

## SUPPLEMENTARY MATERIAL

Table S1. A) Summary of culling effort. Noting that the hours of culling efforts refers only to the dive time associated directly to sea urchin removal, thus excluding the time spent to set up systematic culling transects. B) ANOVAs testing the effect of culling on sea urchin density ( $n^\circ$  individuals  $m^{-2}$ ) across time.

A)

<b>Cull location</b>	<b>Days</b>	<b>Number of divers</b>	<b>Number of urchins removed</b>	<b>Hours of culling effort</b>	<b>Cull rate (urchins/diver/min)</b>
1	1	5	8756	7.5	19.46
	2	8	13122	12	18.23
	3	7	12568	10.5	19.95
	4	8	13898	12	19.30
2	5	6	9112	9	16.87
	6	6	9823	9	18.19
	7	8	13114	12	18.21
	8	8	12125	12	16.84
Totals		56	92518	84	Mean = 18.38

B)

<b>Source of variation</b>	<b>df</b>	<b>MS</b>	<b>F</b>	<b>P</b>
Time = Ti	4	77.41	15.51	0.0008
Treatment = Tr	1	391.68	1120.69	0.0009
Si = Si(Tr)	2	0.35	0.12	0.8844
Ti × Tr	4	12.86	2.58	0.1185
Ti × Si(Tr)	8	4.99	1.76	0.1144
Residuals	40	2.84		
Total	59			

Table S2. List of taxa found in the study. The aggregations adopted for univariate analyses were also reported. Abbreviations used in canonical analysis of principal coordinates (CAP) are also reported. \* Taxa not considered in univariate analyses.

<b>TURF FORMING ALGAE</b>	<b>Abbr.</b>	<b>INVERTEBRATES</b>	<b>Abbr.</b>
<i>Acetabularia acetabulum</i>		<i>Actinia</i> sp.	Act
<i>Amphiroa rigida</i>	Amp	<i>Aiptasia mutabilis</i>	Aip
<i>Anadyomene stellata</i>	Ana	<i>Aplysina aerophoba</i>	
Articulated Corallines	AC	<i>Ascidia mentula</i>	Asc
<i>Caulerpa cylindracea</i>	Cau	<i>Balanophyllia europea</i>	Beur
Dark Filamentous Algae	DFA	<i>Botryllus</i> sp.	
<i>Dasycladus vermicularis</i>	Dver	Calcareus Tube Worms	CTW
Dumontiaceae		<i>Caryophyllia smithii</i>	Cary
Green Filamentous Algae	GFA	<i>Cereus pedunculatus</i>	
<i>Liagora viscida</i>	Lvis	<i>Chondrilla nucula</i>	
Stypocaulaceae		<i>Chondrosia reniformis</i>	
<i>Tricleocarpa fragilis</i>	Tfra	<i>Cladocora caespitosa</i>	
<i>Valonia macrophysa</i>		<i>Cliona</i> spp.	
		Didemnidae	Did
<b>ERECT MACROALGAE</b>		Encrusting Bryozoans	EB
<i>Codium bursa</i>	Cod	Encrusting Red Sponges	ERS
<i>Cystoseira compressa</i>	Cys	<i>Halocynthia papillosa</i>	
Dictyotales	Dic	<i>Hemimycale columella</i>	Hem
<i>Flabellia petiolata</i>	Fla	Hydroids	Hyd
<i>Halimeda tuna</i>		<i>Ircinia variabilis</i>	
<i>Laurencia</i> complex	Lau	<i>Lithophaga lithophaga</i>	
<i>Padina pavonica</i>	Pad	Massive Dark Sponges	MDS
<i>Sphaerococcus coronopifolius</i>		<i>Petrosia ficiformis</i>	
<i>Wrangelia penicillata</i>	Wra	<i>Phorbas tenacior</i>	
		<i>Rocellaria dubia</i>	
<b>ENCRUSTING ALGAE*</b>		<i>Terpios fugax</i>	
Encrusting Calcified Rhodophyceae	ECR	Thin Ramified Bryozoans	
<i>Palmophyllum crassum</i>		Vermetidae	Ver
<i>Peyssonnelia</i> spp.	Pey		



Figure S1. Canonical analysis of principal coordinates (CAP) for the factor  $T_i \times T_r$  based on the distance matrix (Jaccard dissimilarity index, i.e., presence absence data) of areas of assemblages in the different sampling times (T0:T4). Numbers (from 0 to 4) indicate the sampling times. G+ = reference conditions; G- = manipulated conditions.

