



Understanding the genetic complexity of puberty timing across the allele frequency spectrum

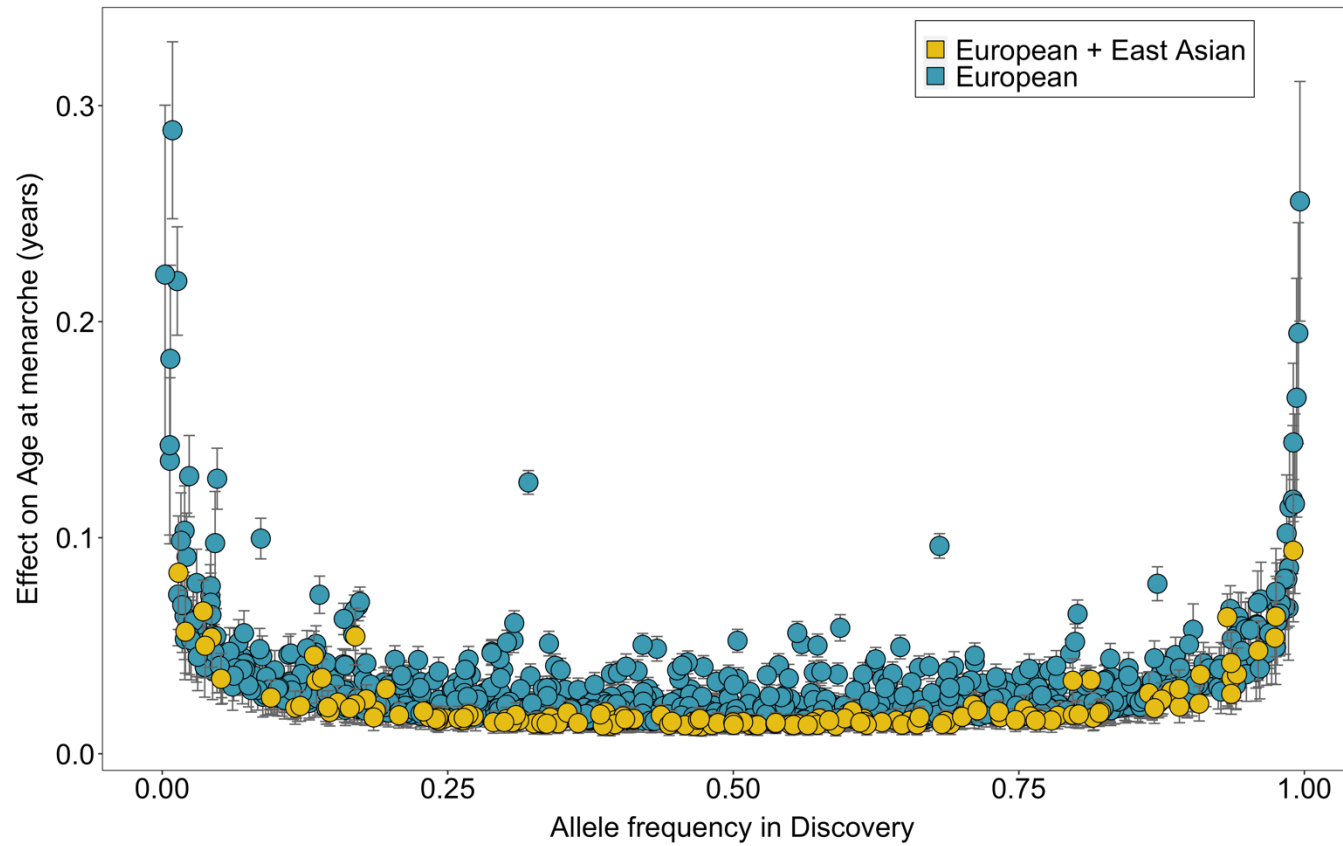
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Supplementary information

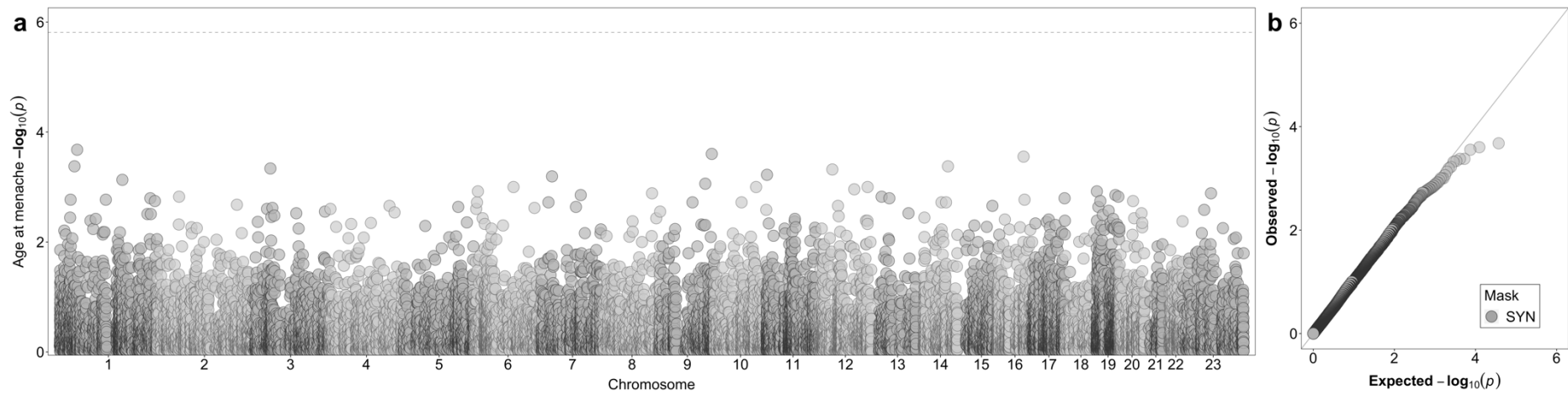
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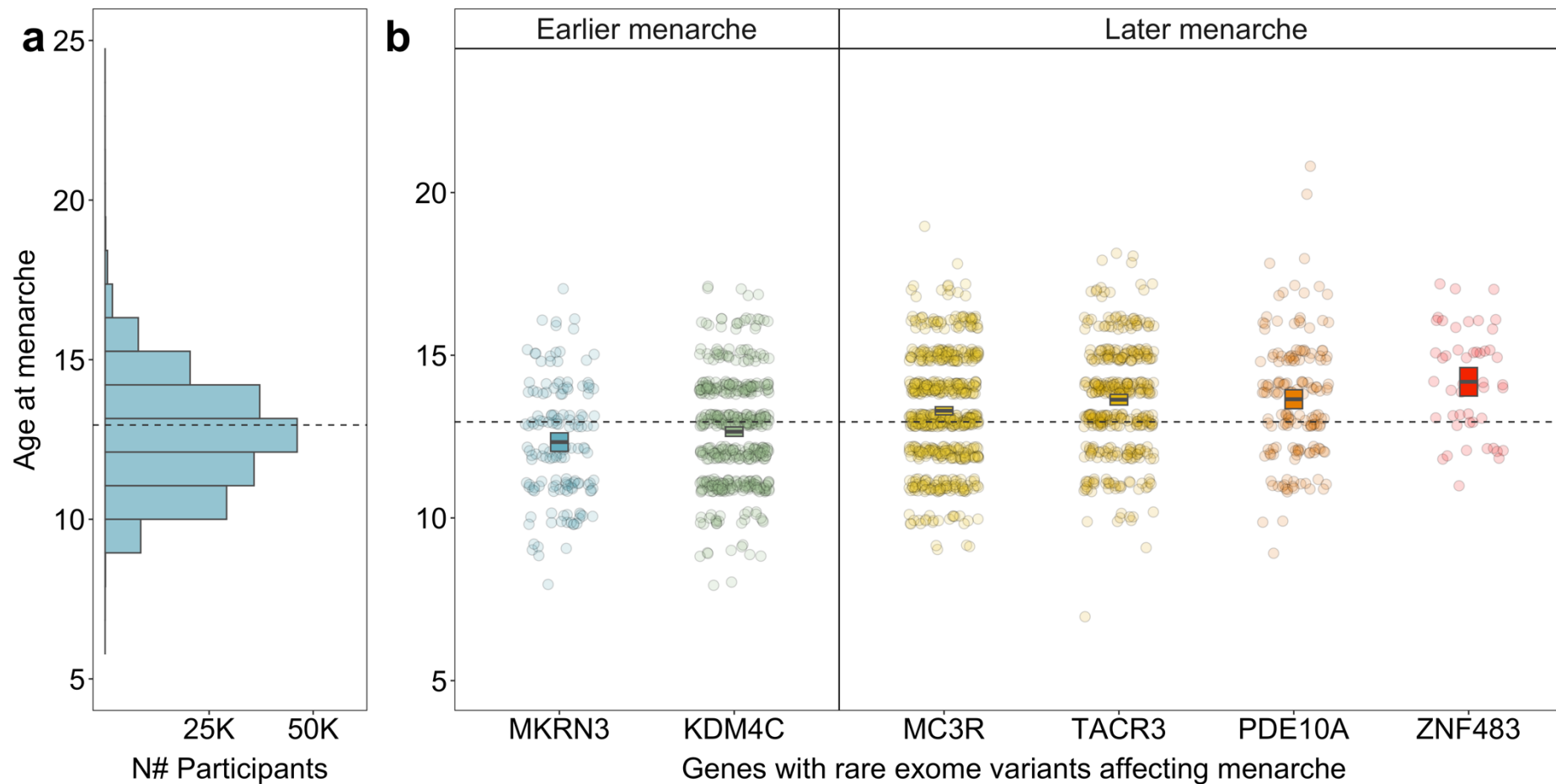
Supplementary Figures



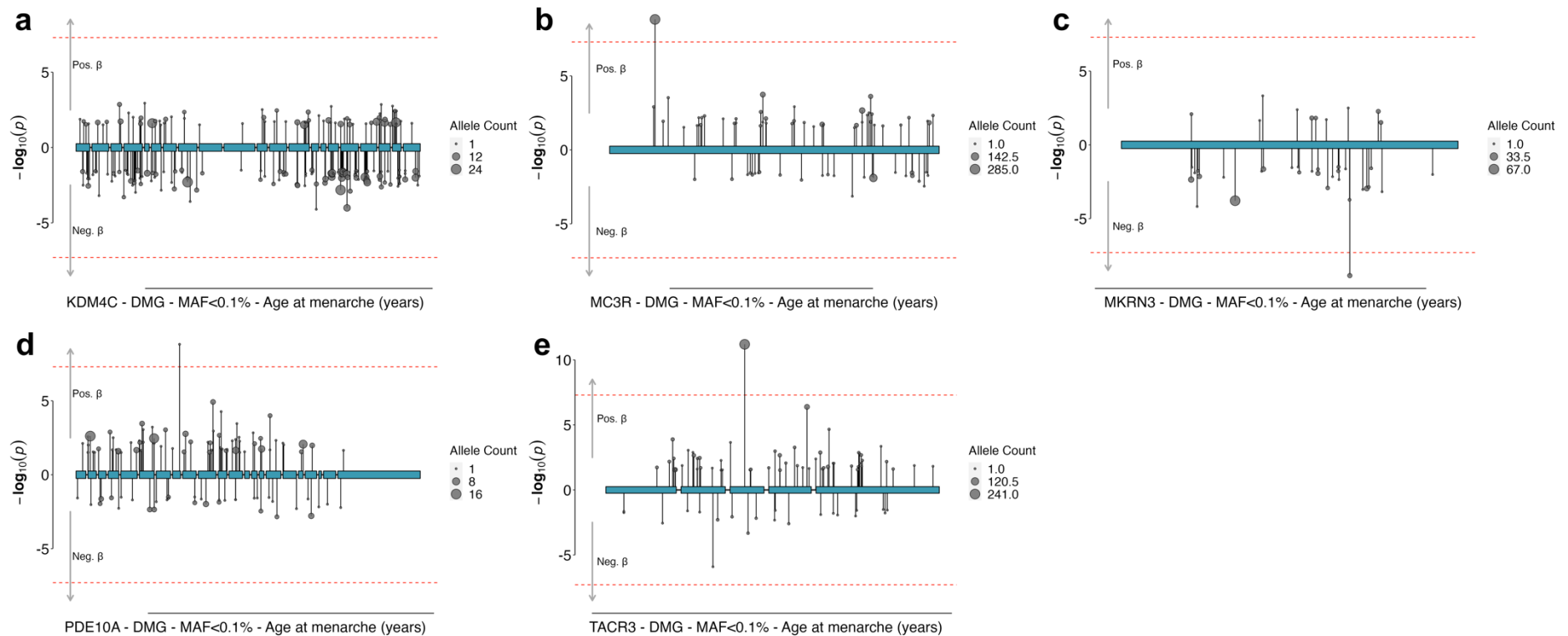
