

SUPPLEMENTARY MATERIAL FOR THE MANUSCRIPT

**ITS1 metabarcoding reveals low specificity of lichen mycobiomes at a
local scale**

by

Fernando FERNÁNDEZ-MENDOZA, Antonia FLEISCHHACKER, Theodora
KOPUN, Martin GRUBE, Lucia MUGGIA

September 2016

INDEX OF TABLES

Supplementary table 1. Structure of the dataset accounting for the percentage of reads assignable to the mycobiome, the mycobiont and considered as spurious reads for the ITS1 and non-ITS sample fractions.	9
Supplementary table 2. Taxonomic composition of the dataset at order level, Ascomycota. Numbers reflect the raw number of reads.	10
Supplementary table 3. Taxonomic composition of the dataset at order level, Basidiomycota. Numbers reflect the raw number of reads.	11
Supplementary table 4. Expanded representation of network statistics for unipartite and bipartite sample networks, as well as for bipartite host species vs. OTUs.	27
Supplementary table 5. Standarized and raw specialization indices calculatæd for each lichen host species (A) in the networks, as well as for those lichenicolous fungi identifiable as the main MOTUs at sample level (B). The values of d' range between 0, mening no specialization and 1 meaning perfect specialization.	28

INDEX OF FIGURES

Supplementary Figure 1. Schematic representation of the analytical pipeline. Files referred by filename can be found on the gitHub repository. Main pipeline is highlighted green and green names in italics highlight the main resulting data structures. Taxonomic filtering was done manually and included multiple manual and semiautomated validation steps.	7
Supplementary Figure 2. Summary of reads included and excluded in the analyses per sample. Upper plots show read counts per sample, lower plots the same figures but normalized per sample. The left subplot includes reads that passed the filtering for fungal ITS1; the right subplot those that did not pass the filter. Colours highlight the reads excluded as they do not contain ITS1 fragments (grey), they belong to the host lichen fungus of the sample (blue), or to those of other lichen species found in the neighbouring community (green). Yellow reads are those included in further analyses.	8
Supplementary Figure 3. Proportion of reads from the ITS1 and from the untrimmed dataset assigned to fungal Divisions. Sequences were identified by blasting to the UNITE database (ITS1) and to NCBI database (complete).	12

Supplementary Figure 4. Summary of the taxonomic assignment of the dataset to the main fungal classes, orders and families. Bars reflect the proportion of reads from the ITS1 dataset per sample.	13
Supplementary Figure 5. Taxonomic assignment of the dataset to fungal classes, in the ITS1 and untrimmed datasets.	14
Supplementary Figure 6. Taxonomic assignment of the dataset to fungal orders, in the ITS1 and untrimmed datasets.	15
Supplementary Figure 7. Taxonomic assignment of the dataset to fungal families, in the ITS1 and untrimmed datasets.	16
Supplementary Figure 8. Graphic representation of the multidimensional scaling axes based on β -diversity estimates between samples. Samples are sorted by whether they are symptomatically infected by lichenicolous fungi. The position of the two centroids (red dots) of the two sets of samples as well as the overlap between the two groups reinforces the idea that all samples form a single group.	17
Supplementary Figure 9. Hierarchical clustergram (dendrogram) showing similarity in OTU composition between samples. Hierarchical clustering was carried out on a pairwise Bray-Curtis distance matrix between samples. The long branch lengths of the cladogram reinforce the lack of structure of the dataset. Bray-Curtis distances tend to overestimate the differences due to minority species.....	18
Supplementary Figure 10. Rarefaction curves at order level. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.	19
Supplementary Figure 11. Rarefaction curves at family level. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.	20
Supplementary Figure 12. Rarefaction curves at genus level. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.	21

Supplementary Figure 13. Rarefaction curves at species (pseudospecies) level. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.....22

Supplementary Figure 14. Rarefaction curves at OTU level, including singletons. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.23

Supplementary Figure 15. Rarefaction curves at OTU level, excluding singletons. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.24

Supplementary Figure 16. Phylogenetic placement of OTUs identified as A) *Endococcus macrosporus*. B) *Skyttea tephromelarum* and C) *Taeniolella sp.* The presented topologies are maximum likelihood topologies calculated in RaxML, numbers above the branches reflect bootstrap support.....25

Supplementary Figure 17. Bipartite network at sample level including all non-lichen OTUs.29

Supplementary Figure 18. Bipartite network at sample level of non-lichen OTUs above a sample specific threshold.29

Supplementary Figure 19. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level of non-lichen OTUs above a sample specific threshold.....30

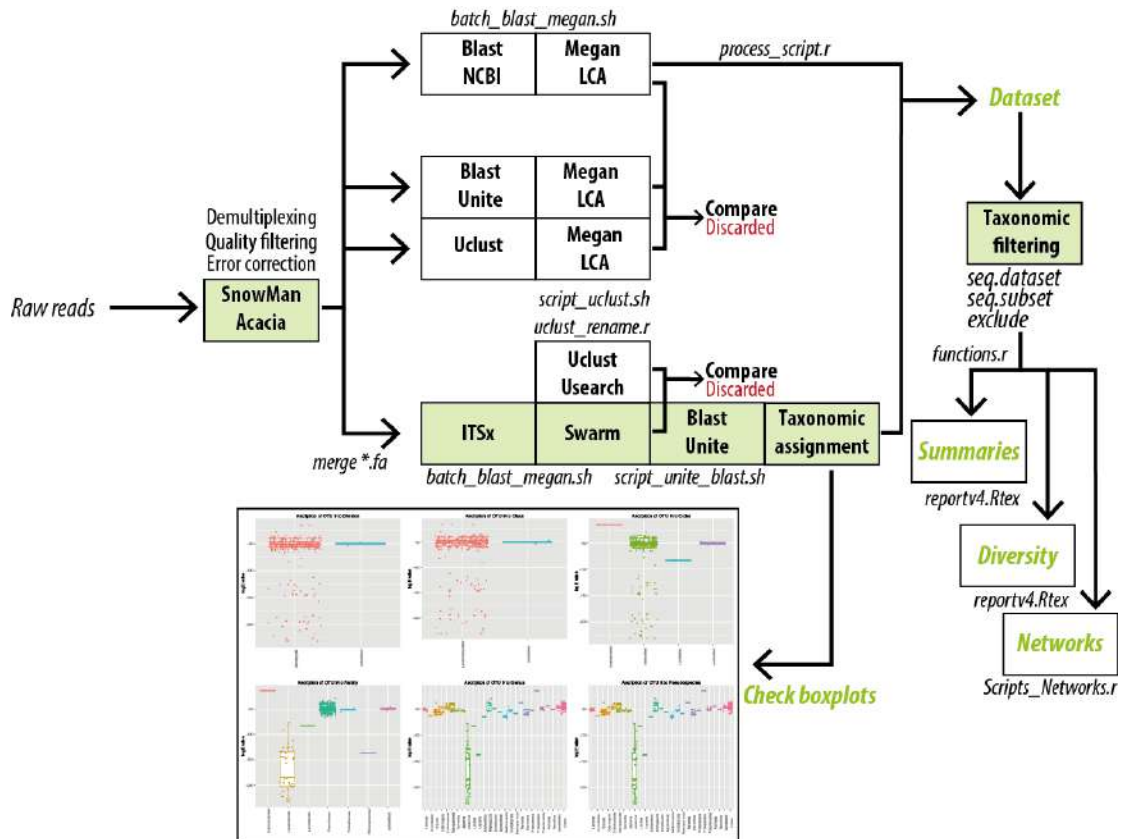
Supplementary Figure 20. Bipartite network at sample level using only OTUs identified as Tremellales.....31

Supplementary Figure 21. Bipartite network at species level using only those OTUs identified as Tremellales found above a sample specific threshold.31

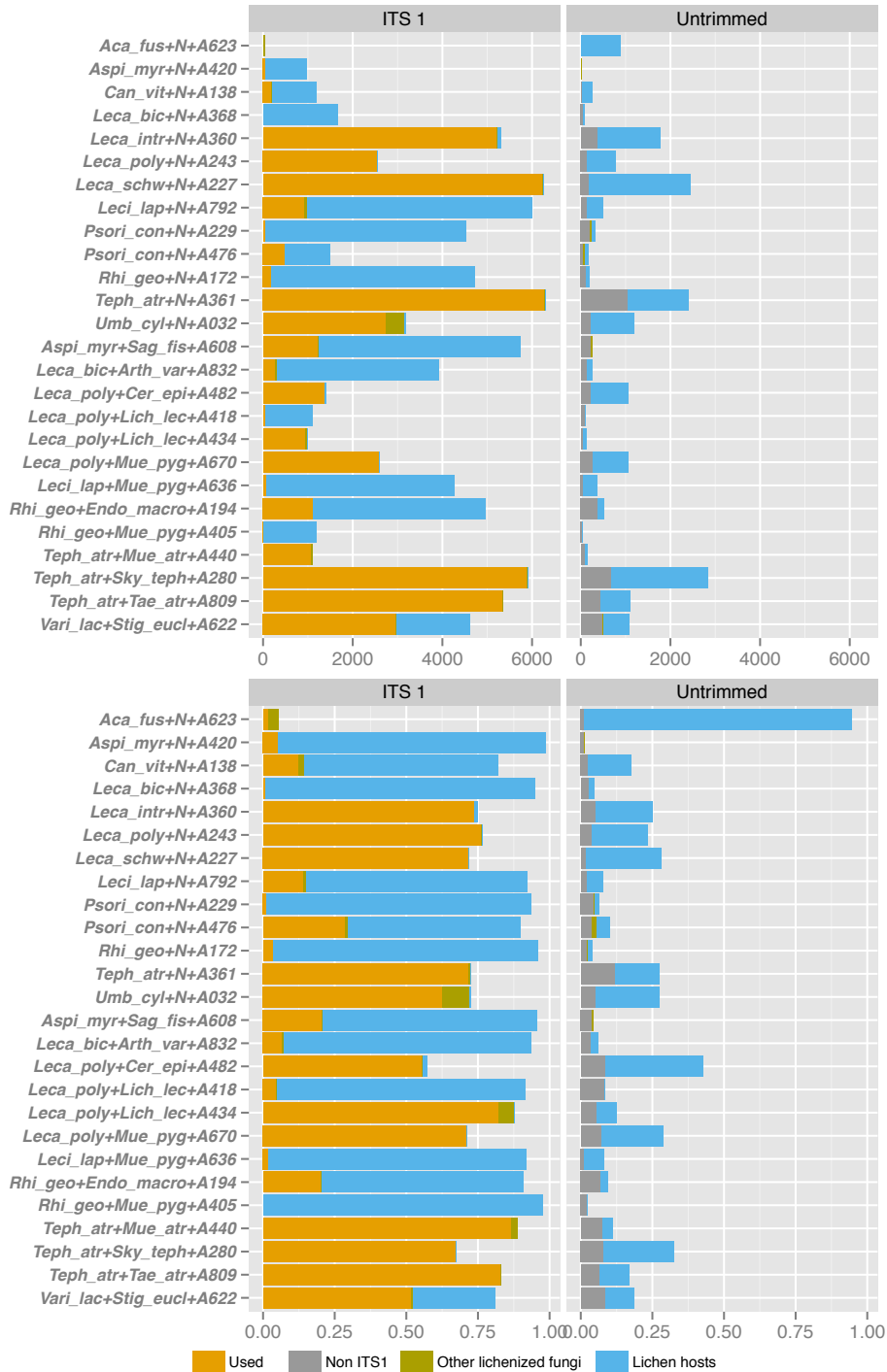
Supplementary Figure 22. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Tremellales.....	32
Supplementary Figure 23. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Tremellales sequence above a sample specific threshold.....	32
Supplementary Figure 24. Bipartite network at sample level using only OTUs identified as Capnodiales.....	33
Supplementary Figure 25. Bipartite network at sample level using only those OTUs identified as Capnodiales above a sample specific threshold.....	33
Supplementary Figure 26. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Capnodiales.....	34
Supplementary Figure 27. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Capnodiales sequence above a sample specific threshold.....	34
Supplementary Figure 28. Bipartite network at sample level using only OTUs identified as Botryosphaerales.....	35
Supplementary Figure 29. Bipartite network at sample level using only those OTUs identified as Botryosphaerales above a sample specific threshold.....	35
Supplementary Figure 30. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Botryosphaerales.....	36
Supplementary Figure 31. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Botryosphaerales sequence above a sample specific threshold.	36
Supplementary Figure 32. Bipartite network at sample level using only OTUs identified as Chaetothyriales.	37

Supplementary Figure 33. Bipartite network at sample level using only those OTUs identified as Chaetothyriales found above a sample specific threshold.	37
Supplementary Figure 34. Matrix representation showing nestedness and compartmentalization of the bipartite network at sample level using all OTUs identified as Chaetothyriales.	38
Supplementary Figure 35. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Chaetothyriales sequence above a sample specific threshold. ...	38
Supplementary Figure 36. Bipartite network at species level including all non-lichen OTUs.	39
Supplementary Figure 37. Bipartite network at species level of non-lichen OTUs above a sample specific threshold.	39
Supplementary Figure 38. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level of non-lichen OTUs above a sample specific threshold.	40
Supplementary Figure 39. Bipartite network at species level using only OTUs identified as Tremellales.	41
Supplementary Figure 40. Bipartite network at species level using only those OTUs identified as Tremellales found above a sample specific threshold.	41
Supplementary Figure 41. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Tremellales.	42
Supplementary Figure 42. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Tremellales sequence above a sample specific threshold.	42
Supplementary Figure 43. Bipartite network at species level using only OTUs identified as Capnodiales.	43
Supplementary Figure 44. Bipartite network at species level using only those OTUs identified as Capnodiales found above a sample specific threshold.	43

Supplementary Figure 45. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Capnodiales.....	44
Supplementary Figure 46. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Capnodiales sequence above a sample specific threshold.....	44
Supplementary Figure 47. Bipartite network at species level using only OTUs identified as Botryosphaerales.....	45
Supplementary Figure 48. Bipartite network at species level using only those OTUs identified as Botryosphaerales found above a sample specific threshold.....	45
Supplementary Figure 49. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Botryosphaerales.....	46
Supplementary Figure 50. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Botryosphaerales sequence above a sample specific threshold.....	46
Supplementary Figure 51. Bipartite network at species level using only OTUs identified as Chaetothyriales.....	47
Supplementary Figure 52. Bipartite network at species level using only those OTUs identified as Chaetothyriales found above a sample specific threshold.	
Supplementary Figure 53. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Chaetothyriales.....	47
Supplementary Figure 54. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Chaetothyriales sequence above a sample specific threshold. ...	48
Supplementary Figure 55. The following pages contain the output of the quality assessment of the raw data obtained using FastQC.....	49



Supplementary Figure 1. Schematic representation of the analytical pipeline. Files referred by filename can be found on the gitHub repository. Main pipeline is highlighted green and green names in italics highlight the main resulting data structures. Taxonomic filtering was done manually and included multiple manual and semiautomated validation steps.



Supplementary Figure 2. Summary of reads included and excluded in the analyses per sample. Upper plots show read counts per sample, lower plots the same figures but normalized per sample. The left subplot includes reads that passed the filtering for fungal ITS1; the right subplot those that did not pass the filter. Colours highlight the reads excluded as they do not contain ITS1 fragments (grey), they belong to the host lichen fungus of the sample (blue), or to those of other lichen species found in the neighbouring community (green). Yellow reads are those included in further analyses.

	Mycobiome		Spurious		Mycobiont		All
	ITS1	Non	ITS1	Non	ITS1	Non	
Aca_fus+N+A623	2	1.1	3.4	0	0.1	93.4	93.5
Aspi_myr+N+A420	5.1	1.1	0.2	0.2	93.4	0	93.4
Can_vit+N+A138	12.2	2.5	2.3	0.2	67.7	15.1	82.8
Leca_bic+N+A368	0.9	2.9	0	0	94.2	2	96.2
Leca_intr+N+A360	73.7	5.4	0.1	0	1	19.7	20.7
Leca_poly+N+A243	76.3	4.1	0.2	0	0.1	19.4	19.5
Leca_schw+N+A227	71.6	2	0.2	0	0	26.1	26.1
Leci_lap+N+A792	14.1	2.3	1.1	0	77.1	5.5	82.6
Psori_con+N+A229	1.1	4.5	0.1	0.5	92.3	1.5	93.8
Psori_con+N+A476	28.8	3.9	0.8	1.5	60.3	4.7	65
Rhi_geo+N+A172	3.6	2.3	0.2	0	92.3	1.7	94
Teph_atr+N+A361	71.9	12.1	0.4	0	0	15.5	15.5
Umb_cyl+N+A032	62.8	5.4	9.2	0	0.6	22	22.6
Aspi_myr+Sag_fis+A608	20.5	3.8	0.5	0.4	74.8	0	74.8
Leca_bic+Arth_var+A832	6.5	3.6	0.8	0	86.4	2.7	89.1
Leca_poly+Cer_epi+A482	55.6	8.6	0.2	0	1.6	34.1	35.7
Leca_poly+Lich_lect+A418	4.8	8.1	0.1	0	86.7	0.3	87
Leca_poly+Lich_lect+A434	82.2	5.6	5.3	0	0.1	6.8	6.9
Leca_poly+Mue_pyg+A670	71	7.3	0.1	0	0	21.5	21.5
Leci_lap+Mue_pyg+A636	1.8	1	0	0.2	90.1	6.9	97
Rhi_geo+Endo_macro+A194	20.3	6.9	0.1	0	70.4	2.4	72.8
Rhi_geo+Mue_pyg+A405	0.2	2.2	0	0	97.4	0.2	97.6
Teph_atr+Mue_atr+A440	86.4	7.8	2.2	0	0	3.5	3.5
Teph_atr+Sky_teph+A280	67.3	7.8	0.2	0	0.1	24.6	24.7
Teph_atr+Tae_atr+A809	82.7	6.7	0.3	0	0	10.3	10.3
Vari_lac+Stig_eucl+A622	51.8	8.7	0.6	0.1	28.8	10	38.8
Average	37.51	4.91	1.10	0.12	42.90	13.46	56.36
							1.22

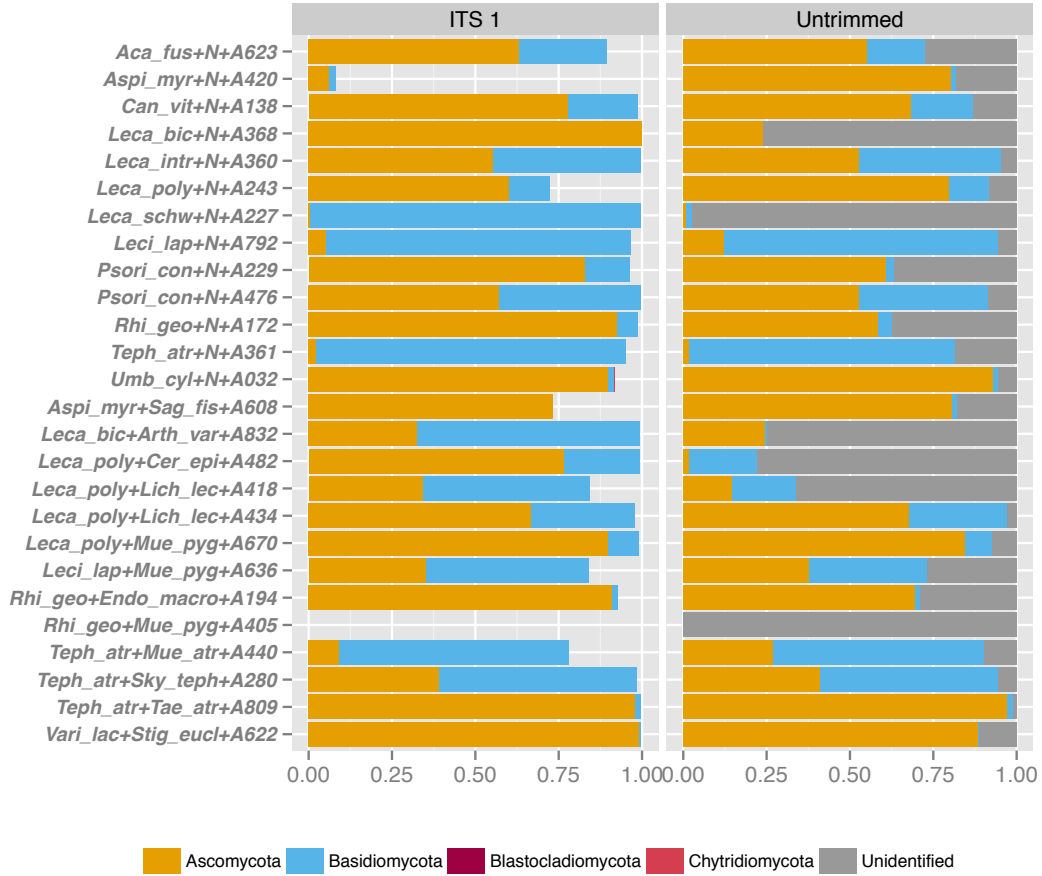
Supplementary table 1. Structure of the dataset accounting for the percentage of reads assignable to the mycobiome, the mycobiont and considered as spurious reads for the ITS1 and non-ITS sample fractions.

