563 main gradient, being at least sensitive to different conditions as carabid beetles (Gioria et al., 2009).
564 In conclusion, plants best represent the diversity of carabid beetles in NAK region and can thus be
565 defined as a surrogate group for carabid beetles, as already found by Lários et al. (2017) for other
566 animal groups in Neotropical regions.

567

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569

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574

575 **Appendix A. Supplementary data**

576

577

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