Supplementary Figure 1 | Distribution of effect sizes across 1080 independent signals with age at menarche (AAM). For each signal the effect size estimate and 95% confidence intervals are shown from the corresponding Discovery analysis, i.e. from the European-only or the ancestry combined meta-analysis, as indicated by point colours. Estimates are aligned towards the menarche-increasing alleles. Extended data are shown in Supplementary Table 2.



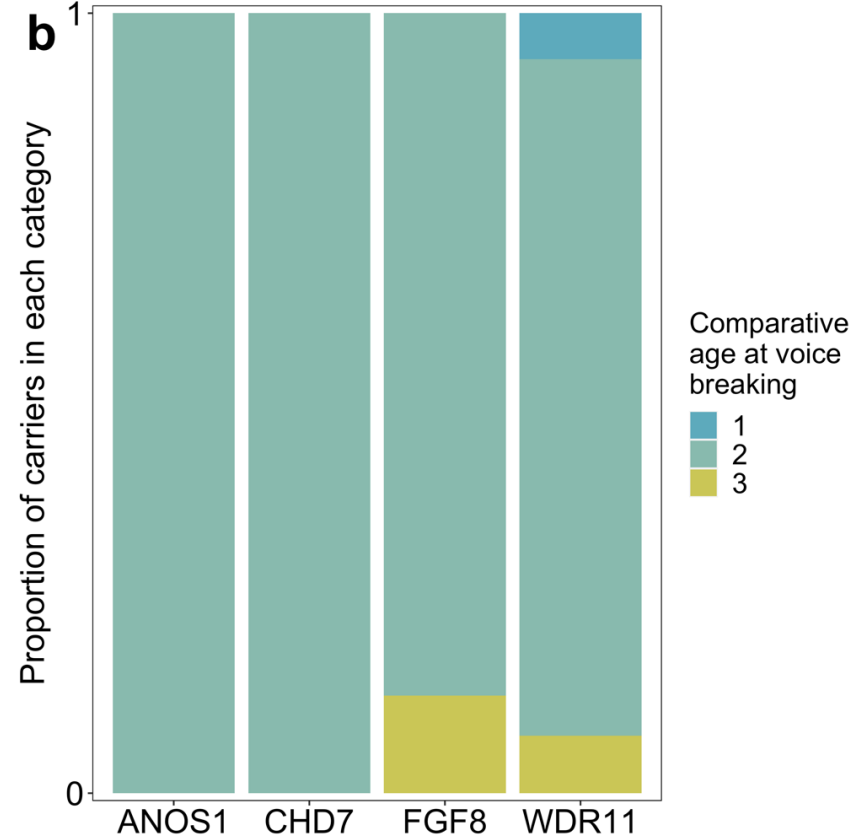