Ascomycota	A623	A420	A138IN	A368	A360	A243	A227	A792	A229	A476	A172	A361	A032	A608	A832	A482	A418	A434	A670	A636	A194	A405	A440	A280	A809	A622	
Dothideomycetes	8	3	60	6	588	223	18	28	15	42	32	53	359	155	28	228	4	126	353	14	178	.	44	463	862	409	
Botryosphaeriales	.	1	3	.	44	2	4	.	.	3	7	4	204	.	.	218	.	.	4	.	8	.	4	4	.	7	
Capnodiales	.	.	16	.	389	42	4	6	.	7	13	45	53	1	8	4	2	48	63	5	4	.	26	179	859	391	
Myrangiiales	.	.	1	.	1	2	2	1	.	1	3	23	.	.	
Pleosporales	2	17	1	3	.	1	.	2	42	1	.	1	.	.	.	3	1	.	
unidentified	
Eurotiomycetes	5	2	39	.	30	132	3	7	2	29	12	1	22	153	13	1	.	1	159	7	166	.	4	158	1	6	
Chaetothyriales	5	2	39	.	29	132	3	7	1	29	12	1	22	153	13	1	.	1	159	7	1	.	4	156	1	6	
Eurotiales
Verrucariales	1	.	.	1	165	
Lecanoromycetes	3	.	1	6	1	17	3	10	13	.	.	.	24	1	.	5	.	5	124	1	.	.	1	93	.	5	
Candelariales	3	.	1	1	
Lecanorales	3	.	.	.	1	11	2	9	13	.	.	.	11	1	.	3	.	.	122	1	
Lecideales	1	5	3	
Ostropales	1	92	.	
Peltigerales	.	.	.	6	5	2	
Pertusariales	1	
Rhizocarpaceles	7	1	
Umbilicariales	1	
unidentified	1	1	1	.	.	
Leotiomycetes	2	
Helotiales	4	1	5	2	3	.	.	.	
Saccharomycetes	4	1	5	2	3	.	.	.	
Saccharomycetales	1	
Sordariomycetes	2	8	2	
Diaporthales	
Hypocreales	5	
Taphrinales	7	3	2	
Unidentified Ascomycete	2	1	.	1	.	1	.	.	7	.	2	67	1		

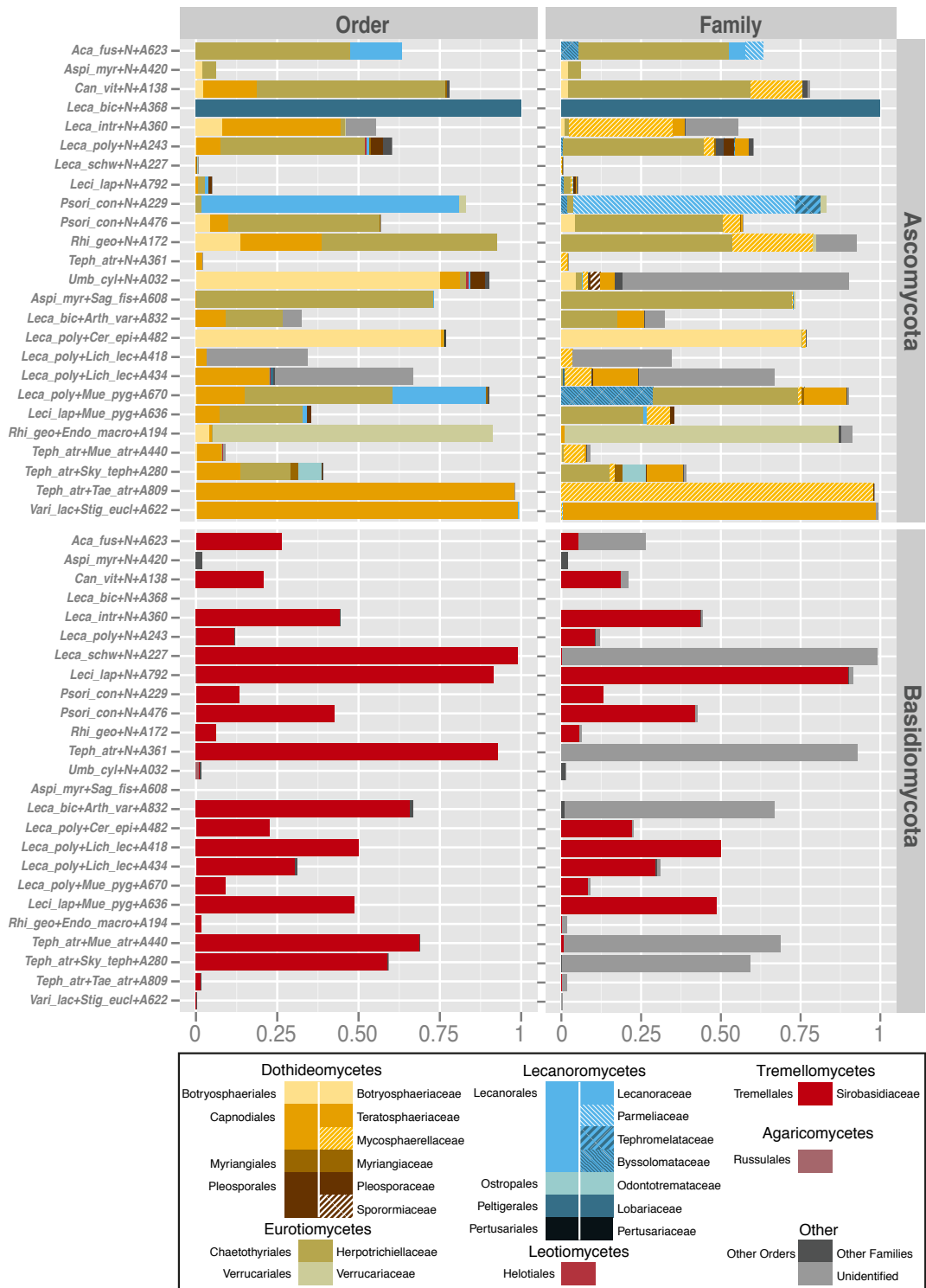
Supplementary table 2. Taxonomic composition of the dataset at order level, Ascomycota. Numbers reflect the raw number of reads.

A623	4	1	13	. . .	276	51	434	176	4	40	3	506	19	. . .	31	52	7	62	48	7	9	. . .	98	319	17	6	
A420	. . .	1	1	1	. . .	1	17	. . .	3	6	7	7	9	. . .	1	14	1	1	
A138	5	. . .	2	6	. . .	1	
A368	1	1	1	
A360	1	1	1	. . .	
A243	6	3	
A227	5	2	
A792	
A229	
A476	
A172	
A361	
A032	
A608	
A832	
A482	
A418	
A434	
A670	
A636	
A194	
A405	
A280	
A809	
A622	
Basidiomycota																											
Agaricomycetes																											
Agaricales																											
Auriculariales																											
Cantharellales																											
Corticiales																											
Hymenochaetales																											
Polyporales																											
Russulales																											
Sebacinales																											
Microbotryomycetes																											
Sporidiobolales																											
Tremellomycetes																											
Cystofilobasidiales																											
Tremellales																											
unidentified																											
Chytridiomycota																											
Blastocladiomycetes																											
Blastocladiiales																											
Unidentified																											

Supplementary table 3. Taxonomic composition of the dataset at order level, Basidiomycota. Numbers reflect the raw number of reads.



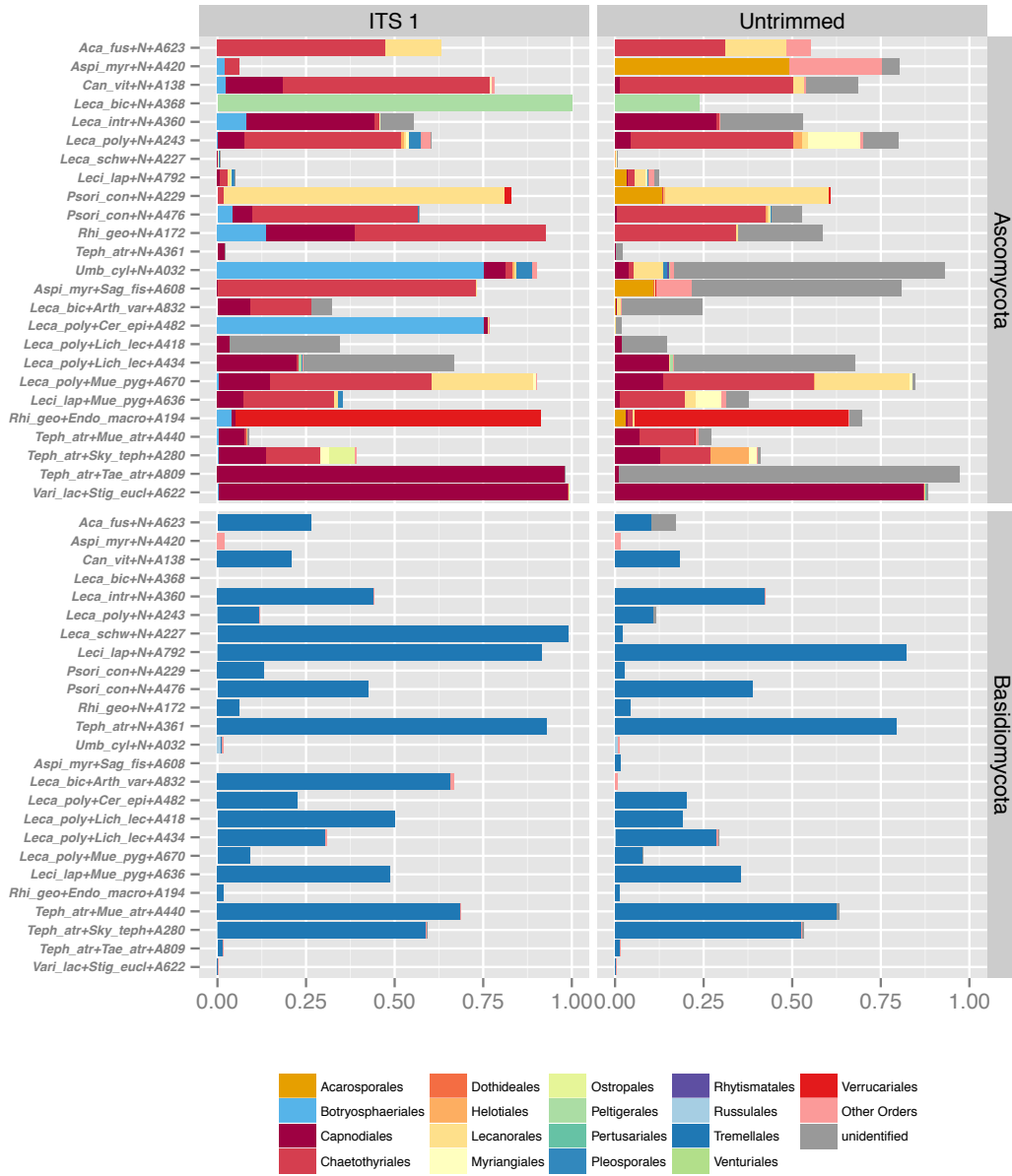
Supplementary Figure 3. Proportion of reads from the ITS1 and from the untrimmed dataset assigned to fungal Divisions. Sequences were identified by blasting to the UNITE database (ITS1) and to NCBI database (complete).



Supplementary Figure 4. Summary of the taxonomic assignment of the dataset to the main fungal classes, orders and families. Bars reflect the proportion of reads from the ITS1 dataset per sample.



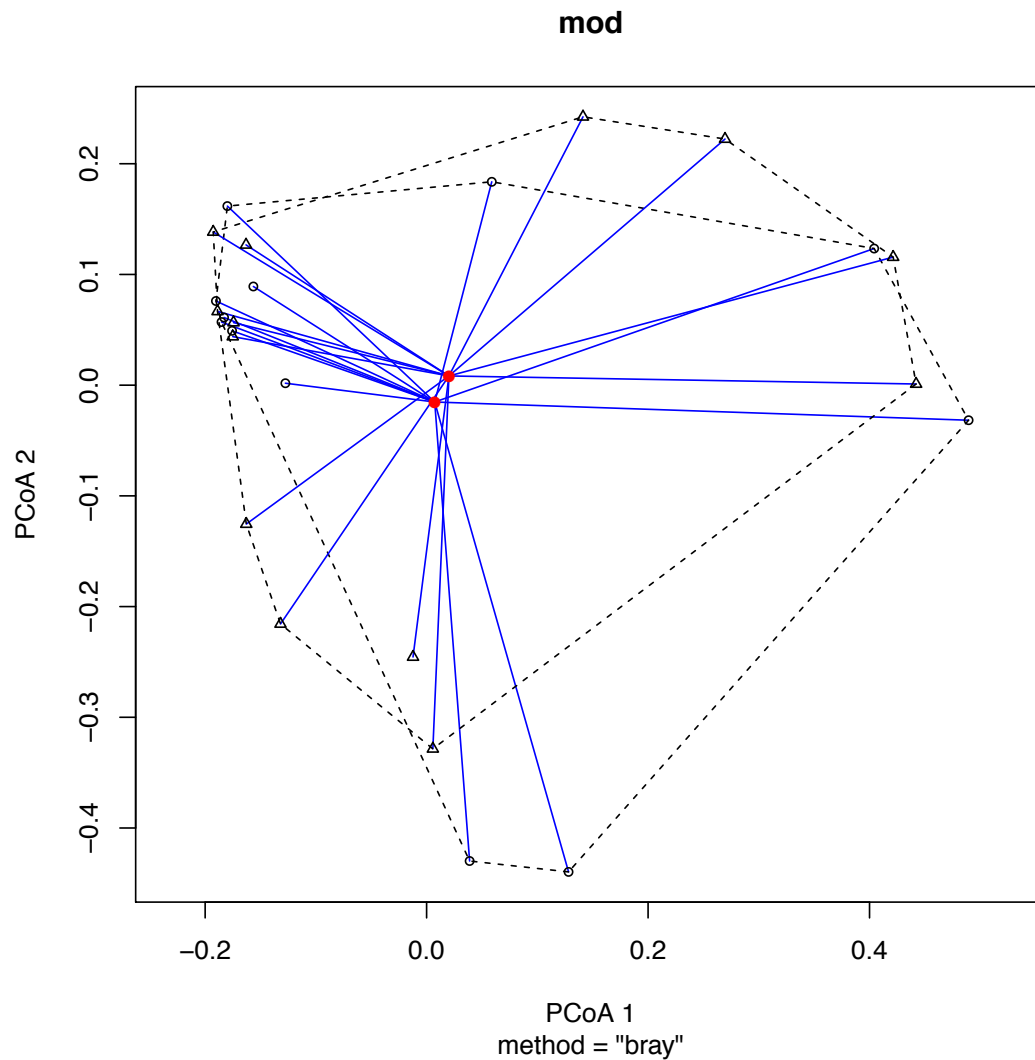
Supplementary Figure 5. Taxonomic assignment of the dataset to fungal classes, in the ITS1 and untrimmed datasets.



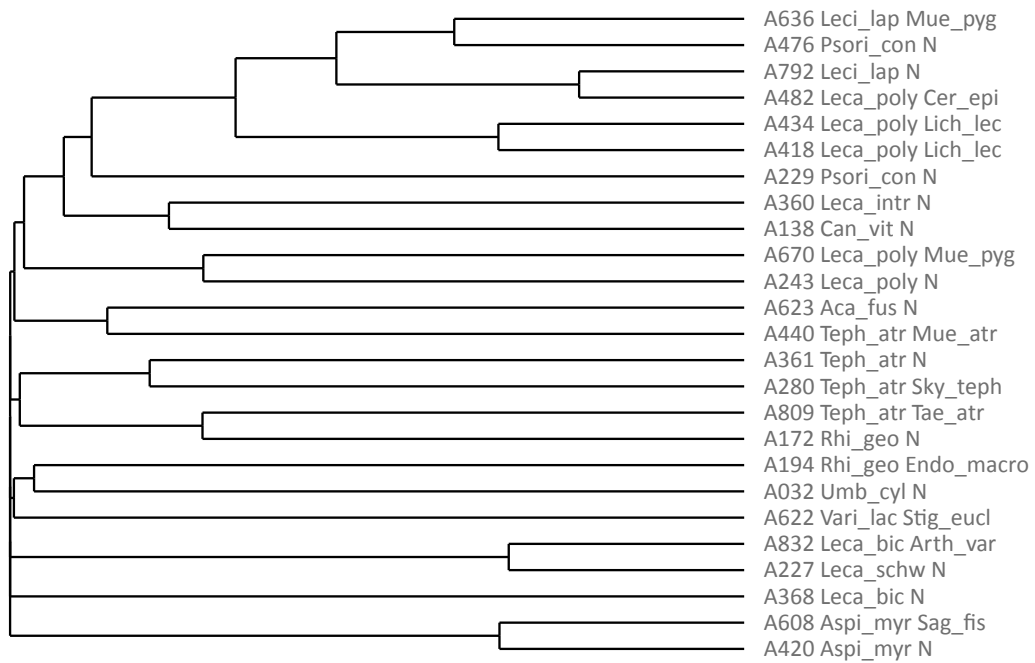
Supplementary Figure 6. Taxonomic assignment of the dataset to fungal orders, in the ITS1 and untrimmed datasets.



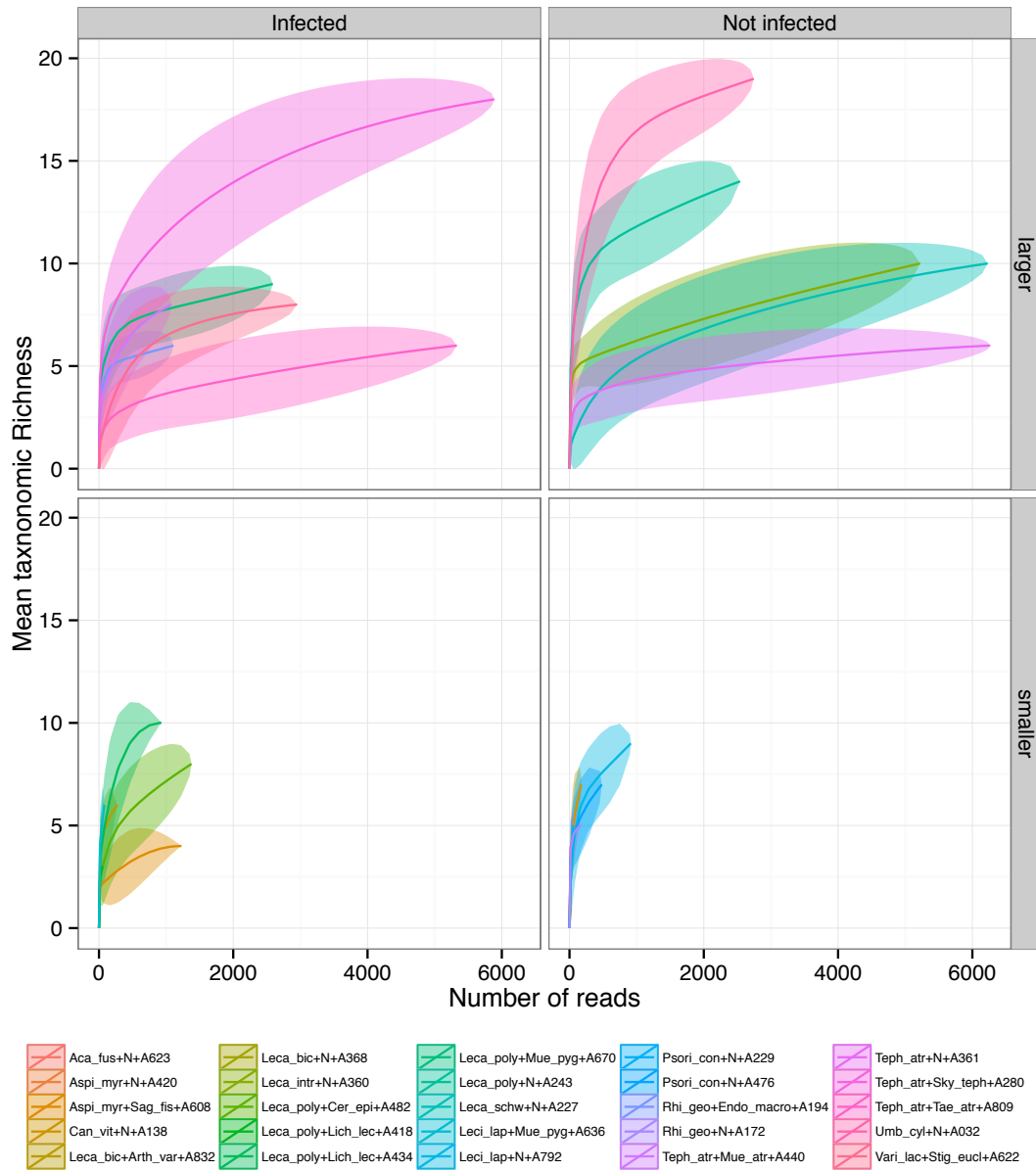
Supplementary Figure 7. Taxonomic assignment of the dataset to fungal families, in the ITS1 and untrimmed datasets.