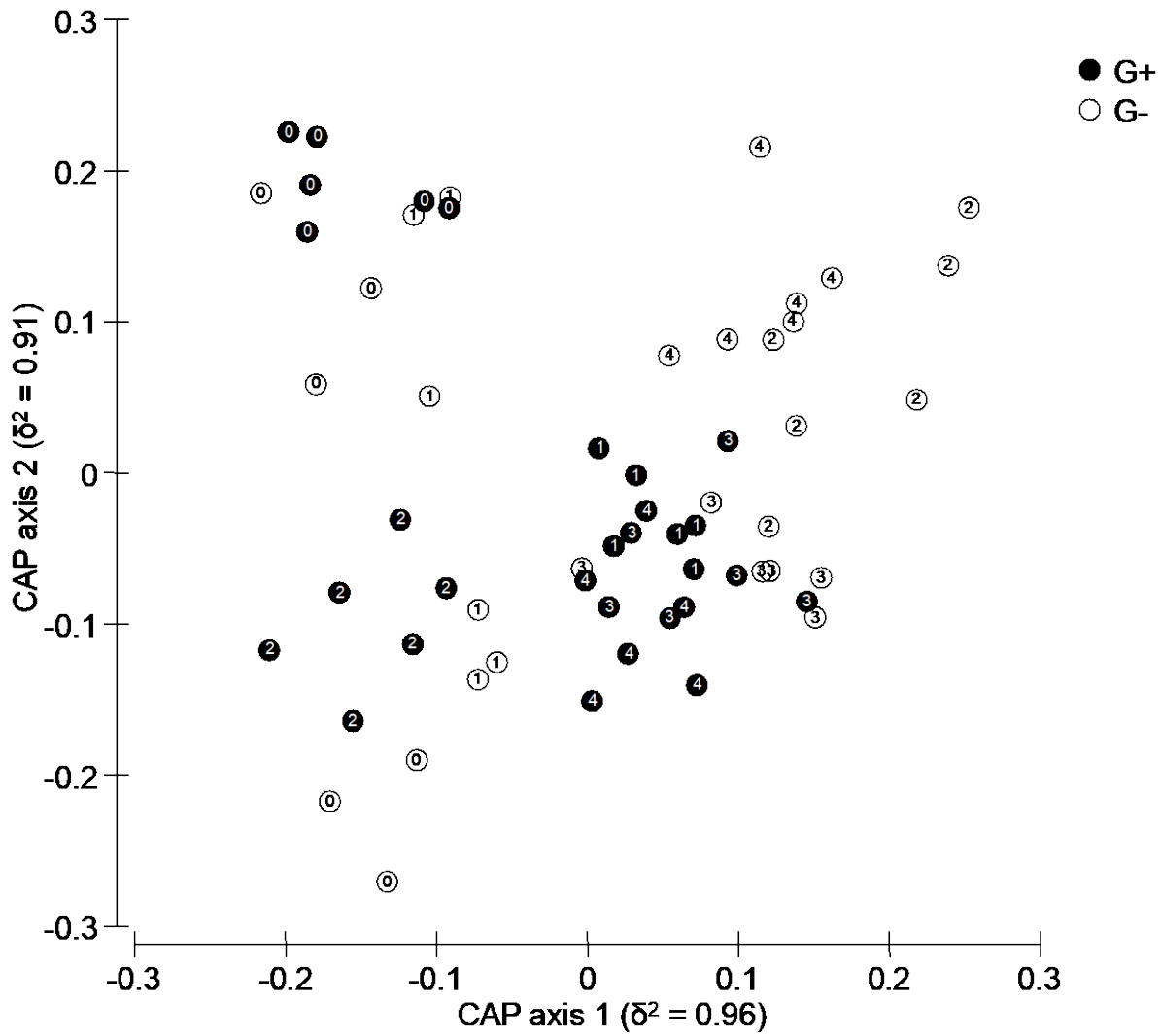


Figure S2. Final state of the rocky reefs characterizing reference (a) and culling (b) sites (i.e. after 3 years from the intervention).