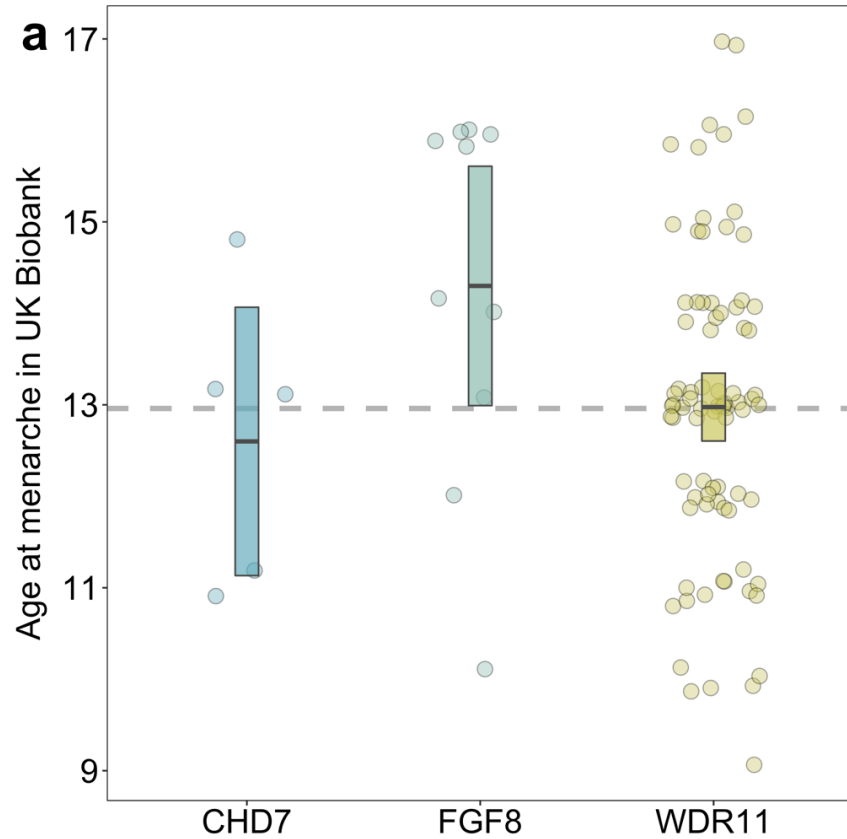
Supplementary Figure 2 | Synonymous variant associations with AAM. (a) Manhattan plot showing the gene burden associations from BOLT-LMM with age at menarche for the synonymous variant mask, as a negative control analysis. The horizontal line indicates the exome-wide significance threshold ($P < 1.54 \times 10^{-6}$). (b) Quantile-quantile (QQ) plot of the gene burden associations from BOLT-LMM for the synonymous variant mask.