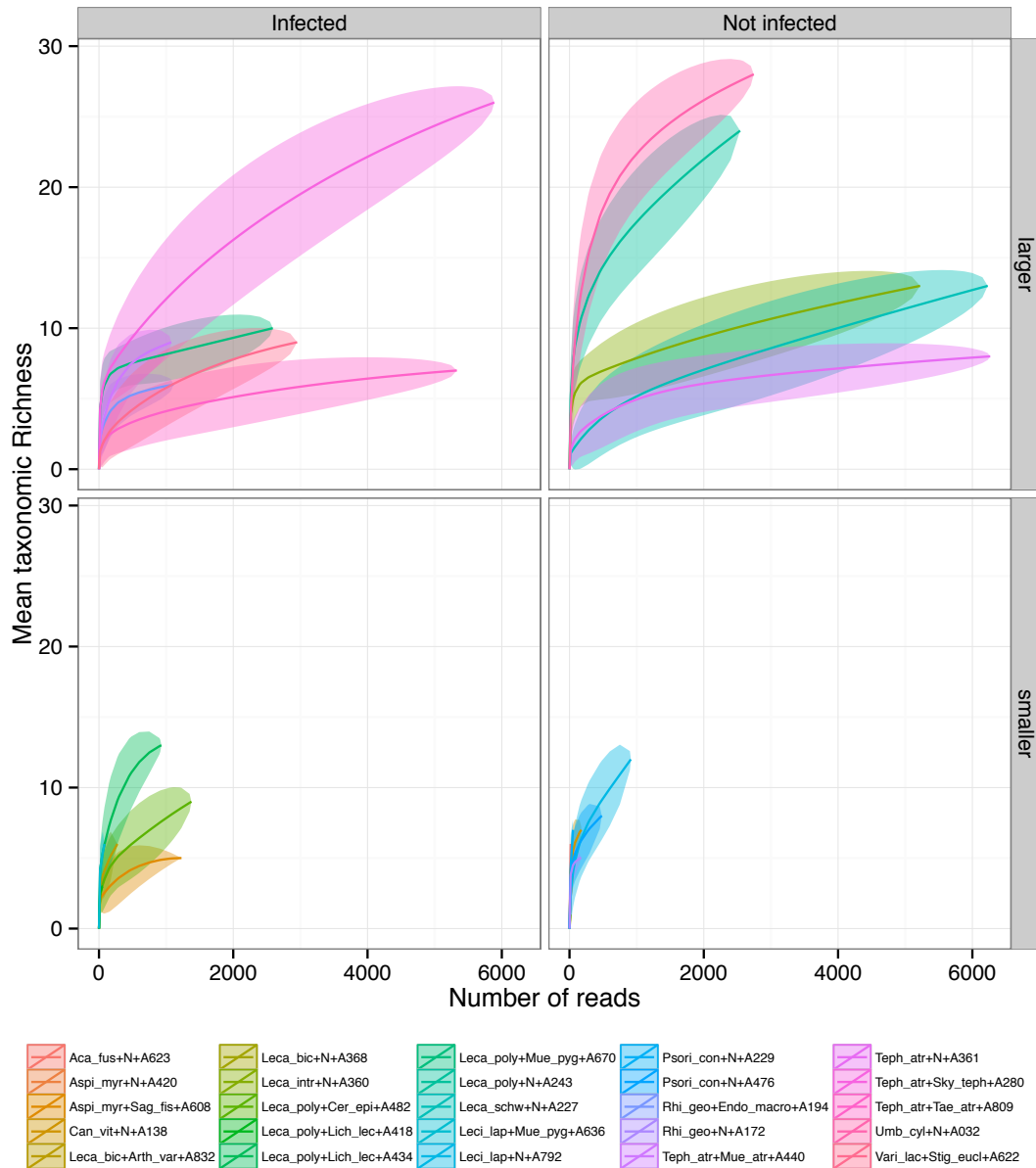
Supplementary Figure 8. Graphic representation of the multidimensional scaling axes based on β -diversity estimates between samples. Samples are sorted by whether they are symptomatically infected by lichenicolous fungi. The position of the two centroids (red dots) of the two sets of samples as well as the overlap between the two groups reinforces the idea that all samples form a single group.



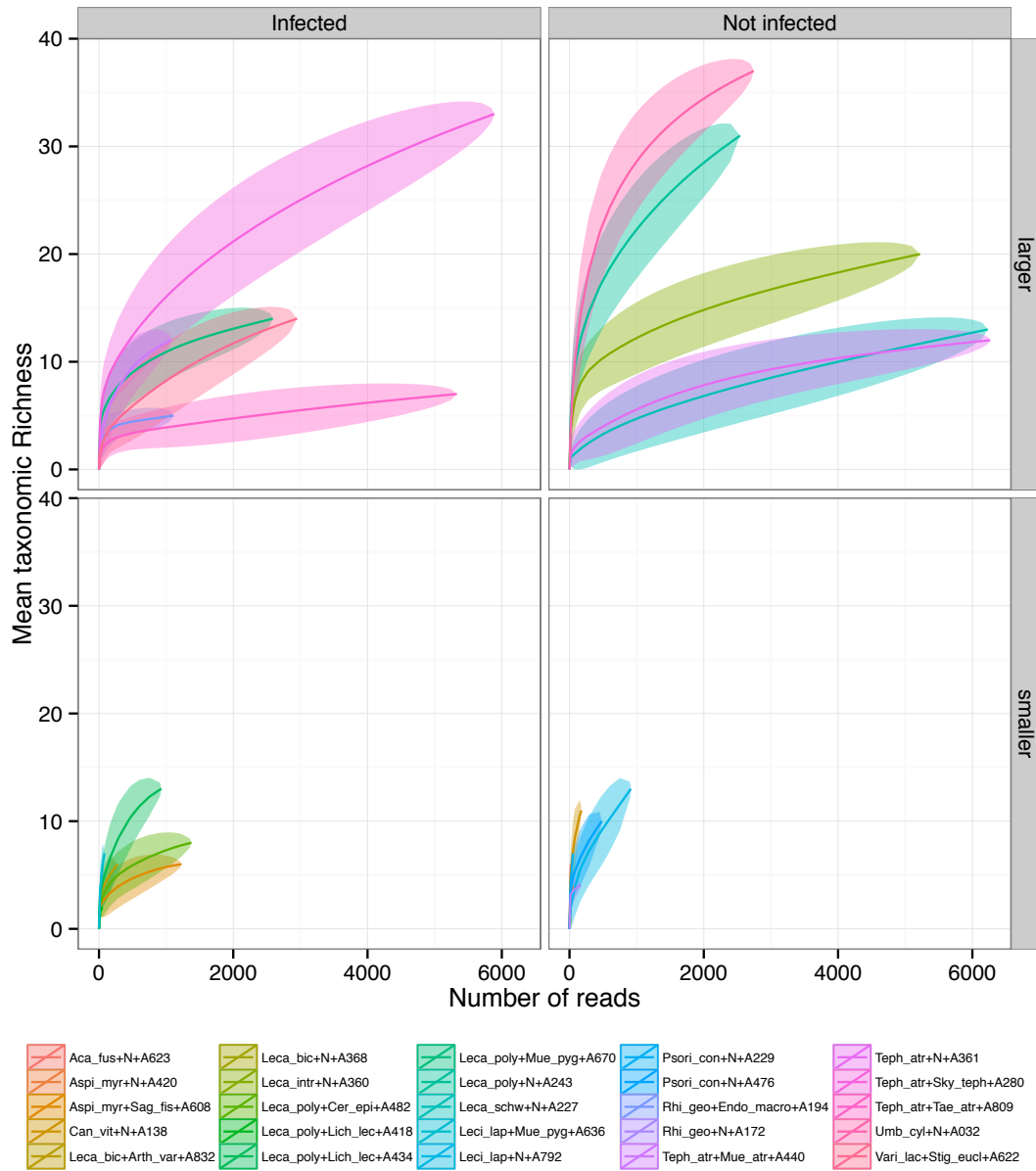
Supplementary Figure 9. Hierarchical clustergram (dendrogram) showing similarity in OTU composition between samples. Hierarchical clustering was carried out on a pairwise Bray-Curtis distance matrix between samples. The long branch lengths of the cladogram reinforce the lack of structure of the dataset. Bray-Curtis distances tend to overestimate the differences due to minority species.



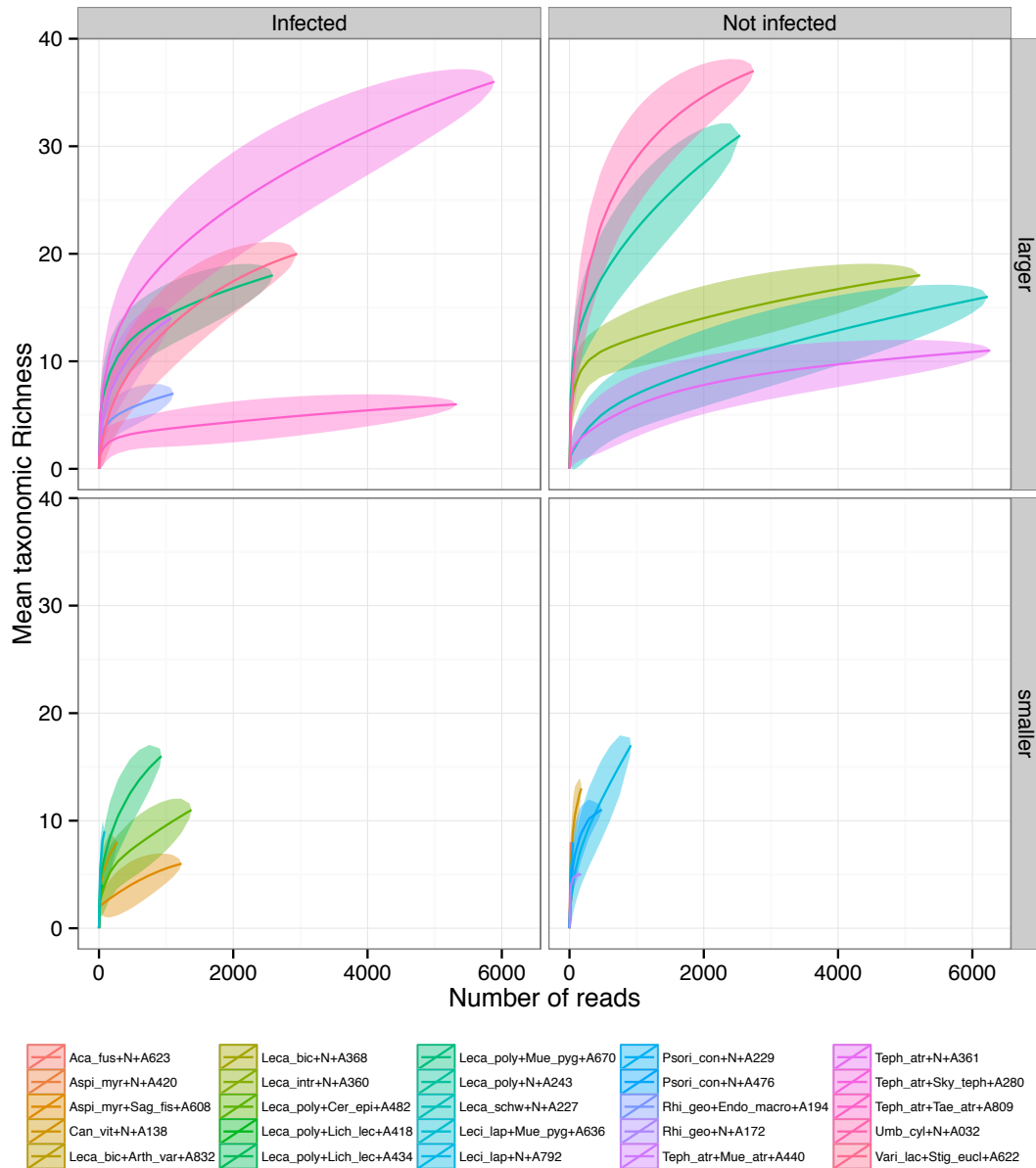
Supplementary Figure 10. Rarefaction curves at order level. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.



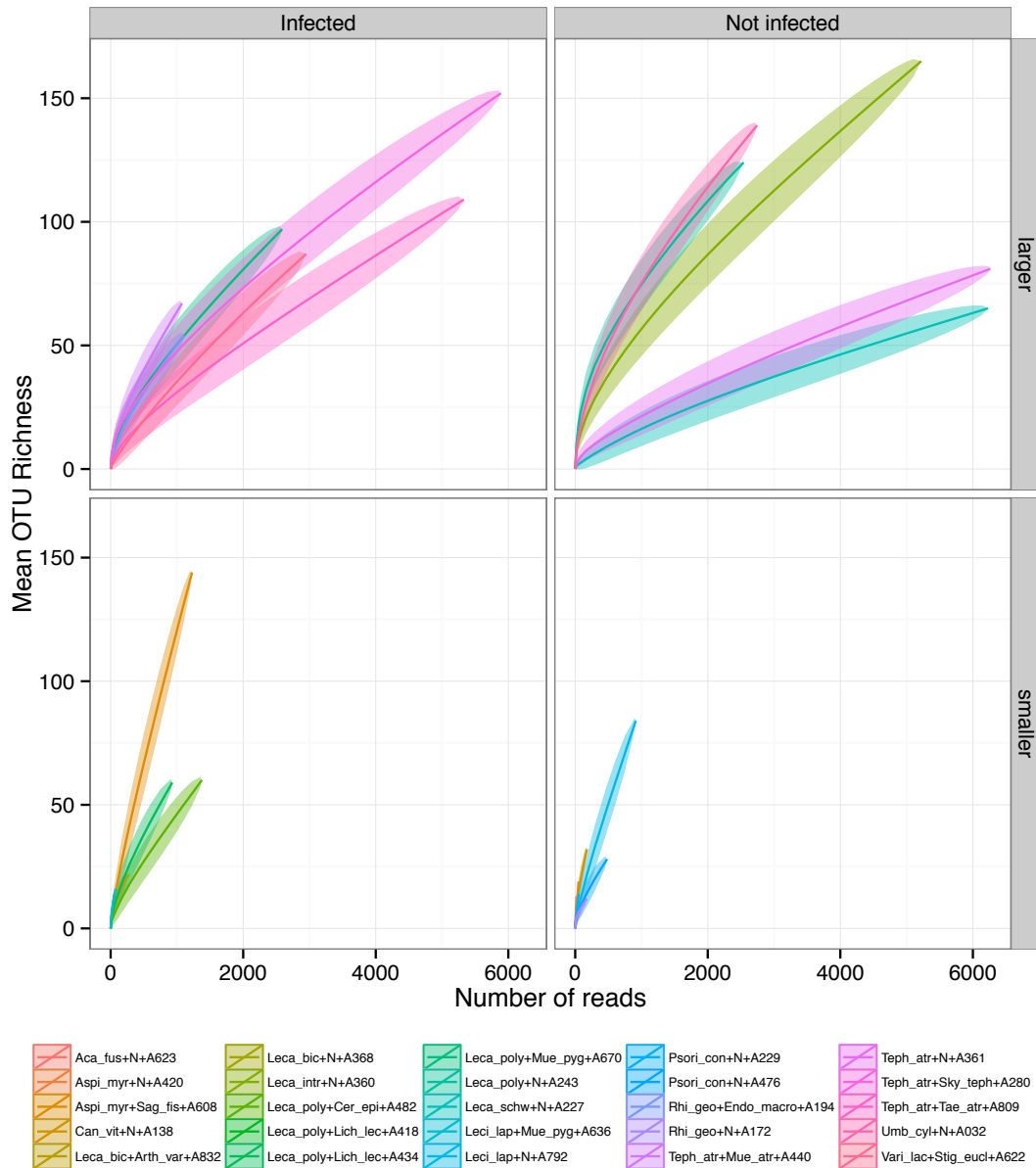
Supplementary Figure 11. Rarefaction curves at family level. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.



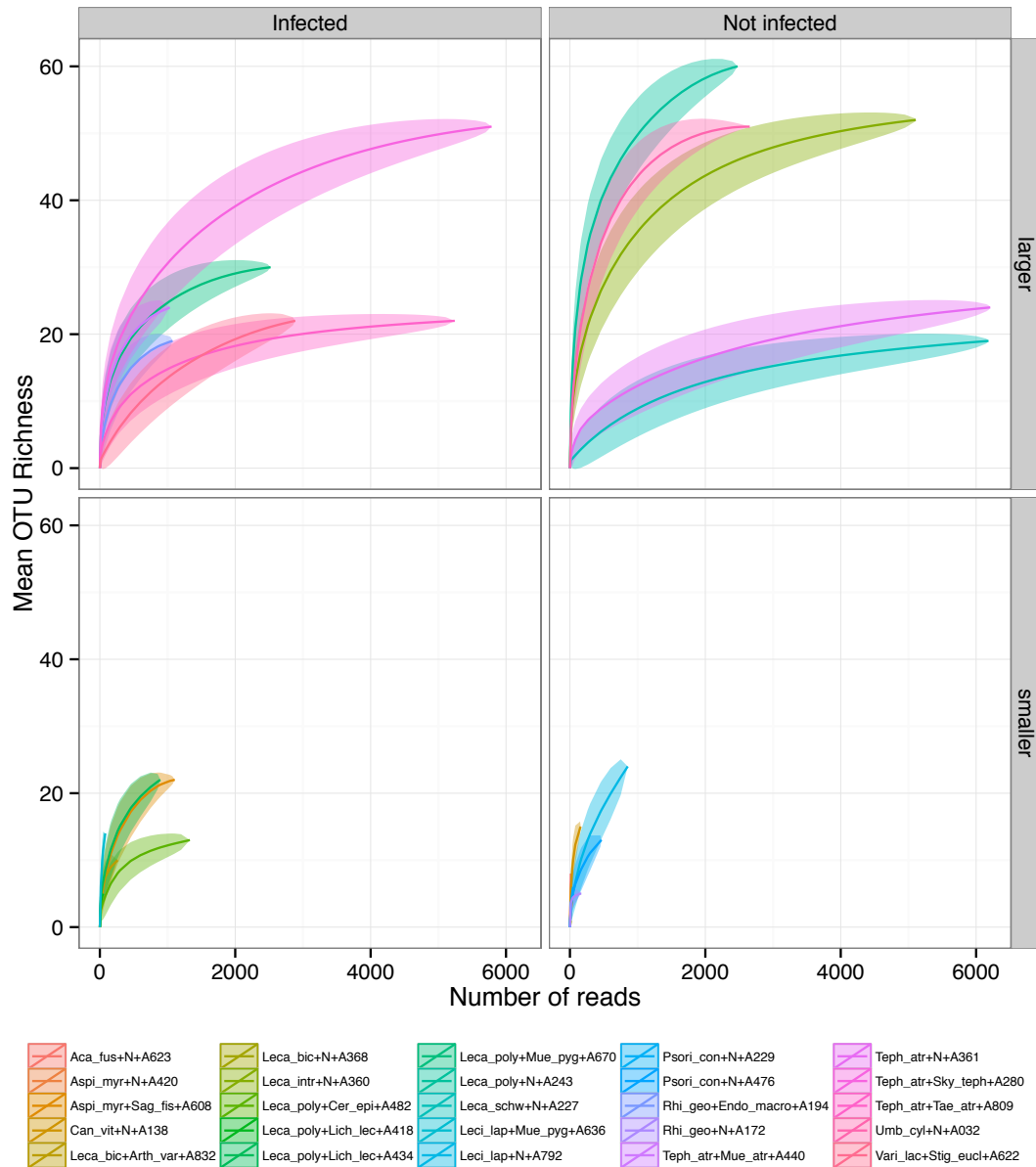
Supplementary Figure 12. Rarefaction curves at genus level. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.