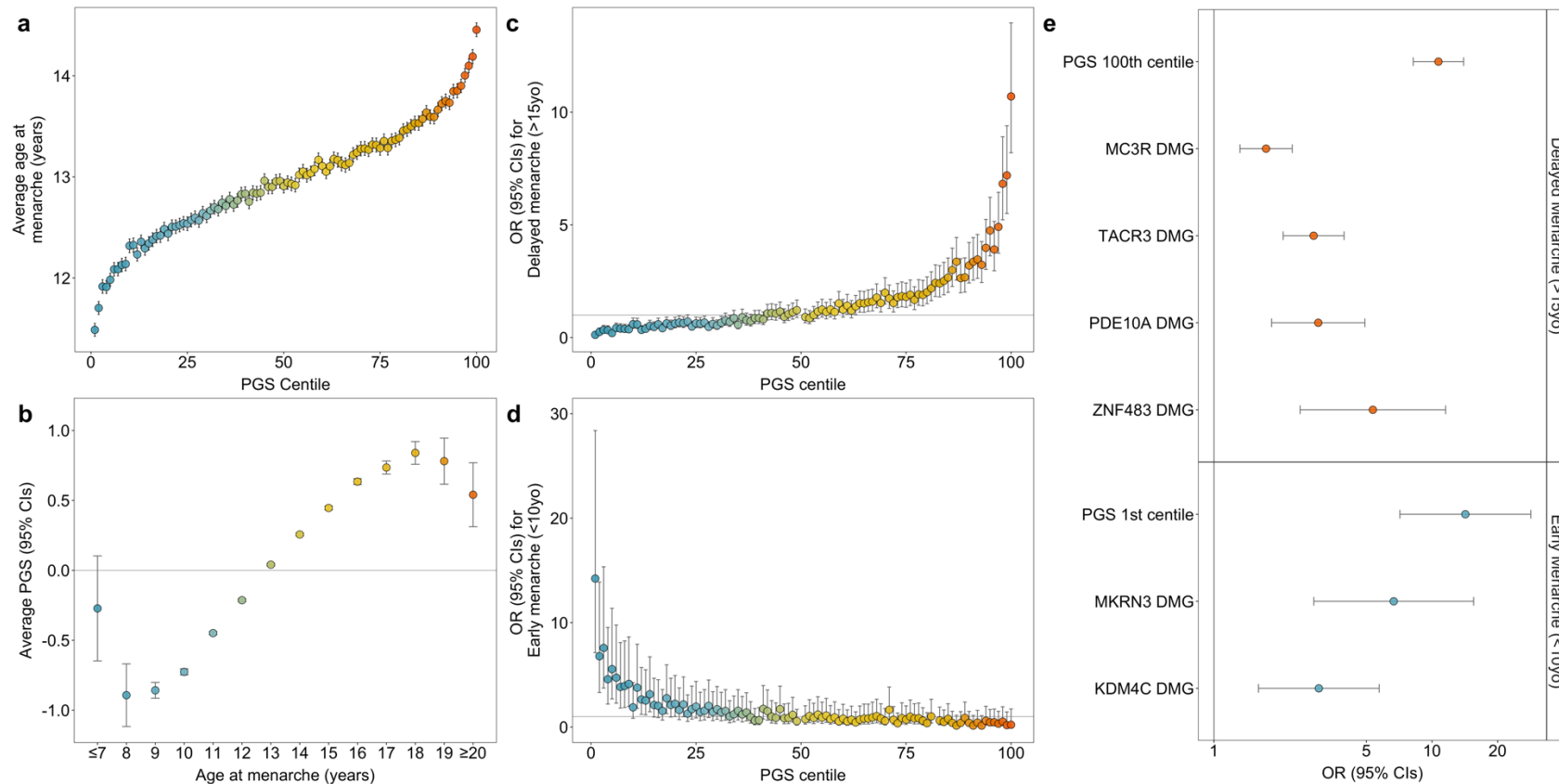
Supplementary Figure 3 | Distribution of age at menarche (AAM) in UK Biobank. Reported AAM in: (a) All white European unrelated female participants (N=187,941) (b) Carriers of qualifying rare variants in the associated genes. The horizontal dotted line indicates the mean AAM among non-carriers (N=185,929). Mean and 95% confidence intervals (CIs) for each carrier group are indicated by horizontal bars and boxes.



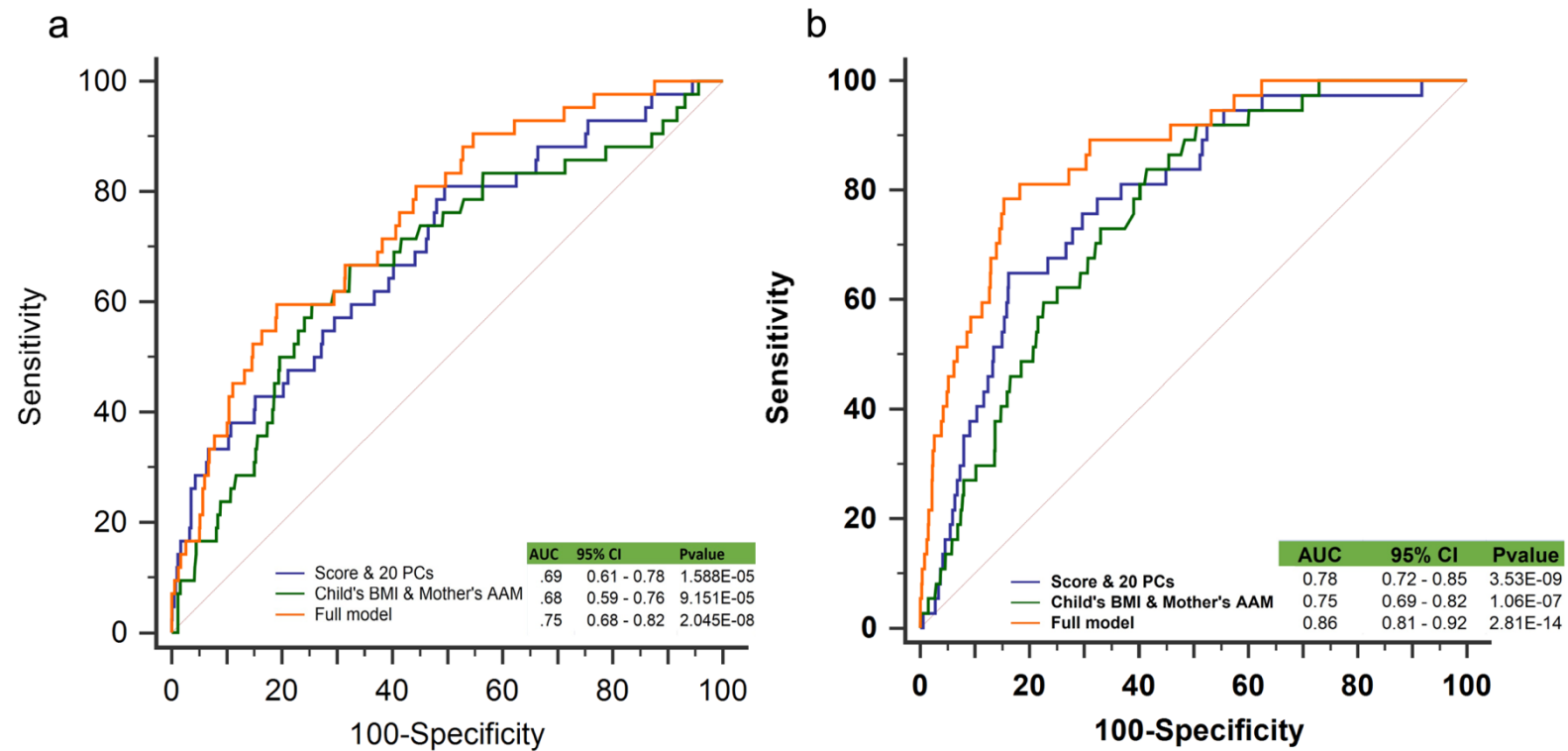
Supplementary Figure 4 | Variant-level associations with AAM in identified genes in UK Biobank. Rare exome variant associations from BOLT-LMM with AAM for variants in *KDM4C* (a), *MC3R* (b), *MKRN3* (c), *PDE10A* (d) and *TACR3* (e). The equivalent plot for *ZNF483* can be found in Fig. 3. Variant collapsing masks included variants with a minor allele frequency (MAF) < 0.1% and annotated as either high-confidence protein truncating variants (HC_PTV) or HC_PTV plus missense variants with a high CADD score (≥ 25 , denoted DMG). Each variant association is represented by a circle and vertical line: the line length indicates the P-value ($-\log_{10}$), in the direction of its effect on AAM in carriers of the rare allele, and the circle size indicates the number of carriers of each variant (i.e. allele count). Exons are indicated by the blue boxes.