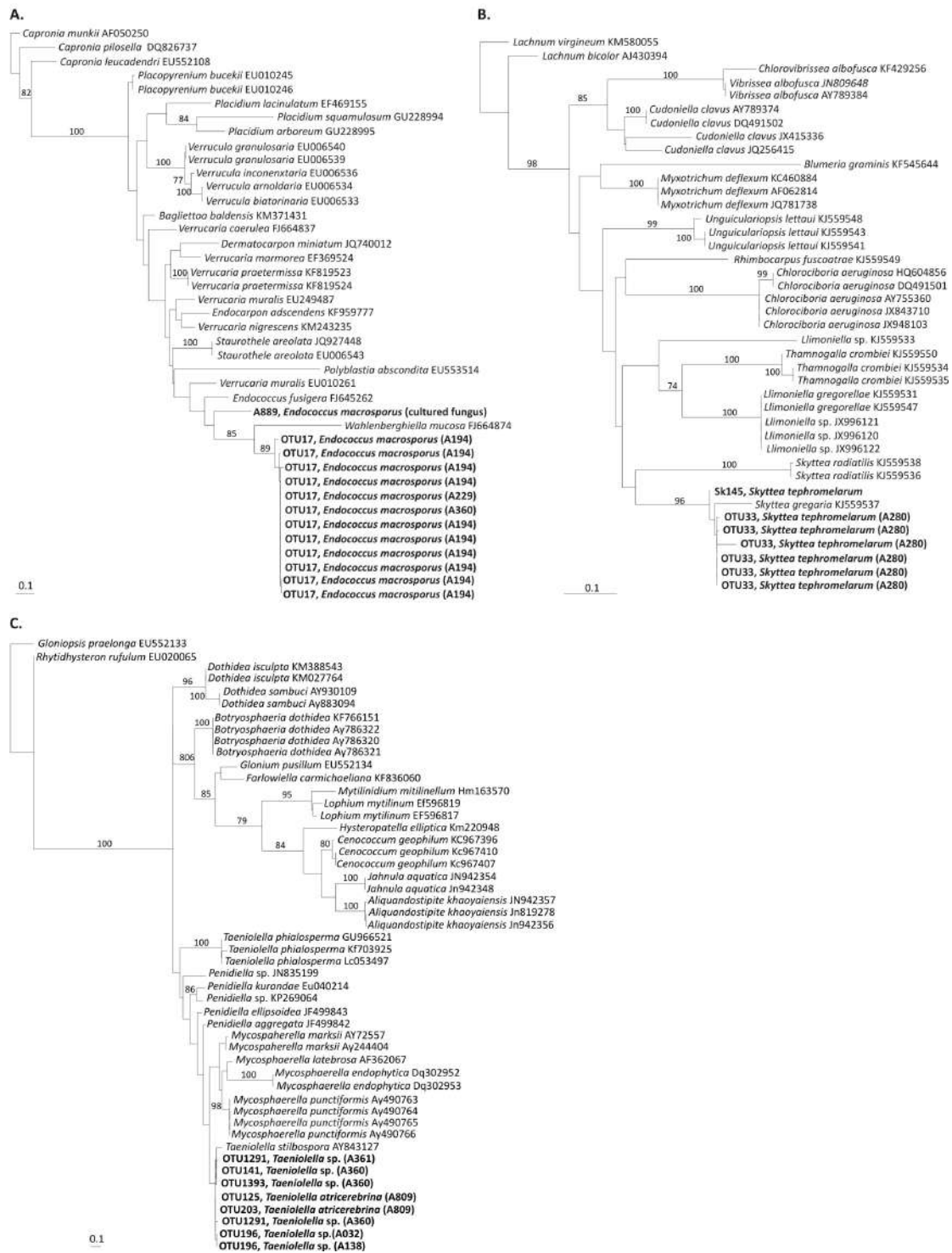
Supplementary Figure 13. Rarefaction curves at species (pseudospecies) level. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.



Supplementary Figure 14. Rarefaction curves at OTU level, including singletons. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.



Supplementary Figure 15. Rarefaction curves at OTU level, excluding singletons. Banana plots of rarefied species richness estimates and (bold line) confidence intervals based on standard error estimates. Symptomatic and non infected samples are presented in separate subplots, as well as are samples with higher and lower number of reads.



Supplementary Figure 16. Phylogenetic placement of OTUs identified as A) *Endococcus macrosporus*. B) *Skyttea tephromelarum* and C) *Taeniolella sp.* The presented topologies are maximum likelihood topologies calculated in RaxML, numbers above the branches reflect bootstrap support.

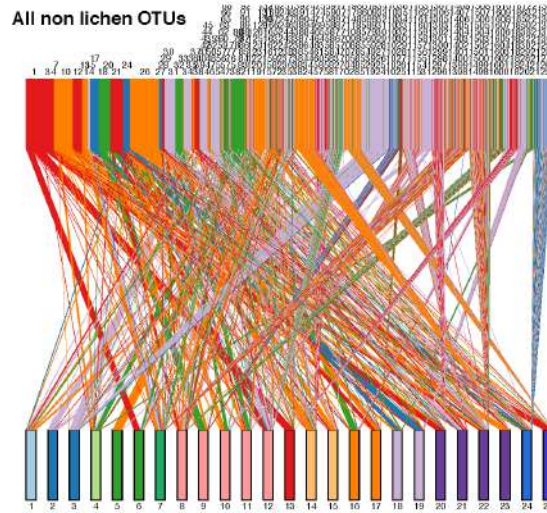
SAMPLES	All		Tremellales		Botryosphaerales		Capnodiales		Chaetothyriales	
	R	T	R	T	R	T	R	T	R	T
Unipartite networks										
Connectance	0.80	0.47	0.64	0.29	0.14	0.06	0.48	0.33	0.42	0.12
Loss of connectance		0.34		0.36		0.08		0.15		0.30
Number of links per sample	9.64	5.60	7.72	3.44	1.72	0.76	5.76	4.00	5.00	1.40
Mean number of common links	15.87	7.40	12.26	3.56	1.67	0.77	8.98	5.66	6.33	1.64
Bipartite networks										
Number of MOTUs	1225	130	219	21	63	6	292	26	153	29
Number of Samples	25	24	23	20	15	10	21	20	23	14
Mean Shared MOTUs per pair of samples	2.95	1.13	1.32	0.54	0.58	0.56	1.07	0.60	0.66	0.47
Mean common samples per pair of MOTUs	0.10	0.22	0.14	0.39	0.57	0.87	0.21	0.24	0.19	0.36
Connectance	0.05	0.08	0.06	0.13	0.09	0.30	0.06	0.10	0.06	0.12
Number of compartments	1	2	1	1	2	2	2	4	3	3
Nestedness	7.16	9.13	5.81	11.37	5.77	27.24	3.81	8.26	4.06	9.19
Weighted nestedness	0.46	0.37	0.59	0.54	0.79	0.26	0.74	0.52	0.62	0.53
Weighted NODF	2.32	7.80	3.89	15.64	10.67	22.22	2.87	12.22	5.71	13.02
cluster coefficient	0.04	0.04	0.04	0.10	0.07	0.35	0.05	0.05	0.04	0.07
cluster coefficient MOTUs	0.31	0.22	0.51	0.35	0.31	0.34	0.43	0.38	0.30	0.28
cluster coefficient Sample	0.05	0.08	0.09	0.15	0.43	0.55	0.17	0.15	0.09	0.15
weighted cluster coefficient MOTUs	0.89	0.65	0.51	0.08	0.34	0.18	0.42	0.20	0.29	0.07
weighted cluster coefficient SAMPLE	0.04	0.22	0.04	0.11	0.04	0.42	0.01	0.12	0.02	0.03
C score MOTU	0.91	0.83	0.86	0.73	0.44	0.58	0.79	0.81	0.82	0.68
C score SAMPLE	0.86	0.77	0.66	0.62	0.70	0.57	0.58	0.54	0.73	0.67
SPECIES										
Bipartite networks										
Number of MOTUs	1225	176	219	41	63	6	292	35	153	25
Number of Species	13	12	13	10	10	6	12	8	13	9
Connectance	0.09	0.11	0.09	0.14	0.13	0.44	0.10	0.17	0.10	0.17
Weighted connectance	0.00	0.02	0.01	0.04	0.03	0.17	0.01	0.05	0.01	0.05
Number of compartments	1	1	1	1	2	1	2	2	1	1
nestedness	11.95	8.44	10.24	10.86	11.55	21.94	8.99	16.53	9.84	12.53
weighted nestedness	0.53	0.77	0.63	0.85	0.78	0.27	0.75	0.68	0.57	0.72
weighted NODF	4.19	20.06	5.06	21.20	11.02	31.11	5.07	17.01	9.40	27.61
Web asymmetry	0.98	0.87	0.89	0.61	0.73	0.00	0.92	0.63	0.84	0.47
Links per species	1.14	1.26	1.12	1.12	1.16	1.33	1.10	1.12	1.14	1.15
cluster coefficient	0.08	0.08	0.08	0.10	0.10	0.50	0.08	0.13	0.08	0.11
cluster coefficient HL	0.38	0.28	0.49	0.45	0.45	0.50	0.48	0.45	0.39	0.37
cluster coefficient LL	0.13	0.24	0.19	0.29	0.44	0.60	0.25	0.26	0.14	0.25
weighted cluster coefficient HL	0.96	0.74	0.66	0.23	0.68	0.48	0.68	0.35	0.46	0.03
weighted cluster coefficient LL	0.03	0.07	0.02	0.03	0.06	0.44	0.02	0.06	0.03	0.02
C score HL	0.81	0.37	0.81	0.28	0.42	0.50	0.67	0.53	0.68	0.09
C score LL	0.86	0.70	0.73	0.49	0.60	0.38	0.59	0.48	0.73	0.51

interaction strength asymmetry	1.45	-0.16	-10.58	1.27	-1.56	-0.37	-6.53	0.80	-4.21	-4.73
specialisation asymmetry	-0.24	-0.07	-0.01	0.06	0.07	0.04	-0.13	-0.01	-0.05	0.14
linkage density	5.31	4.25	2.47	2.27	2.21	2.07	2.69	2.33	2.25	1.84
Shannon diversity	4.36	3.85	2.77	2.50	2.28	1.81	2.75	2.37	2.79	2.18
interaction evenness	0.45	0.50	0.35	0.42	0.35	0.50	0.34	0.42	0.37	0.40
Alatalo interaction evenness	0.49	0.63	0.61	0.71	0.58	0.64	0.43	0.54	0.47	0.60
H2	0.76	0.76	0.71	0.73	0.79	0.71	0.69	0.71	0.83	0.86
mean number of shared partners HL	0.20	0.66	0.23	0.74	0.59	1.13	0.33	0.49	0.33	0.97
mean number of shared partners LL	5.41	1.65	1.38	0.78	1.11	1.20	1.48	0.96	1.26	0.72
niche overlap HL	0.16	0.48	0.18	0.53	0.53	0.33	0.29	0.38	0.27	0.62
niche overlap LL	0.07	0.07	0.20	0.25	0.16	0.22	0.30	0.33	0.20	0.26
togetherness HL	0.05	0.18	0.06	0.25	0.19	0.21	0.10	0.17	0.09	0.33
togetherness LL	0.02	0.03	0.02	0.05	0.06	0.28	0.01	0.06	0.02	0.08
V ratio HL	0.54	1.13	0.80	1.84	1.50	1.22	0.63	1.21	1.10	2.12
V ratio LL	116.38	79.53	26.46	21.45	25.15	1.00	65.56	9.87	32.12	15.58
discrepancy HL	988	97	195	14	34	3	173	20	105	9
discrepancy LL	927	71	167	13	22	4	173	20	93	7
extinction slope HL	1.04	1.20	1.07	1.19	1.11	2.46	1.08	1.25	1.05	1.55
extinction slope LL	1.00	1.00	1.00	1.00	1.00	3.09	1.00	1.00	1.00	1.00
robustness HL	0.51	0.55	0.52	0.54	0.53	0.70	0.52	0.55	0.52	0.60
robustness LL	0.98	0.80	0.85	0.62	0.75	0.76	0.84	0.70	0.87	0.65
functional complementarity HL	18.12	16.94	8.18	7.95	1.32	1.00	3.67	3.60	5.28	4.81
functional complementarity LL	15.55	15.18	7.15	6.99	1.45	1.15	3.83	3.73	5.35	4.95
partner diversity HL	0.57	0.54	0.81	0.76	0.39	0.33	0.75	0.70	0.53	0.50
partner diversity LL	1.96	1.56	0.78	0.63	0.92	0.92	0.91	0.68	0.76	0.42
generality HL	2.05	1.98	2.52	2.41	1.65	1.55	2.50	2.33	1.93	1.85
vulnerability LL	8.58	6.52	2.42	2.13	2.76	2.59	2.88	2.34	2.56	1.82

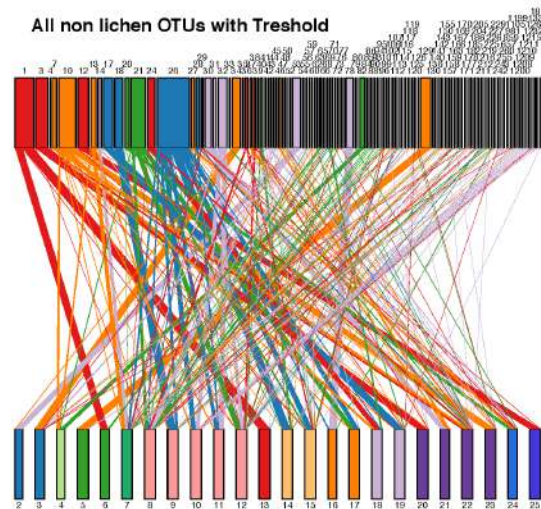
Supplementary table 4. Expanded representation of network statistics for unipartite and bipartite sample networks, as well as for bipartite host species vs. OTUs.

	All OTUs						Main Lichenicole MOTUs only									
	Complete			Threshold filtered			Complete			Threshold filtered						
	d'	d	dmin	dmax	d'	d	dmin	dmax	d'	d	dmin	dmax	d'	d	dmin	dmax
<i>Acarospora fuscata</i>	0.39	2.54	2.12	3.22	.	2.68	2.02	3..	0.97	2.26	1.52	2.28	.	2.27	1.52	2.27
<i>Aspilidea myrinii</i>	0.97	2.50	1.73	2.53	0.99	2.48	2.03	2.69	0.99	0.83	0.83	2.52	.	0.99	1.00	3.59
<i>Candelariella vitellina</i>	0.00	2.08	2.08	3.22	0.33	2.17	2.03	3.40	0.00	.	.	3.52	.	0.00	1.00	3.59
<i>Lecanora bicincta</i>	0.60	2.20	1.73	2.53	0.29	1.94	1.94	2.51
<i>Lecanora intricata</i>	0.00	2.03	2.03	3.22	0.00	0.96	0.96	3.13	0.00	1.32	1.32	4.15	.	0.00	1.33	4.14
<i>Lecanora polytropa</i>	0.00	1.02	1.02	1.61	0.00	2.59	2.03	1.50	0.00	0.79	0.79	1.19	.	0.00	0.79	1.19
<i>Lecanora swartzii</i>	0.51	2.68	2.12	3.22	0.52	1.44	2.03	3.11	0.00	1.09	1.09	8.23
<i>Lecidea lapicida</i>	0.00	1.50	1.50	2.53	0.00	1.44	1.44	2.44	0.00	1.06	1.06	3.56	.	0.00	1.08	3.56
<i>Psorinia conglomerata</i>	0.00	1.66	1.66	2.53	0.00	1.64	1.64	2.53	0.00	1.23	1.23	2.60	.	0.00	1.37	2.64
<i>Rhizocarpon geographicum</i>	0.33	1.99	1.73	2.53	0.00	2.00	2.00	2.47	0.00	1.41	1.41	1.88	.	0.00	1.46	1.88
<i>Tephromela atra</i>	0.48	1.53	1.25	1.83	0.46	1.52	1.32	1.75	0.00	1.36	1.36	1.83	.	0.00	1.36	1.82
<i>Umbilicaria cylindrica</i>	0.78	2.97	2.12	3.22	0.85	3.08	2.03	3.26	0.02	1.71	1.52	9.20
<i>Varicellaria lactea</i>	0.94	3.15	2.12	3.22	0.99	3.12	2.03	3.13	0.95	1.99	1.52	2.02	.	1.00	2.01	2.01
<i>Muellerella pygmaea</i> 21									0.00	0.93	0.93	1.71	.	0.00	0.94	1.70
<i>Sagediopsis fissurisedens</i> 34									0.96	2.23	1.19	2.27	.	0.96	2.23	2.27
<i>Cercidospora epipolytropa</i> 18									0.00	1.13	1.13	2.28	.	0.00	1.19	2.30
<i>Lichenocodium lecanorae</i> 32									0.00	1.14	1.14	2.21	.	0.00	1.15	2.20
<i>Endococcus macrosporus</i> 17									0.64	1.79	1.19	2.13	.	0.73	1.88	2.14
<i>Muellerella atricola</i> 46									0.24	1.81	1.19	3.75	.	0.25	1.82	3.75
<i>Skyttea tephromelarum</i> 33									0.17	1.77	1.19	4.63	.	0.18	1.82	4.63
<i>Taeniolella atricerebrina</i> 10									0.00	0.74	0.74	1.52	.	0.00	0.75	1.52
<i>Stigmatidium eucline</i> 12									0.93	1.96	1.19	2.01	.	1.00	2.01	2.01

Supplementary table 5. Standardized and raw specialization indices calculated for each lichen host species (A) in the networks, as well as for those lichenicolous fungi identifiable as the main MOTUs at sample level (B). The values of d' range between 0, meaning no specialization and 1 meaning perfect specialization.

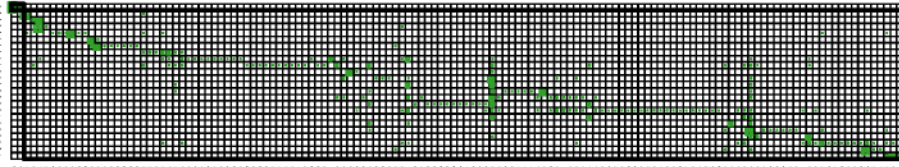


Supplementary Figure 17. Bipartite network at sample level including all non-lichen OTUs.

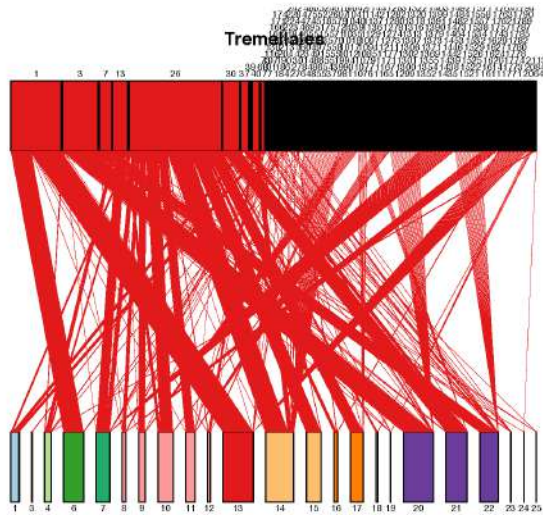


Supplementary Figure 18. Bipartite network at sample level of non-lichen OTUs above a sample specific threshold.

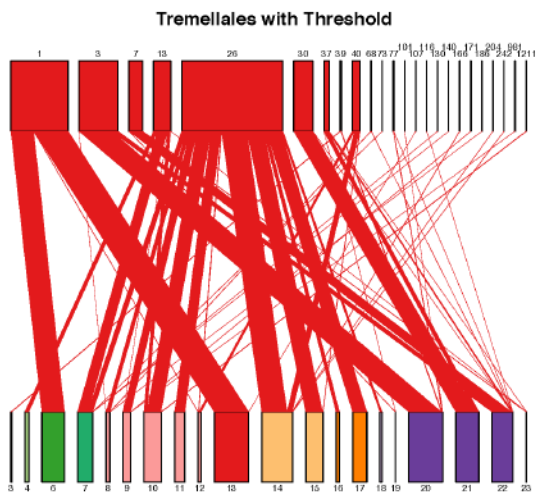
All non lichen OTUs with Treshold



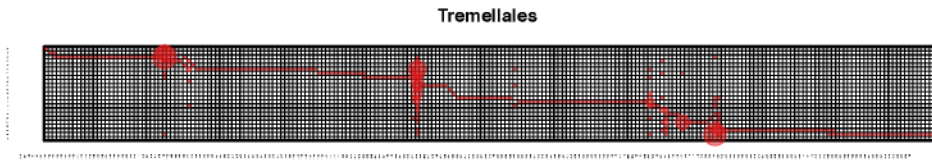
Supplementary Figure 19. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level of non-lichen OTUs above a sample specific threshold.



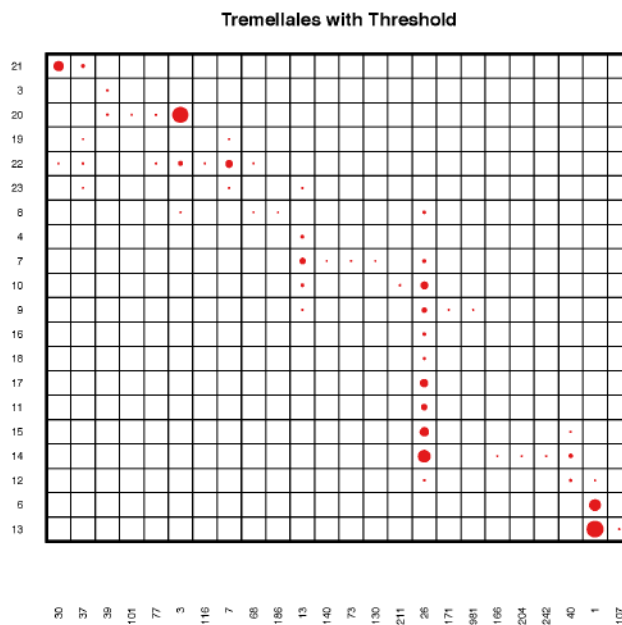
Supplementary Figure 20. Bipartite network at sample level using only OTUs identified as Tremellales.



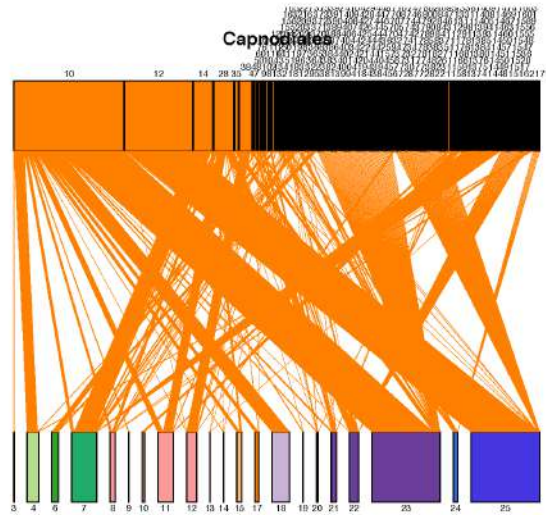
Supplementary Figure 21. Bipartite network at species level using only those OTUs identified as Tremellales found above a sample specific threshold.



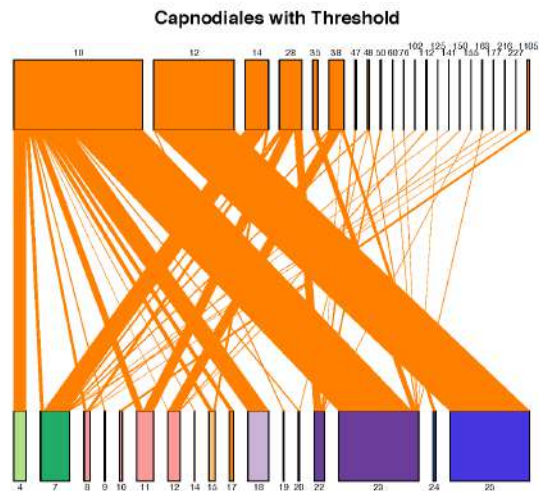
Supplementary Figure 22. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Tremellales.



Supplementary Figure 23. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Tremellales sequence above a sample specific threshold.

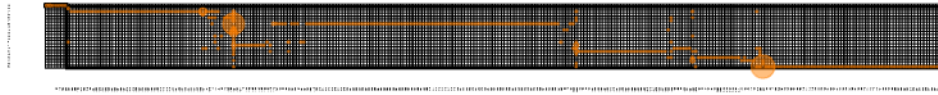


Supplementary Figure 24. Bipartite network at sample level using only OTUs identified as Capnodiales.



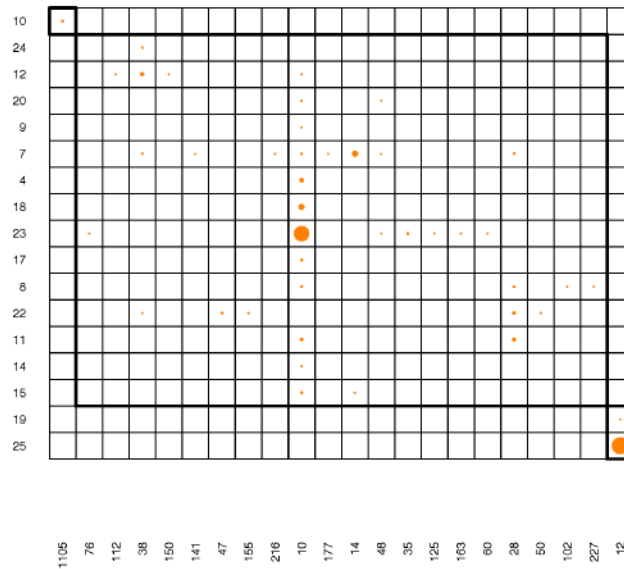
Supplementary Figure 25. Bipartite network at sample level using only those OTUs identified as Capnodiales above a sample specific threshold.

Capnodiales

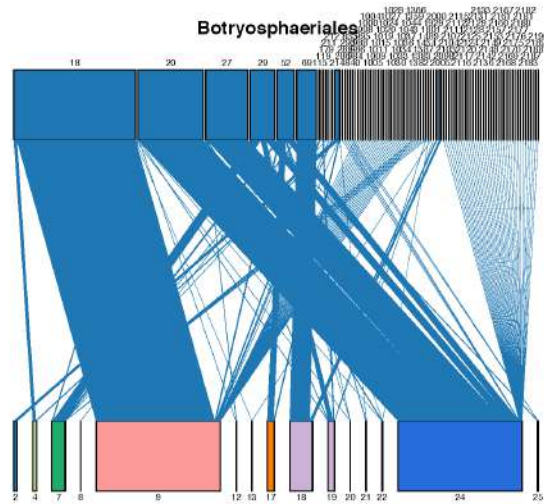


Supplementary Figure 26. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Capnodiales.

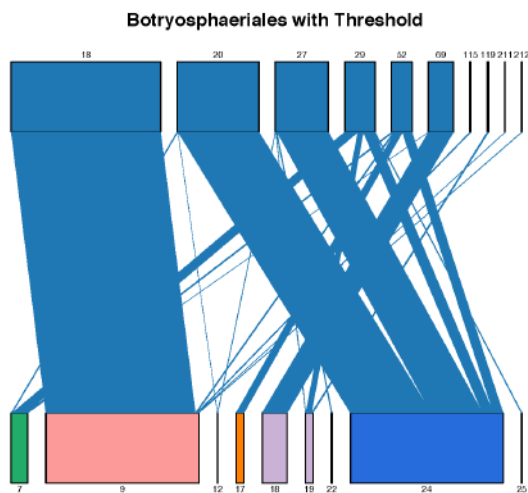
Capnodiales with Threshold



Supplementary Figure 27. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Capnodiales sequence above a sample specific threshold.

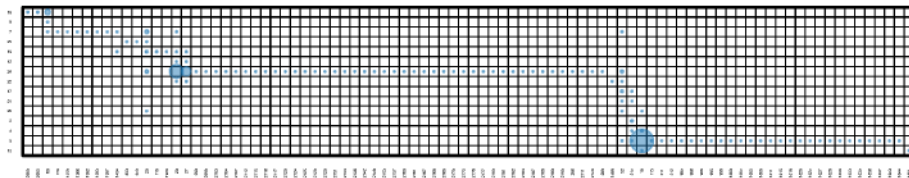


Supplementary Figure 28. Bipartite network at sample level using only OTUs identified as Botryosphaerales.

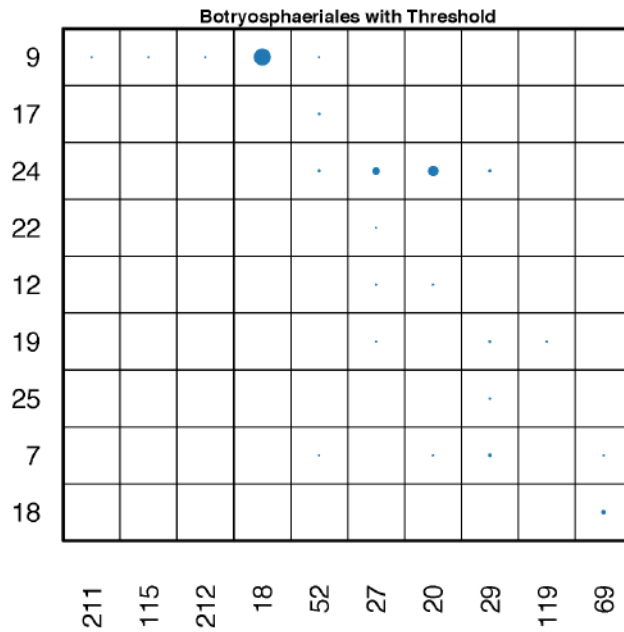


Supplementary Figure 29. Bipartite network at sample level using only those OTUs identified as Botryosphaerales above a sample specific threshold.

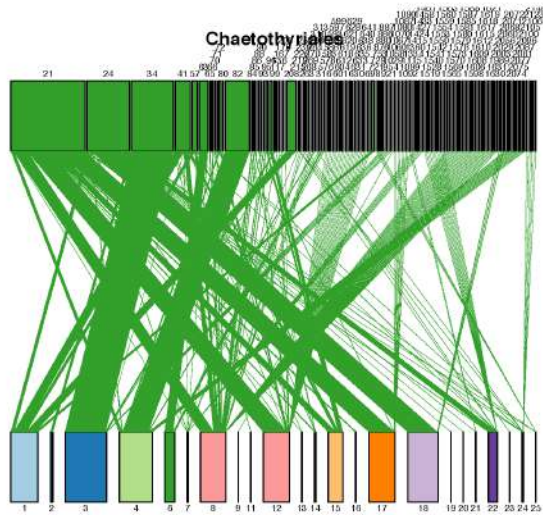
Botryosphaerales



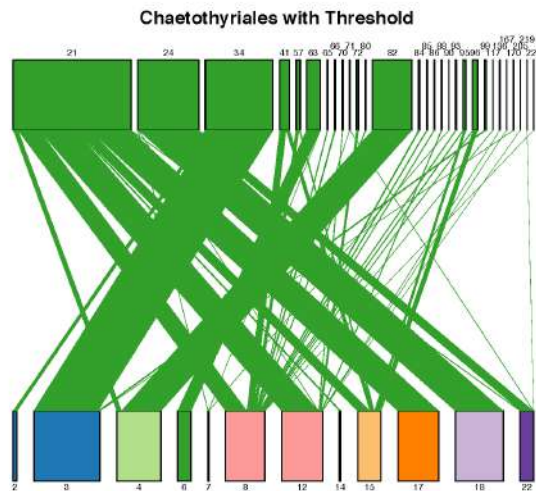
Supplementary Figure 30. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Botryosphaerales.



Supplementary Figure 31. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Botryosphaerales sequence above a sample specific threshold.

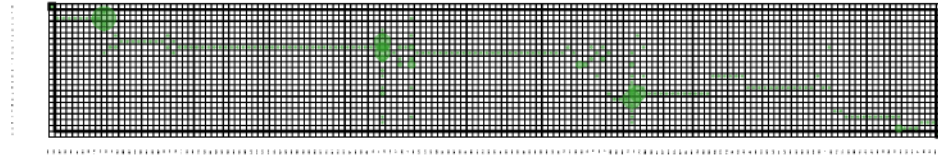


Supplementary Figure 32. Bipartite network at sample level using only OTUs identified as Chaetothyriales.

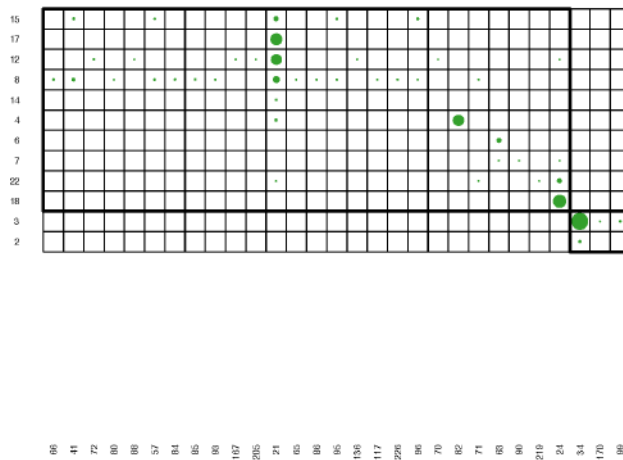


Supplementary Figure 33. Bipartite network at sample level using only those OTUs identified as Chaetothyriales found above a sample specific threshold.

Chaetothyriales

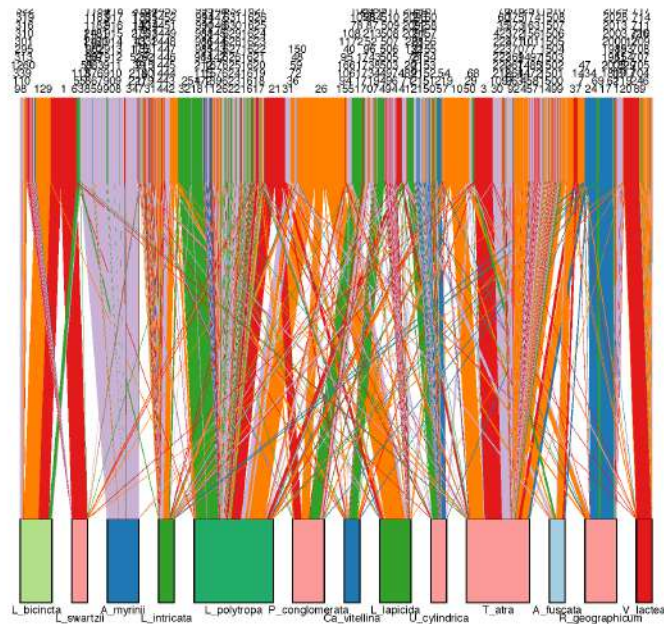


Supplementary Figure 34. Matrix representation showing nestedness and compartmentalization of the bipartite network at sample level using all OTUs identified as Chaetothyriales.



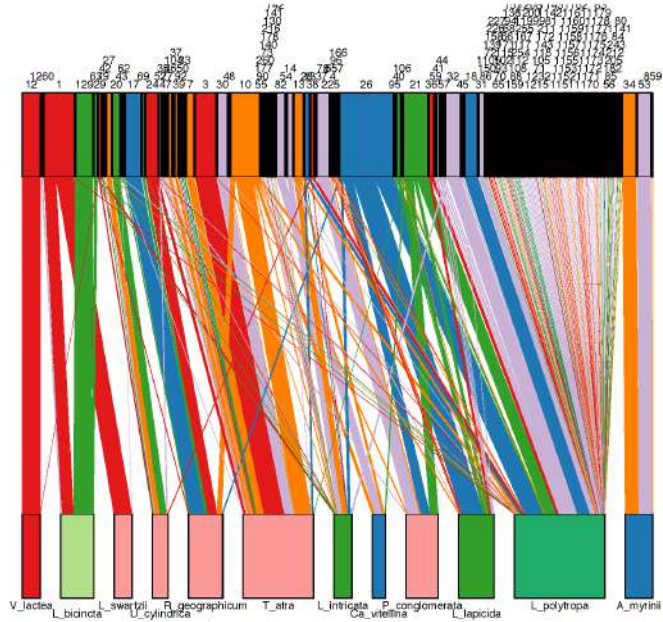
Supplementary Figure 35. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Chaetothyriales sequence above a sample specific threshold.

All non lichen OTUs



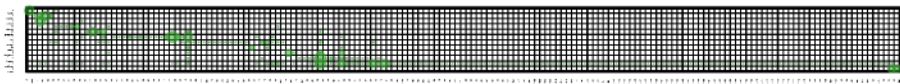
Supplementary Figure 36. Bipartite network at species level including all non-lichen OTUs.

All non lichen OTUs with Treshold

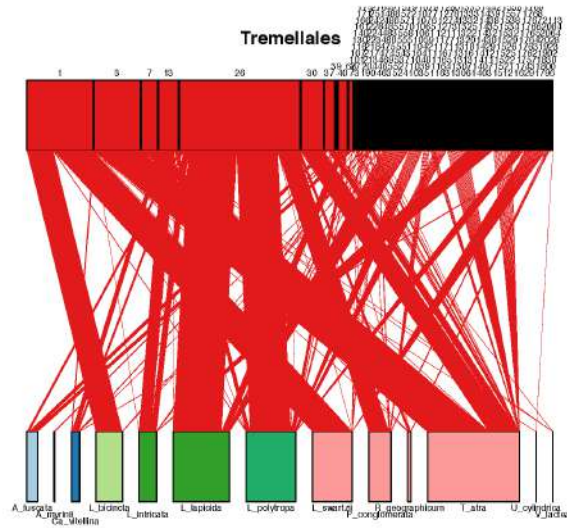


Supplementary Figure 37. Bipartite network at species level of non-lichen OTUs above a sample specific threshold.

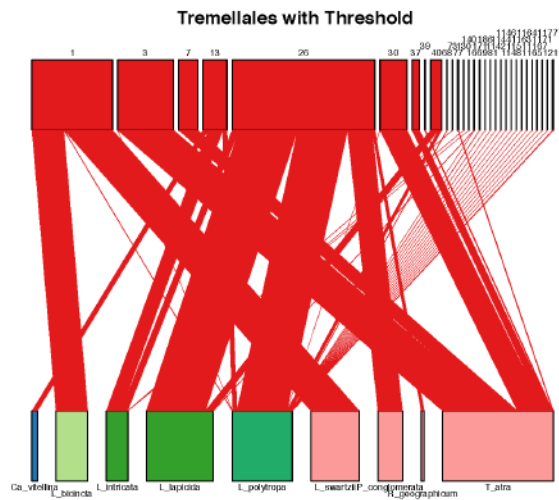
All non lichen OTUs with Treshold



Supplementary Figure 38. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level of non-lichen OTUs above a sample specific threshold.