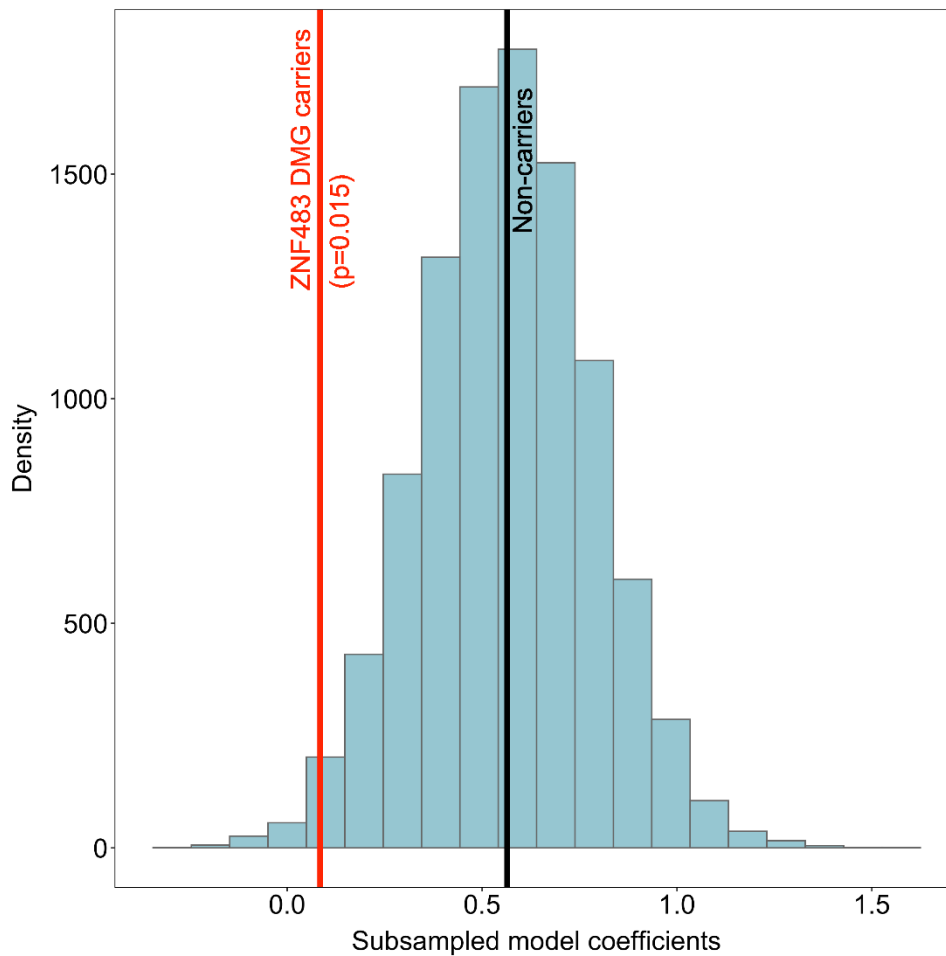
Supplementary Figure 5 | Puberty onset in carriers of IHH panel genes in UK Biobank. (a) Distribution of age at menarche (AAM) in UK Biobank white European unrelated female participants carrying qualifying rare variants in IHH panel genes. The horizontal dotted line indicates mean AAM among non-carriers (N=185,929). Mean and 95% confidence intervals (CIs) for each carrier group are indicated by horizontal bars and boxes. (b) Comparative age at voice breaking in men carrying qualifying rare variants in IHH panel genes. 1 indicates self-reported “younger than average age” at voice breaking, 2 “average age” and 3 “later than average age”.



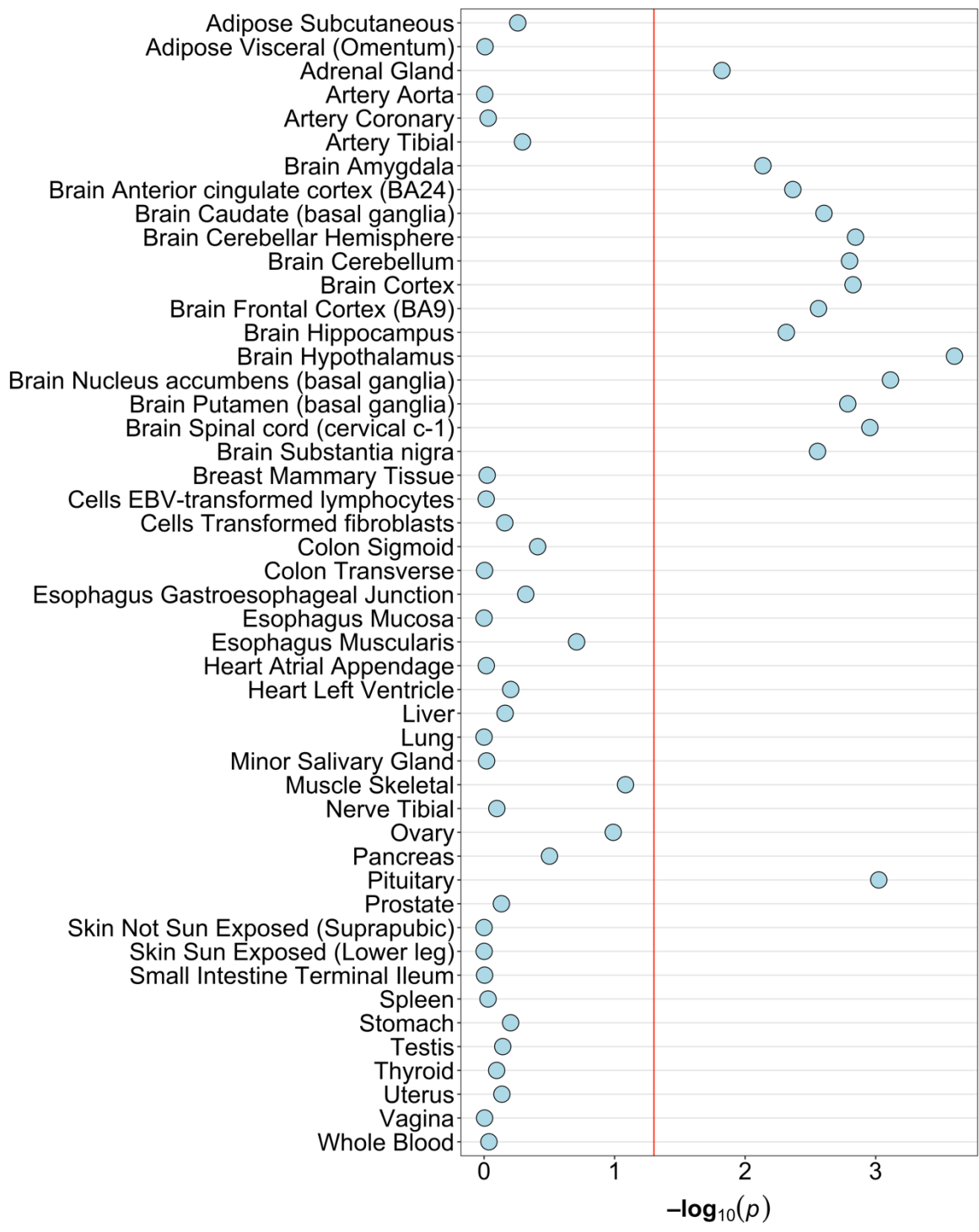
Supplementary Figure 6 | Polygenic score (PGS) associations with age at menarche (AAM). The PGS for AAM was derived using summary statistics from the European-only MA and excluding UK Biobank, and then applied to white European, unrelated female participants in UK Biobank (N=187,941). (a) Means with 95% confidence intervals (CIs) of AAM by PGS centile (N=1,880 per centile). (b) Means with 95% CIs of standardised PGS by AAM. (c-d) Associations of each PGS centile compared to the 50th PGS centile (OR with 95% CIs) with (c) delayed menarche (menarche after 15 years) and (d) early menarche (menarche before 10 years). Controls were women who reported menarche at 12 or 13 years. PGS centiles with fewer than 2 participants were omitted. (e) Gene burden associations between qualifying variants in the exome-identified genes compared to extreme PGS centiles, with delayed or early menarche defined as above, presented as ORs with 95% CIs. Extended data are shown in Supplementary Tables 8-10.



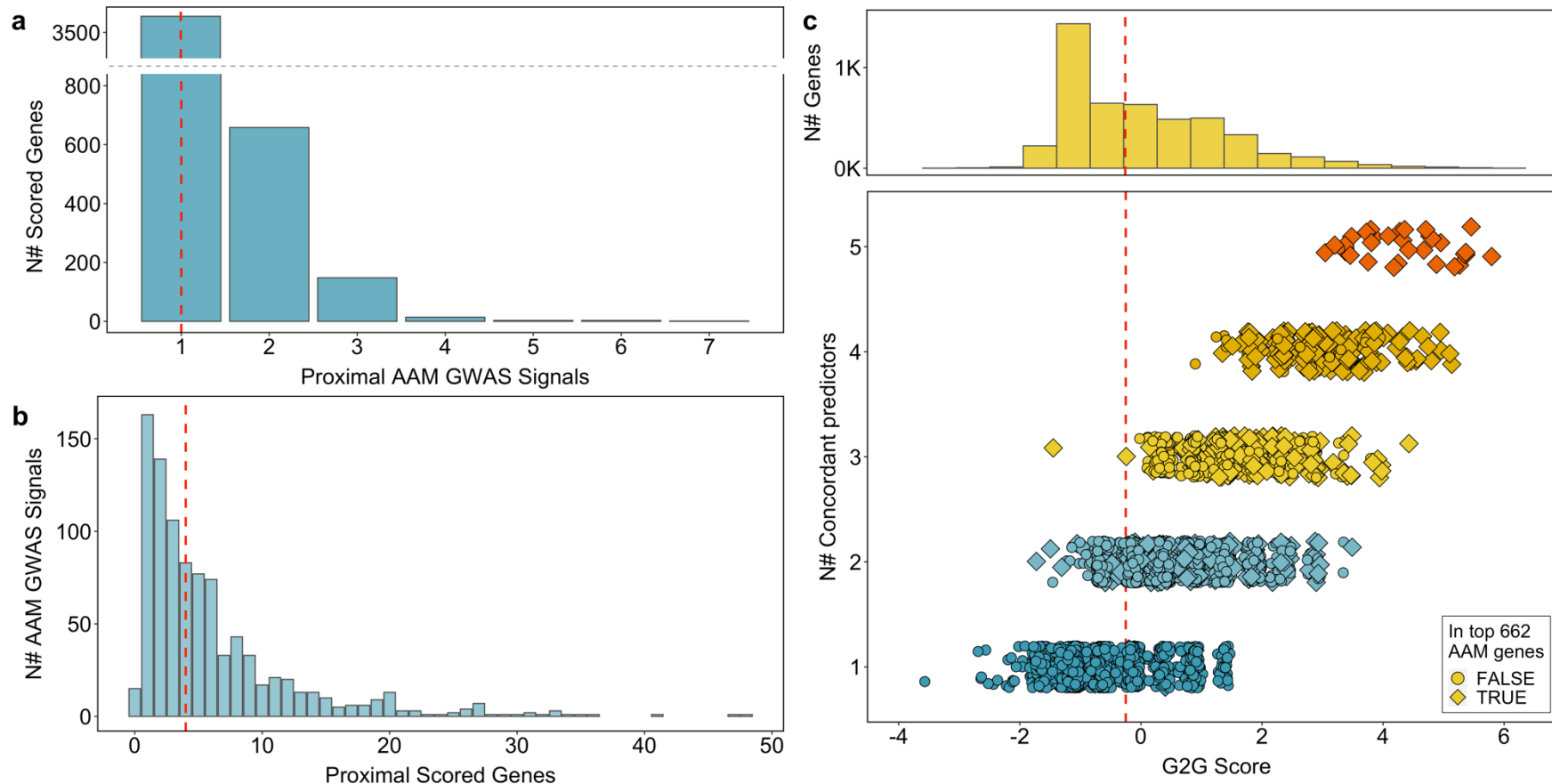
Supplementary Figure 7 | Receiver operating characteristic (ROC) curve for predicting extremes of age at menarche (AAM) in the ALSPAC study. Predictive performance of linear regressions adjusting for the genetic, clinical, and combined predictor and presented as ROC graphs against (a) early AAM and (b) delayed AAM in the ALSPAC study (N= 3,140). Extended data are shown in Supplementary Table 11.