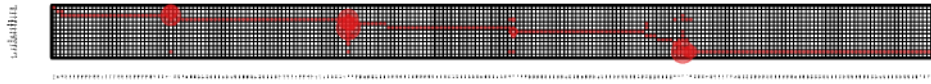


Supplementary Figure 39. Bipartite network at species level using only OTUs identified as Tremellales.



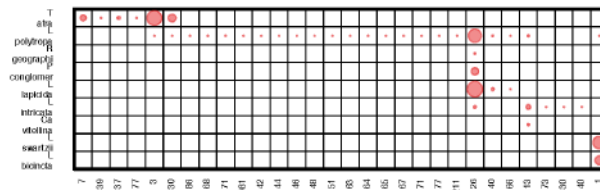
Supplementary Figure 40. Bipartite network at species level using only those OTUs identified as Tremellales found above a sample specific threshold.

Tremellales

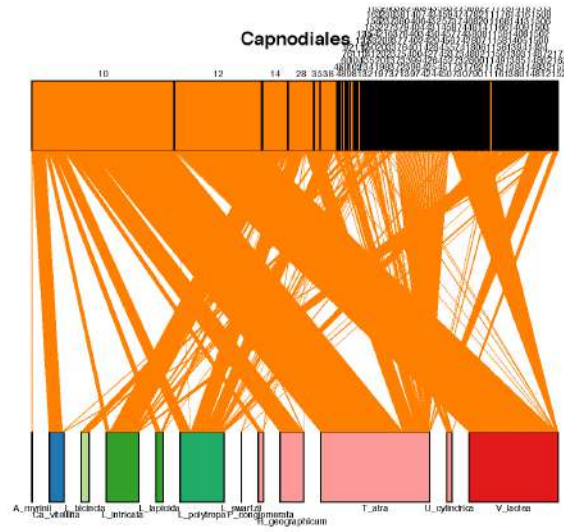


Supplementary Figure 41. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Tremellales.

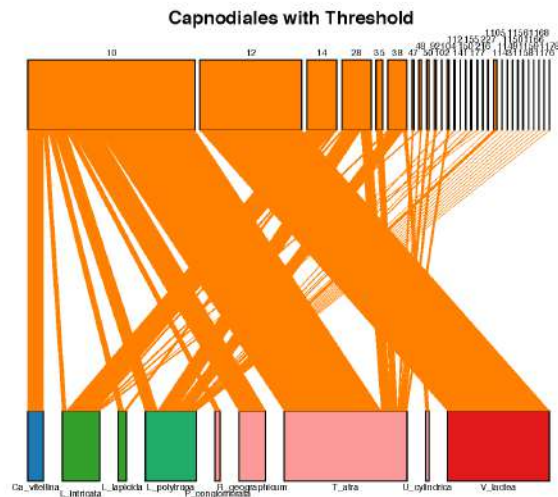
Tremellales with Threshold



Supplementary Figure 42. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Tremellales sequence above a sample specific threshold.

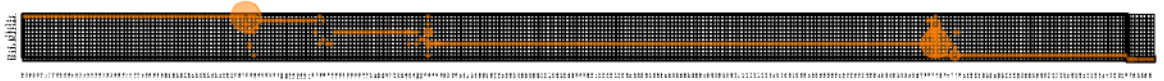


Supplementary Figure 43. Bipartite network at species level using only OTUs identified as Capnodiales.



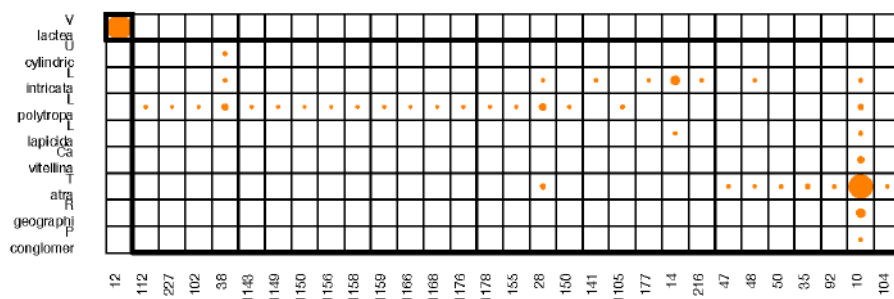
Supplementary Figure 44. Bipartite network at species level using only those OTUs identified as Capnodiales found above a sample specific threshold.

Capnodiales

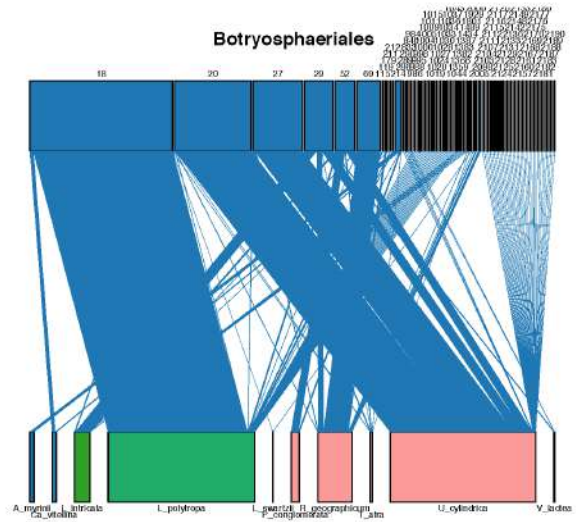


Supplementary Figure 45. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Capnodiales.

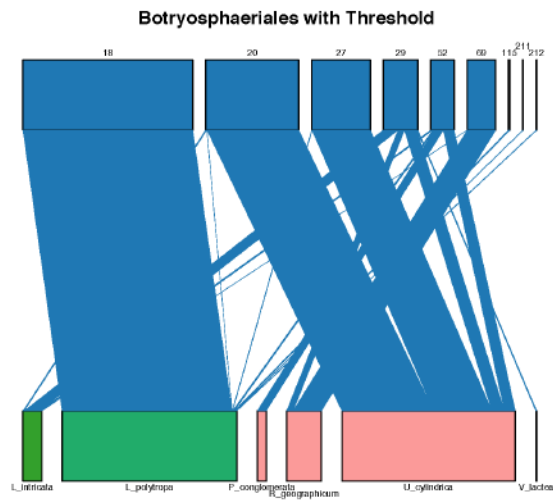
Capnodiales with Threshold



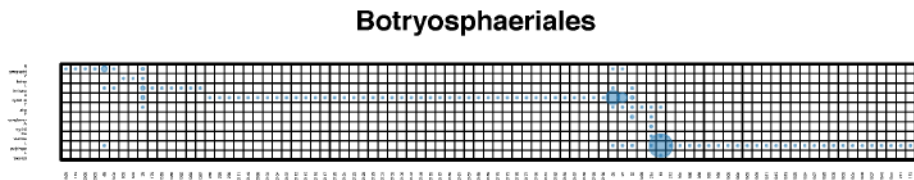
Supplementary Figure 46. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Capnodiales sequence above a sample specific threshold.



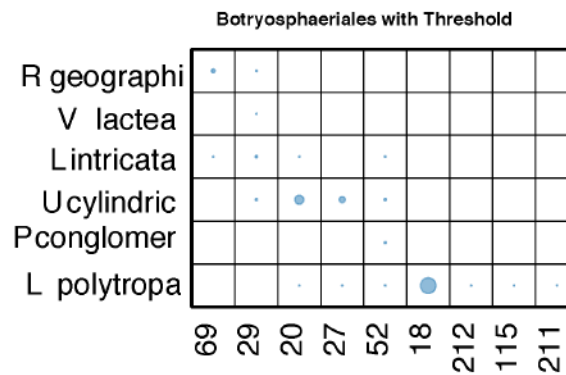
Supplementary Figure 47. Bipartite network at species level using only OTUs identified as Botryosphaerales.



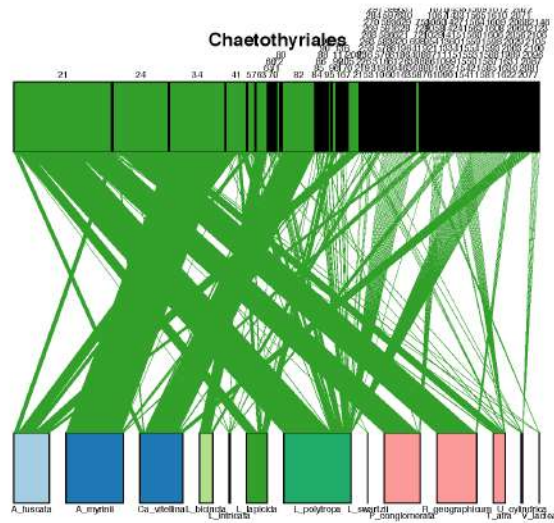
Supplementary Figure 48. Bipartite network at species level using only those OTUs identified as Botryosphaerales found above a sample specific threshold.



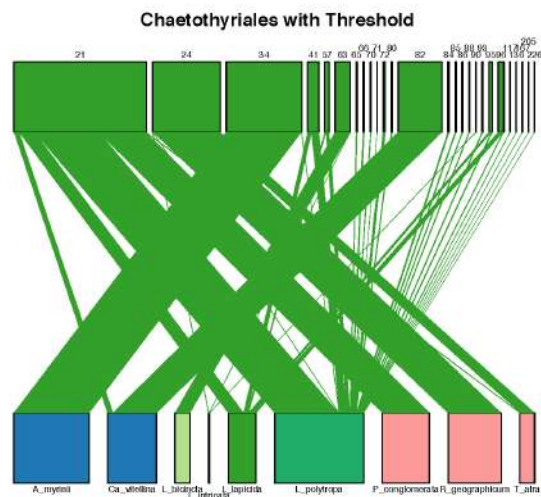
Supplementary Figure 49. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Botryosphaerales.



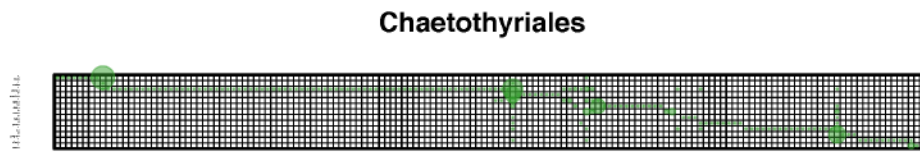
Supplementary Figure 50. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Botryosphaerales sequence above a sample specific threshold.



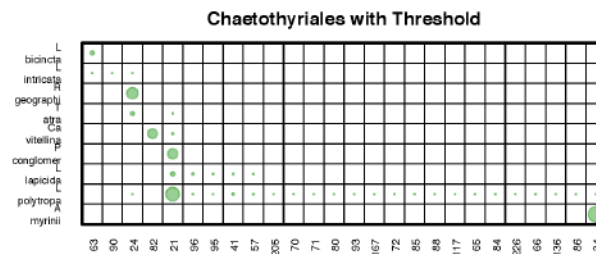
Supplementary Figure 51. Bipartite network at species level using only OTUs identified as Chaetothyriales.



Supplementary Figure 52. Bipartite network at species level using only those OTUs identified as Chaetothyriales found above a sample specific threshold.



Supplementary Figure 53. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using all OTUs identified as Chaetothyrales.














Supplementary Figure 54. Matrix representation showing nestedness and compartmentalization of the bipartite network at species level using only those OTUs identified as Chaetothyrales sequence above a sample specific threshold.

Supplementary Figure 55. The following pages contain the output of the quality assessment of the raw data obtained using FastQC.

FastQC Report

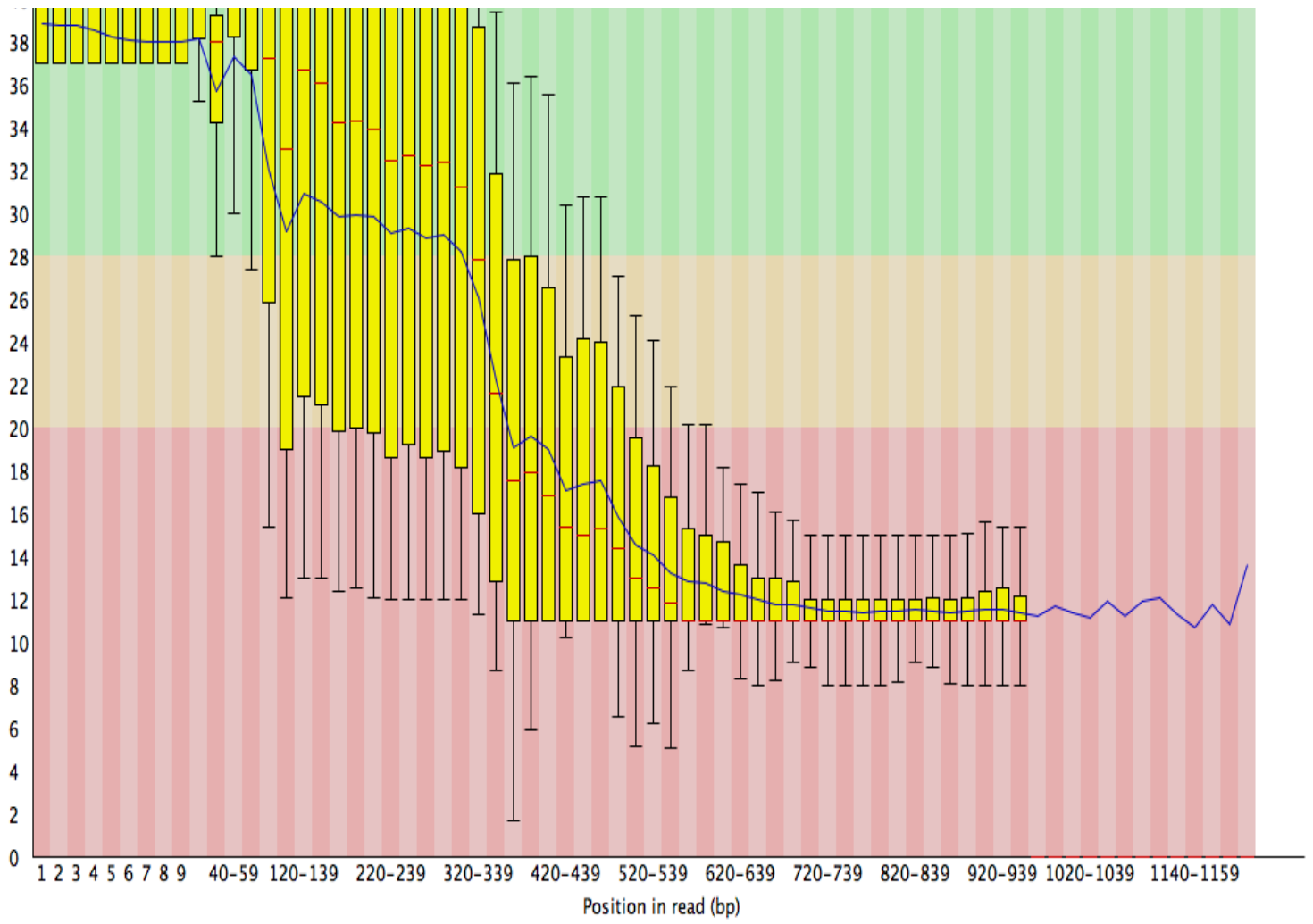
Summary

-  [Basic Statistics](#)
-  [Per base sequence quality](#)
-  [Per sequence quality scores](#)
-  [Per base sequence content](#)
-  [Per sequence GC content](#)
-  [Per base N content](#)
-  [Sequence Length Distribution](#)
-  [Sequence Duplication Levels](#)
-  [Overrepresented sequences](#)
-  [Adapter Content](#)
-  [Kmer Content](#)

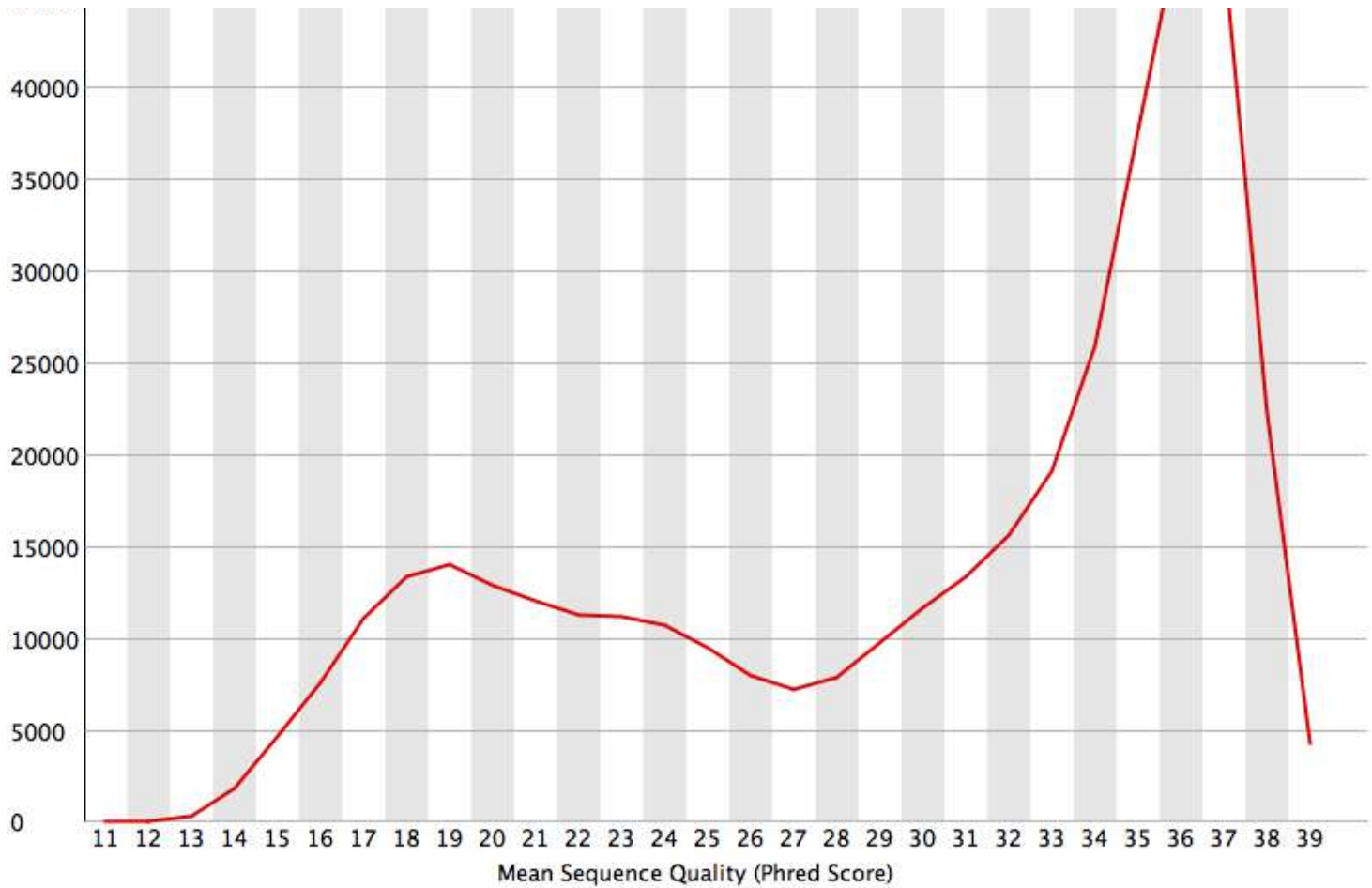
Basic Statistics

Measure	Value
Filename	merged.fastq
File type	Conventional base calls
Encoding	Sanger / Illumina 1.9
Total Sequences	399593
Sequences flagged as poor quality	0
Sequence length	60–1200
%GC	52

Per base sequence quality

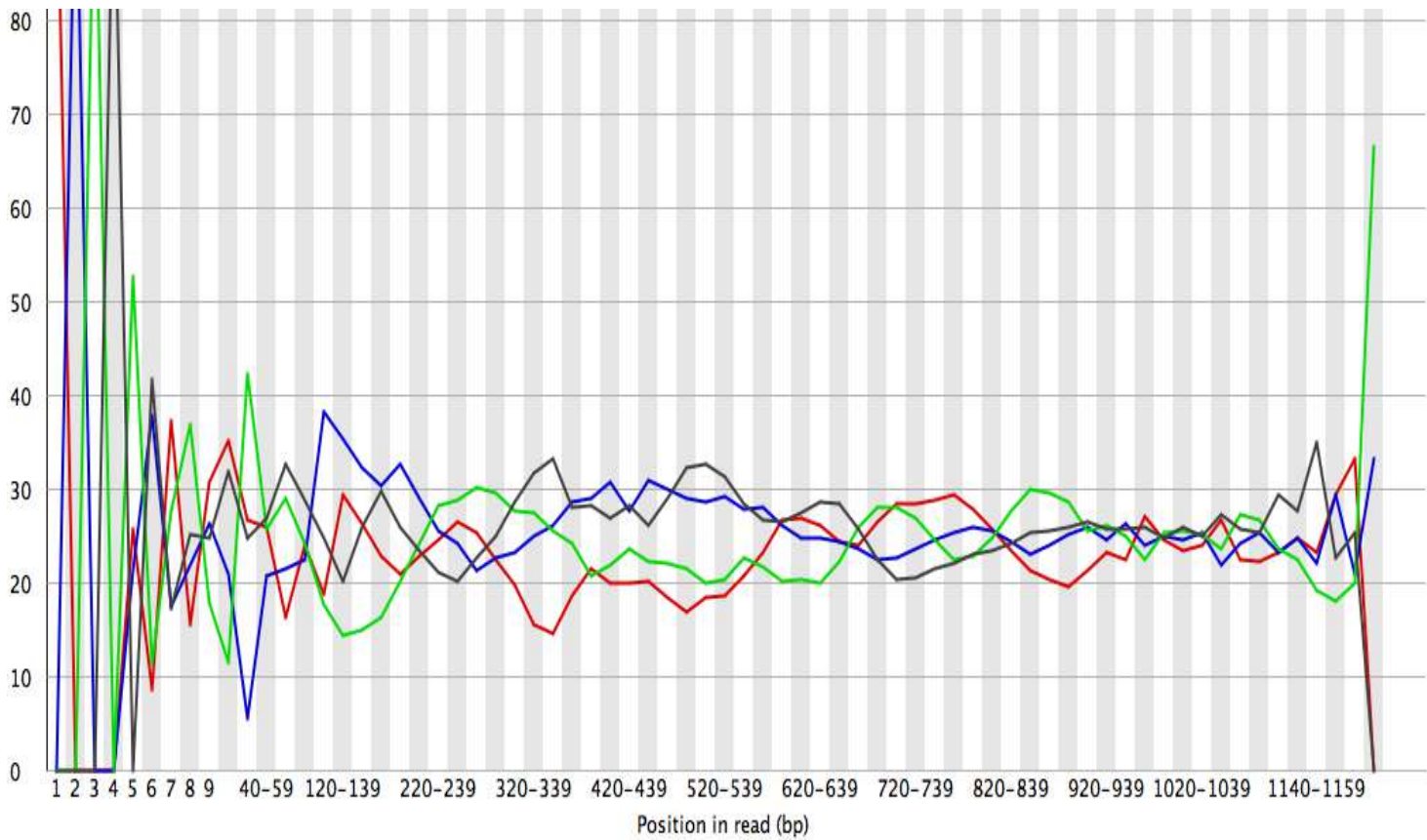


 **Per sequence quality scores**

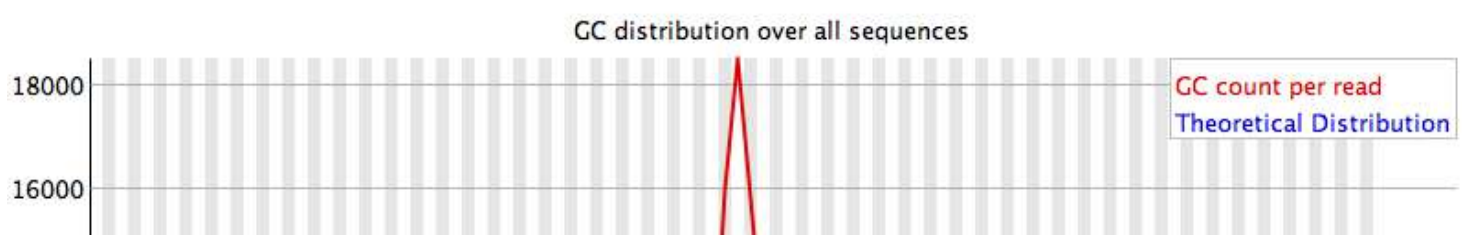


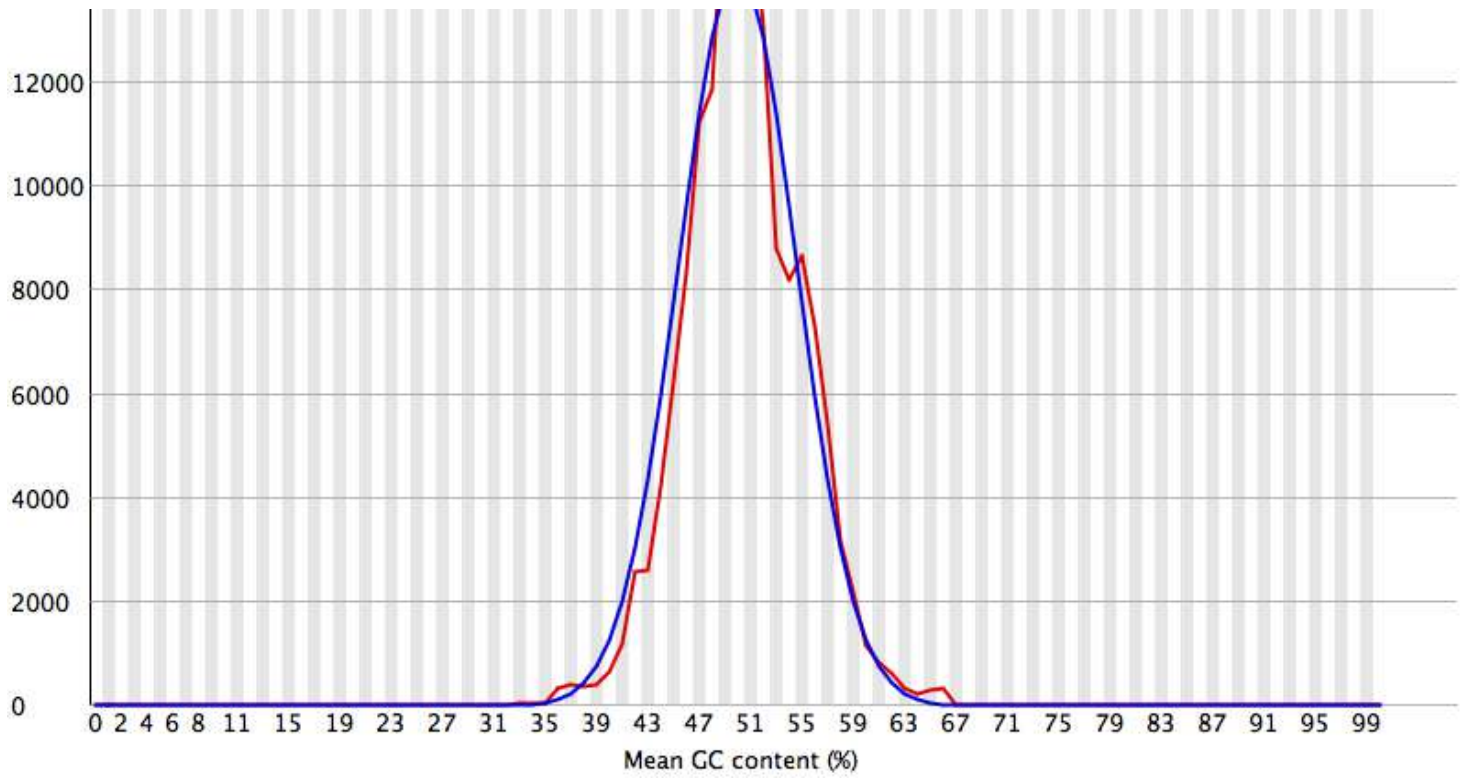
✖ Per base sequence content



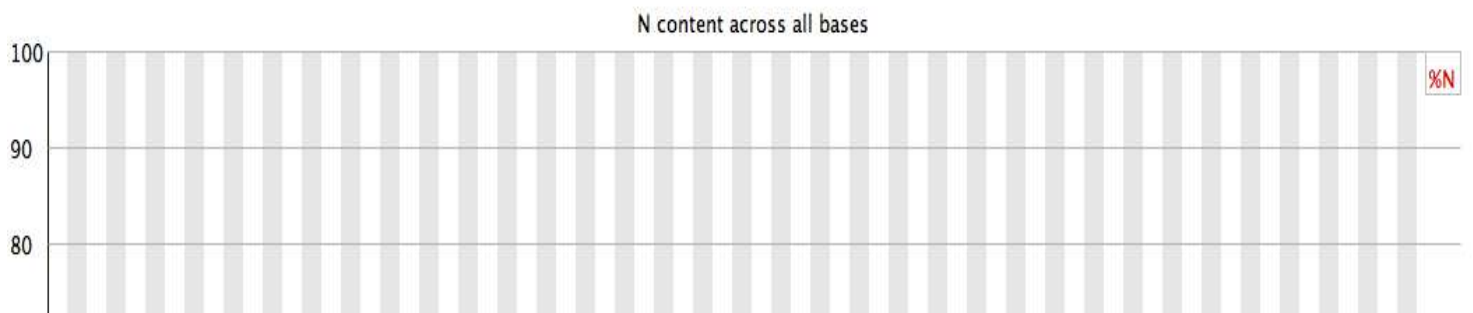


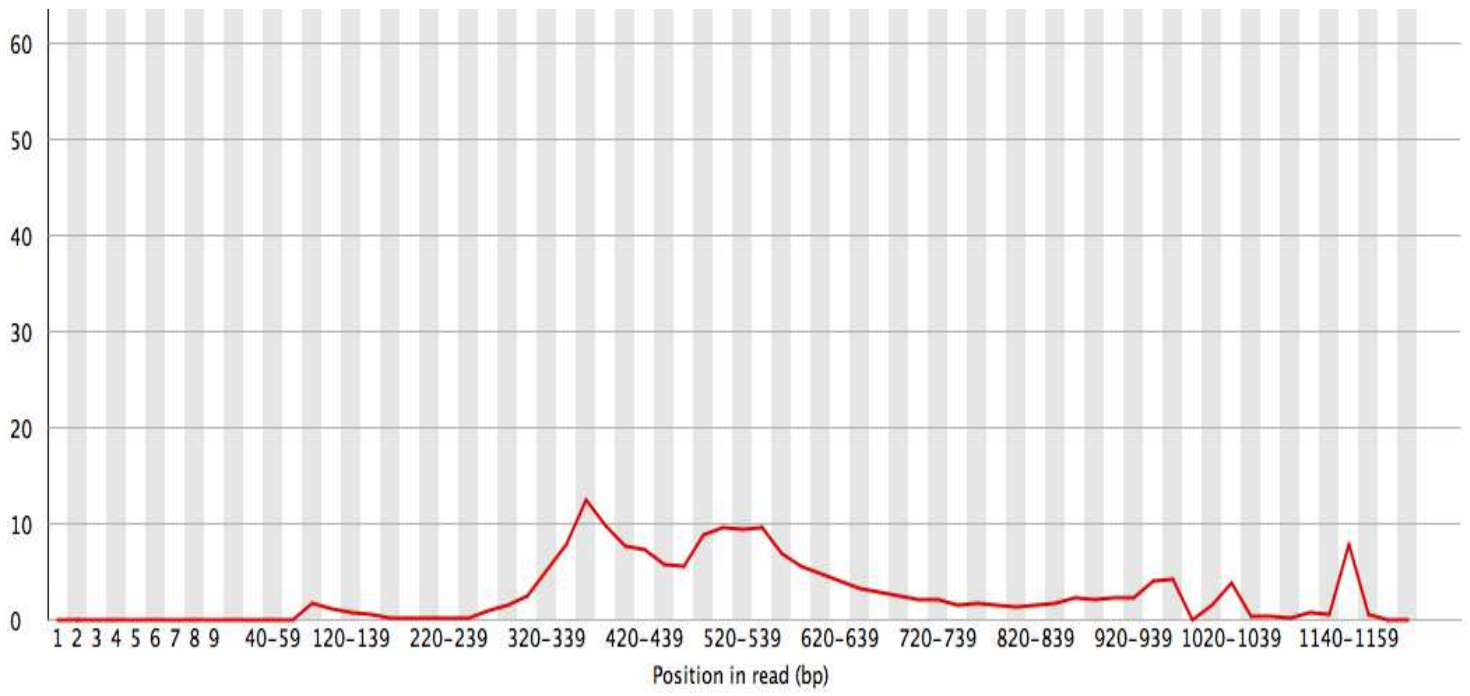
 **Per sequence GC content**



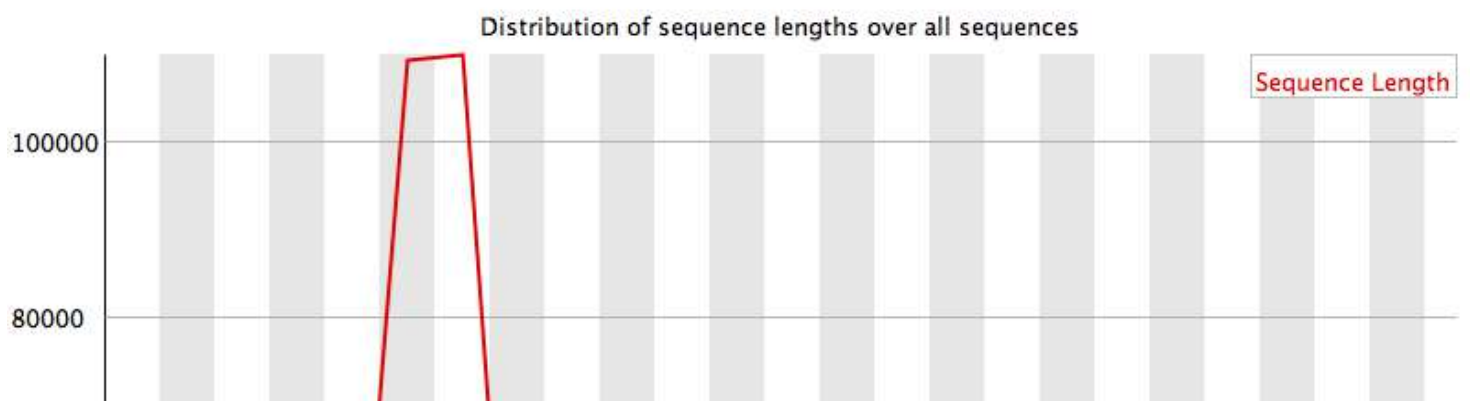


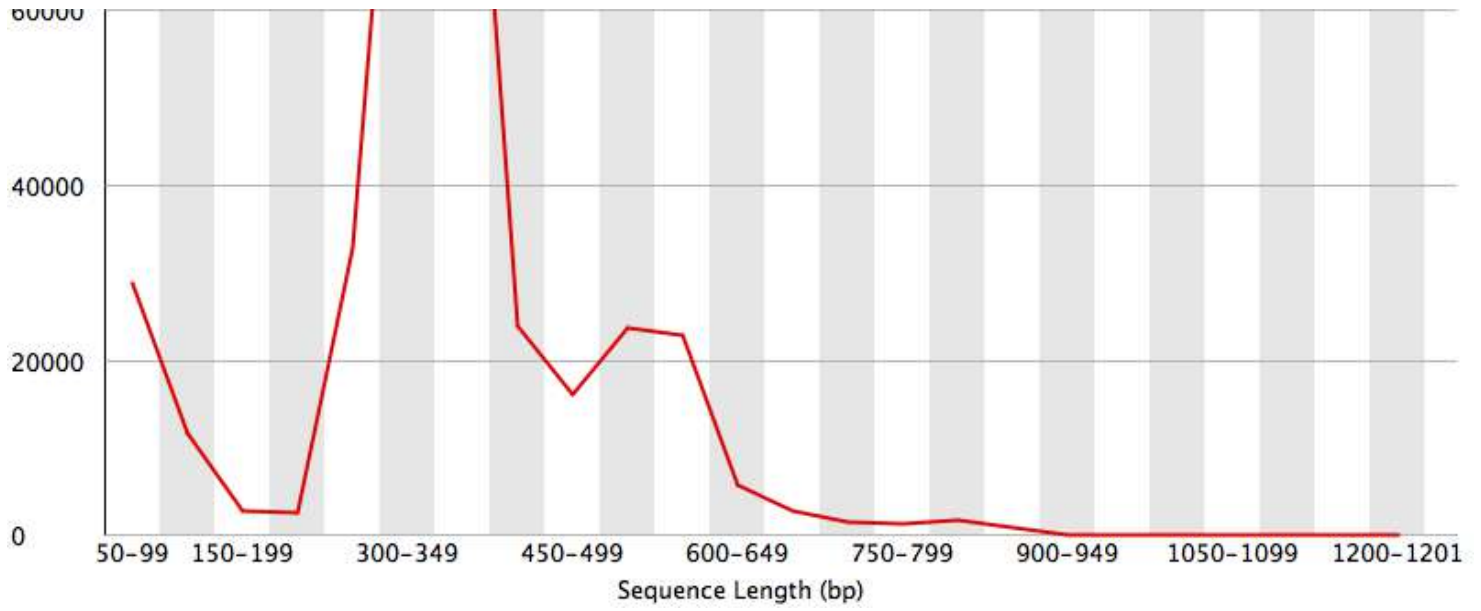
 **Per base N content**



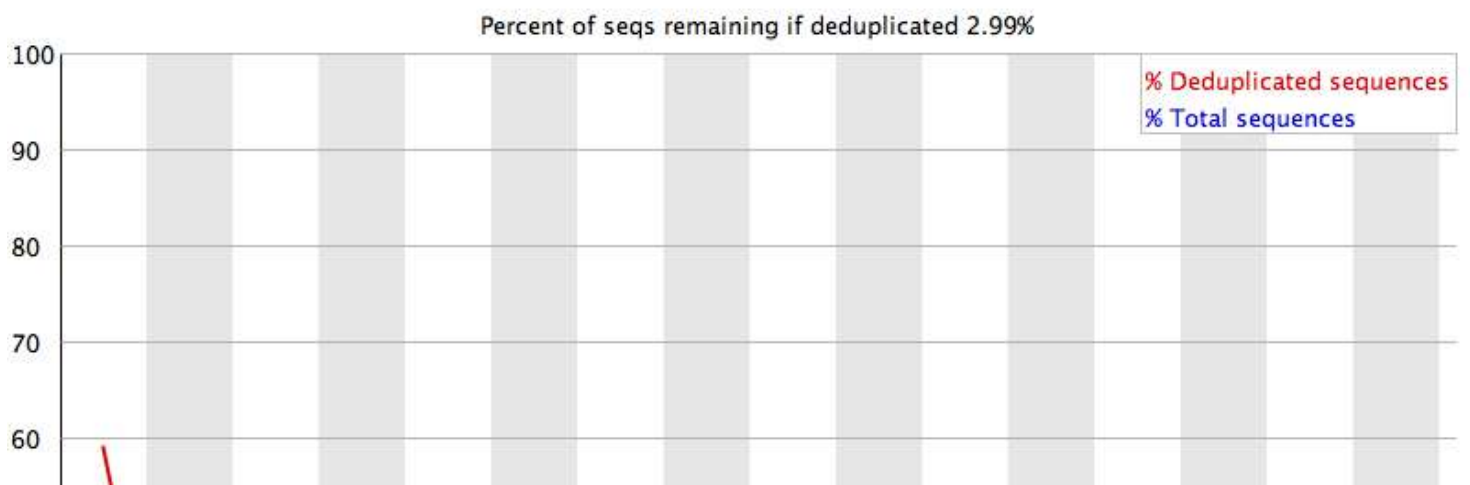


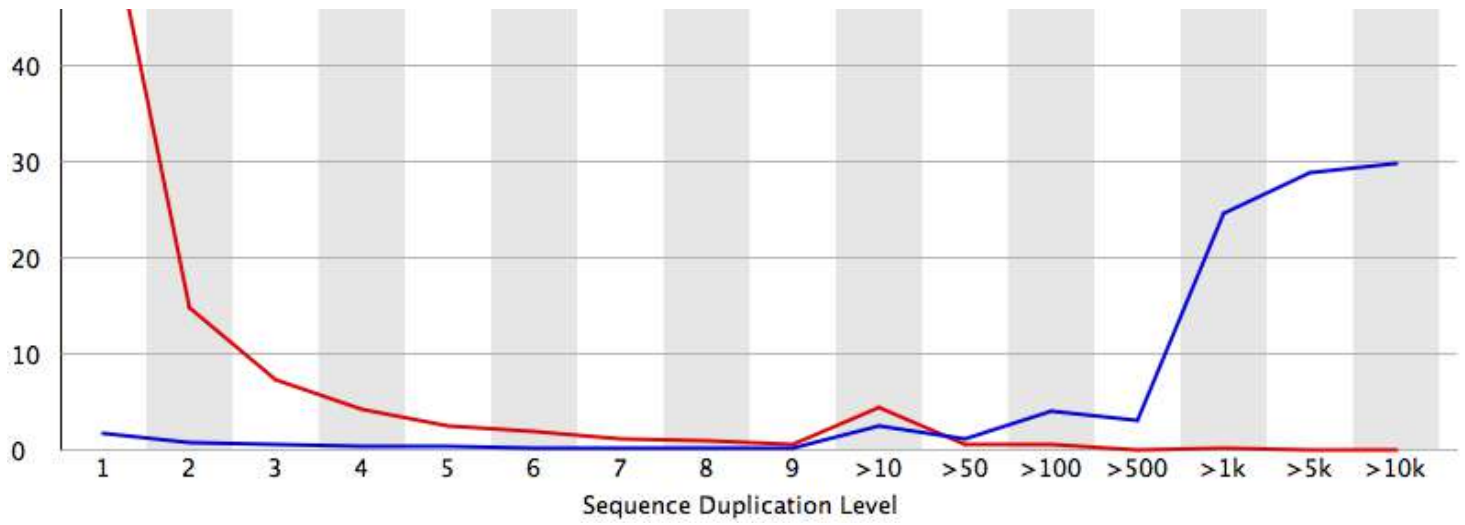
Sequence Length Distribution





✖ Sequence Duplication Levels





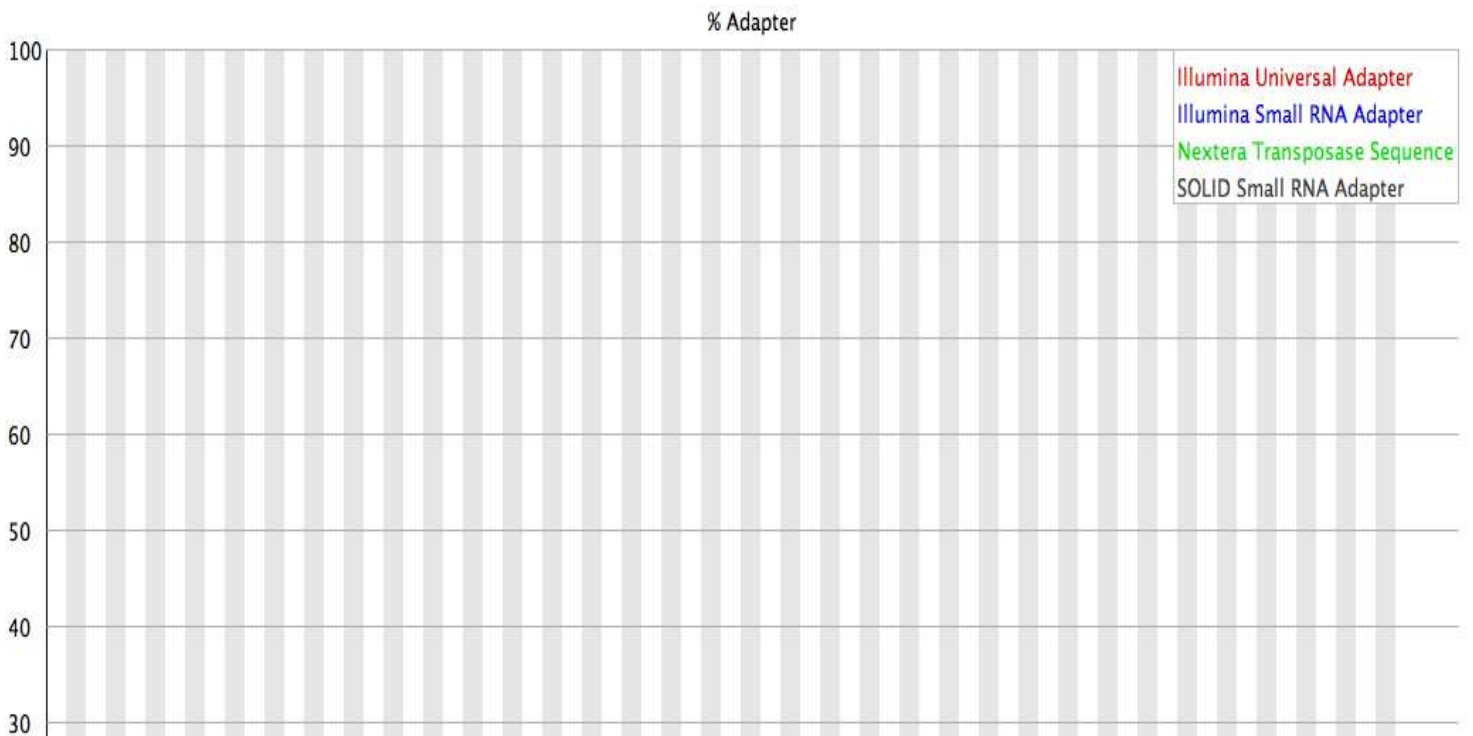
Overrepresented sequences

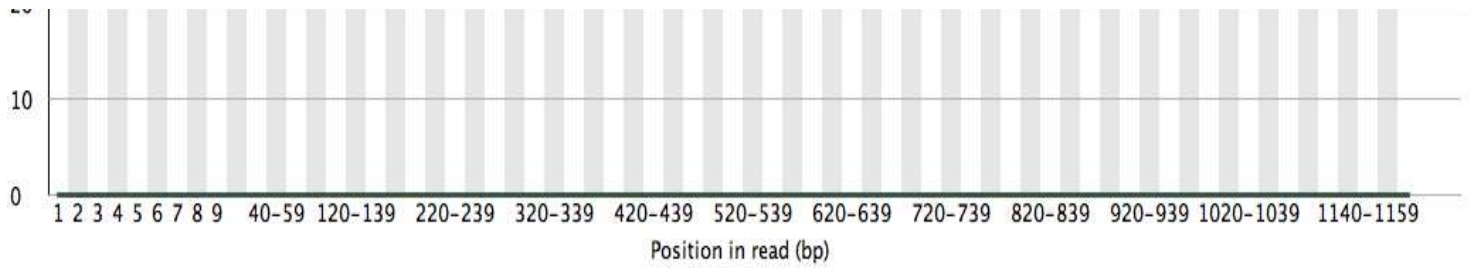
Sequence	Count	Percentage	Possible Source
TCAGACTACTATGTCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	17507	4.3812078790169	No Hit
TCAGAGACTATACTCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	17043	4.265089728799053	No Hit
TCAGTGATACGTCTCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	16360	4.094165813715456	No Hit
TCAGACTGTACAGTCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	15458	3.8684361337661075	No Hit
TCAGACGAGTGCCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	11356	2.8418916247281607	No Hit
TCAGAGCGTCTCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	10826	2.709256668660362	No Hit
TCAGCGTAGACTAGCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	10479	2.6224183106310672	No Hit
TCAGTCTCTATGCGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	10473	2.620916782826526	No Hit
TCAGCGAGAGATACCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	10331	2.5853806247857194	No Hit
TCAGCATAGTAGTGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	9202	2.3028431428978986	No Hit
TCAGATATCGCGAGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	8945	2.2385277019367207	No Hit
TCAGACATACGCGTCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	7974	1.9955304522351491	No Hit
TCAGCGTAGACTAGCTTGGTCATTTAGAGGAAGTAAAGTCGATGAAGAA	7653	1.9151987146921994	No Hit
TCAGACTGTACAGTCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	7425	1.8581406581196367	No Hit
TCAGCTCGCGTGTCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	7234	1.810342023008411	No Hit
TCAGAGACTATACTCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	7224	1.8078394766675092	No Hit
TCAGTGTACTACTCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	7054	1.7652961888721774	No Hit
TCAGACTACTATGTCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	6825	1.7079878776655246	No Hit
TCAGACGACTACAGCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	6567	1.6434221820702564	No Hit
TCAGCGAGAGATACCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	6244	1.5625899352591262	No Hit
TCAGATCAGACACGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	5904	1.4775033596684628	No Hit
TCAGACGCTCGACACTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	5760	1.4414666923594757	No Hit
TCAGCGTGTCTCTACTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	5599	1.4011756962709556	No Hit
TCAGACGCGAGTATCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	5328	1.3333566904325151	No Hit
TCAGAGCACTGTAGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	5285	1.3225957411666371	No Hit

TCAGACGCTCGACACTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	4550	1.13865858511035	No Hit
TCAGAGCGTCGTCTCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	4537	1.1354052748671775	No Hit
TCAGTGATACGTCTCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	4523	1.1319017099899147	No Hit
TCAGCGTCTAGTACCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	4515	1.1298996729171933	No Hit
TCAGTACTCTCGTGCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	4325	1.0823512924400578	No Hit
TCAGAGTACGTATCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	4275	1.0698385607355485	No Hit
TCAGTCTCTATGCGCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	4273	1.069338051467368	No Hit
TCAGTAGAGACGAGCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	4213	1.0543227734219567	No Hit
TCAGATACGACGTACTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	4179	1.0458141158628904	No Hit
TCAGTACGAGTATGCTTGGTCATTTAGAGGAAGTAAGCATCGATGAAGAA	3972	0.9940114066062218	No Hit
TCAGAGCGTCGTCTCTTGGTCATTTAGAGGAAGTAAGCATCGATGAAGAA	3875	0.9697367070994738	No Hit
TCAGTCACGTACTACTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	3795	0.9497163363722588	No Hit
TCAGTACTCTCGTGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	3544	0.886902423215622	No Hit
TCAGTAGAGACGAGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	3485	0.8721373998043008	No Hit
TCAGACATACGCGTCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	3460	0.8658810339520462	No Hit
TCAGCTCGCGTGTCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	3171	0.7935574446999821	No Hit
TCAGTCTACGTAGCCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	2797	0.6999622115502524	No Hit
TCAGACGACTACAGCTTGGTCATTTAGAGGAAGTAAGCATCGATGAAGAA	2594	0.6491605208299445	No Hit
TCAGTCGTGCTCGCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	2366	0.5921024642573819	No Hit
TCAGTGATACGTCTCTTGGTCATTTAGAGGAAGTAAGCATCGATGAAGAA	2351	0.588348644746029	No Hit
TCAGCGTAGACTAGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	2324	0.581591769625594	No Hit
TCAGACGACTACAGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	2176	0.5445540837802464	No Hit
TCAGACGCGAGTATCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	2165	0.5418012828052544	No Hit
TCAGCGTGTCTCTACTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	2124	0.5315408428075566	No Hit
TCAGTGTACTACTCTTGGTCATTTAGAGGAAGTAAGCATCGATGAAGAA	1917	0.479738133550888	No Hit
TCAGCATAGTAGTGCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	1906	0.476985332575896	No Hit
TCAGTACGAGTATGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	1530	0.38288959015798574	No Hit
TCAGACGCTCGACACTTGGTCATTTAGAGGAAGTAAGCATCGATGAAGAA	1445	0.36161794626031984	No Hit
TCAGAGACGCACTCCTTGGTCATTTAGAGGAAGTAAAAGTCGTAACAAGG	1392	0.34835445065354	No Hit
TCAGTAGAGACGAGCTTGGTCATTTAGAGGAAGTAAGCATCGATGAAGAA	1254	0.31381931114909417	No Hit
TCAGAGTACGTATCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	1234	0.3088142184672905	No Hit
TCAGAGACTATACTCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGTT	990	0.24775208774928487	No Hit
TCAGCGTAGACTAGCTTGGTCATTAGAGGAGTAGCATCGATGAAGAACGC	961	0.24049470336066947	No Hit
TCAGTCTACGTAGCCTTGGTCATTTAGAGGAAGTAAGCATCGATGAAGAA	930	0.2327368097038737	No Hit
TCAGCGTAGACTAGCTTGGTCATTAGAGGAAGTAAGCATCGATGAAGAAC	888	0.22222611507208587	No Hit
TCAGAGACGCACTCCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	882	0.2207245872675447	No Hit
TCAGCGTCTAGTACCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	861	0.21546923995165082	No Hit
TCAGACTACTATGTCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGTT	823	0.2059595638562237	No Hit
TCAGCGAGAGATACCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGTT	741	0.1854386838608284	No Hit
TCAGATACGACGTACTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	673	0.1684213687426957	No Hit
TCAGTCTCTATGCGCTTGGTCATTTAGAGGAAGTAAGCATCGATGAAGAA	668	0.16717009557224474	No Hit
TCAGACTGTACAGTCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGTT	644	0.16116398435408028	No Hit

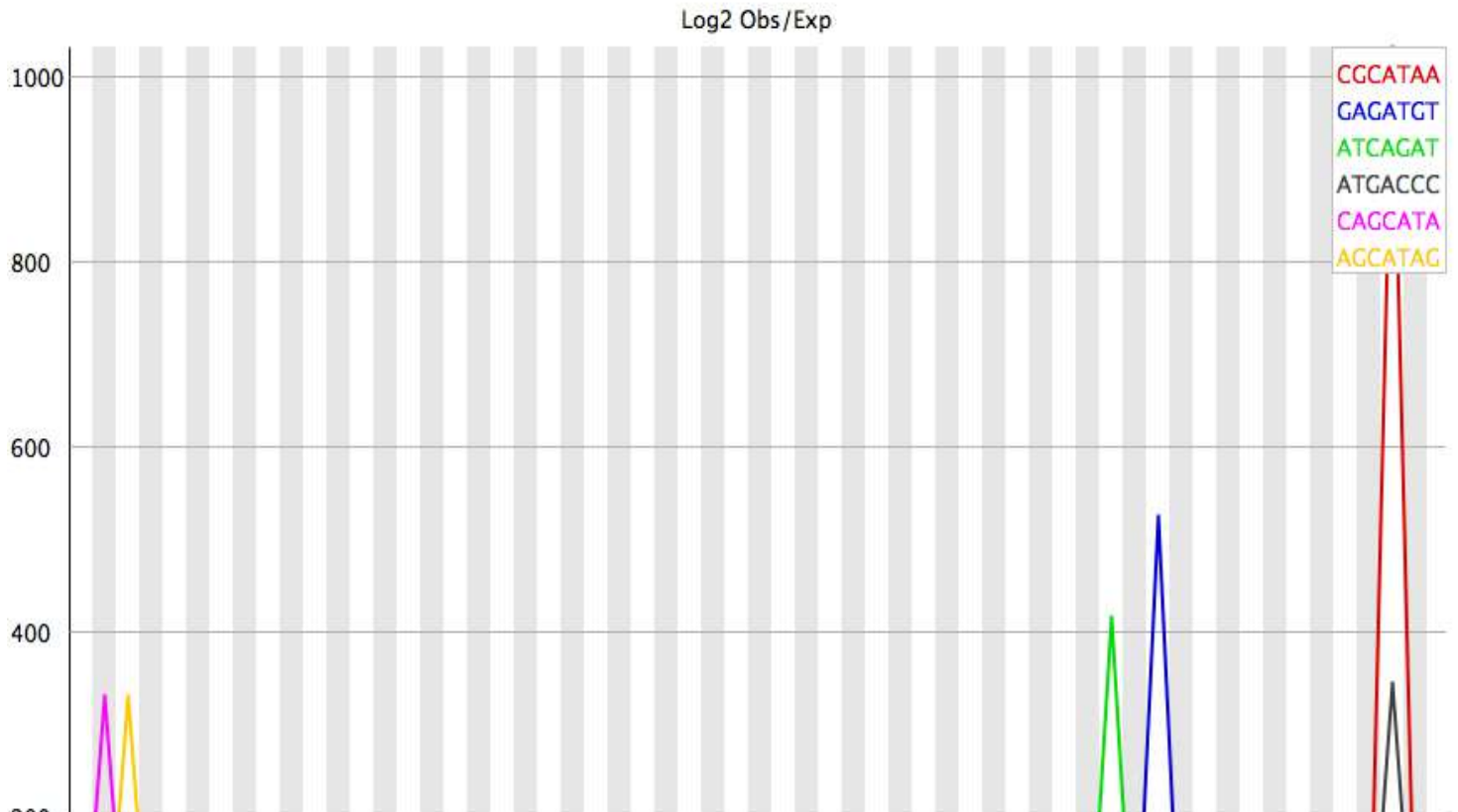
TCAGCGTAGACTAGCTTGGTCATTTAGAGGAGTAGCATCGATGAAGACGCA	620	0.1588594009404589	No Hit
TCAGTCGTCGCTCGCTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	621	0.15540812777000598	No Hit
TCAGACGAGTGCCTTGGTCATTTAGAGGAAGTAAGTCGTAACAAGGTT	575	0.14389641460185737	No Hit
TCAGTGTACTACTCCTTGGTCATTTAGAGGAAGTAAAAAGTCGTAACAAG	568	0.1421446321632261	No Hit
TCAGAGACTATACTCCTTGGTCATTTAGAGGAAGTAAGCATCGATGAAGAA	567	0.1418943775291359	No Hit
TCAGTACTCTCGTGCCTTGGTCATTTAGAGGAAGTAAGTCGTAACAAGGTT	465	0.11636840485193685	No Hit
TCAGCGTAGACTAGCTTGGTCATTTAGAGGAAGTAATATGCATCGATGAA	450	0.11261458534058404	No Hit
TCAGTACGAGTATGCTTGGTCATTAGAGGAGTAGCATCGATGAAGAACGC	405	0.10135312680652564	No Hit
TCAGTCACGTACTACTTGGTCATTTAGAGGAAGTAAAGTCGTAACAAGGT	402	0.10060236290425509	No Hit

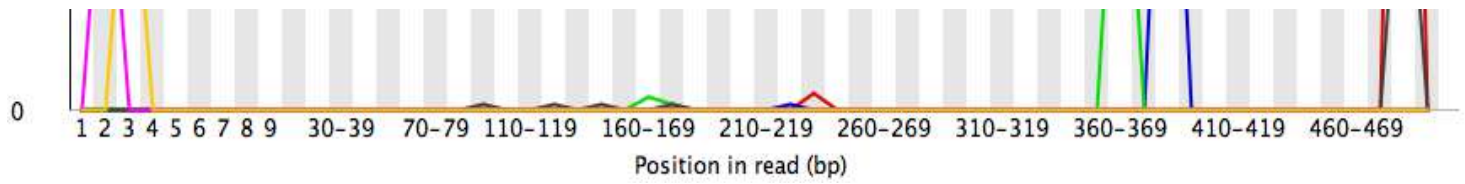
Adapter Content





 **Kmer Content**





Sequence	Count	PValue	Obs/Exp Max	Max Obs/Exp Position
CGCATAA	10	0.0076963855	1030.9615	480-489
GAGATGT	25	3.5106495E-9	523.90875	380-389
ATCAGAT	25	3.0247826E-4	415.7794	360-369
ATGACCC	30	5.334286E-4	344.05515	480-489
CAGCATA	1235	0.0	330.6081	2
AGCATAG	1235	0.0	330.6081	3
TATGCGC	1720	0.0	328.42105	9
GCGTAGA	2690	0.0	328.25	4
CAGCGTA	2720	0.0	327.68063	2
GATATCG	1105	0.0	327.44592	4
CAGATAC	700	0.0	327.20987	2
AGTATGC	1275	0.0	327.0725	9
AGATACC	1855	0.0	326.91098	9
GCATAGT	1250	0.0	326.6408	4
AGACTAT	2785	0.0	326.62924	5
CAGACGC	2315	0.0	326.2164	2
GAGTATG	1280	0.0	325.7949	8
TGATACG	2520	0.0	325.40643	5
AGACATA	1245	0.0	325.28635	3
CAGACAT	1245	0.0	325.28635	2

Produced by [FastQC](#) (version 0.11.4)