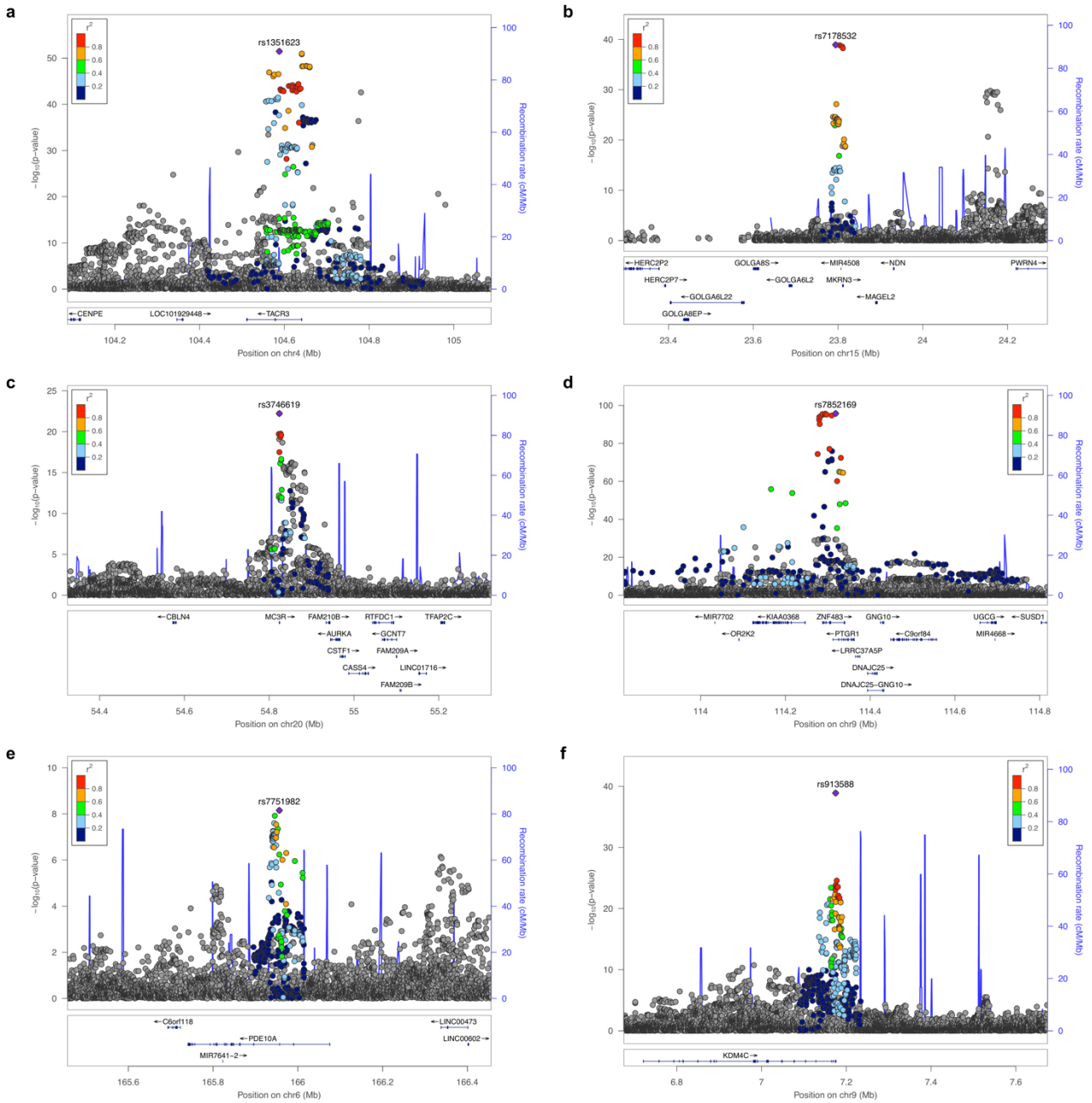
Supplementary Figure 8 | Epistatic interaction between the polygenic score (PGS) and carriage of rare damaging variants in *ZNF483* on age at menarche (AAM). Blue columns indicate the beta coefficients for the PGS-AAM association among 10,000 random subsamples of 49 white European unrelated female non-carriers in UK Biobank, tested in a linear model. The beta coefficient from the full sample of non-carriers is indicated by the black vertical line (0.564 years per SD, SE 0.003, $P < 2 \times 10^{-16}$) and the beta coefficient seen in *ZNF483* variant carriers is indicated by the red vertical line (difference in coefficients: -0.480, SE 0.214, $P = 0.025$). The probability of observing a beta coefficient smaller than that in *ZNF483* variant carriers was estimated as the proportion of subsampled coefficients that were smaller than 0.084 (i.e., $0.564 - 0.480$).



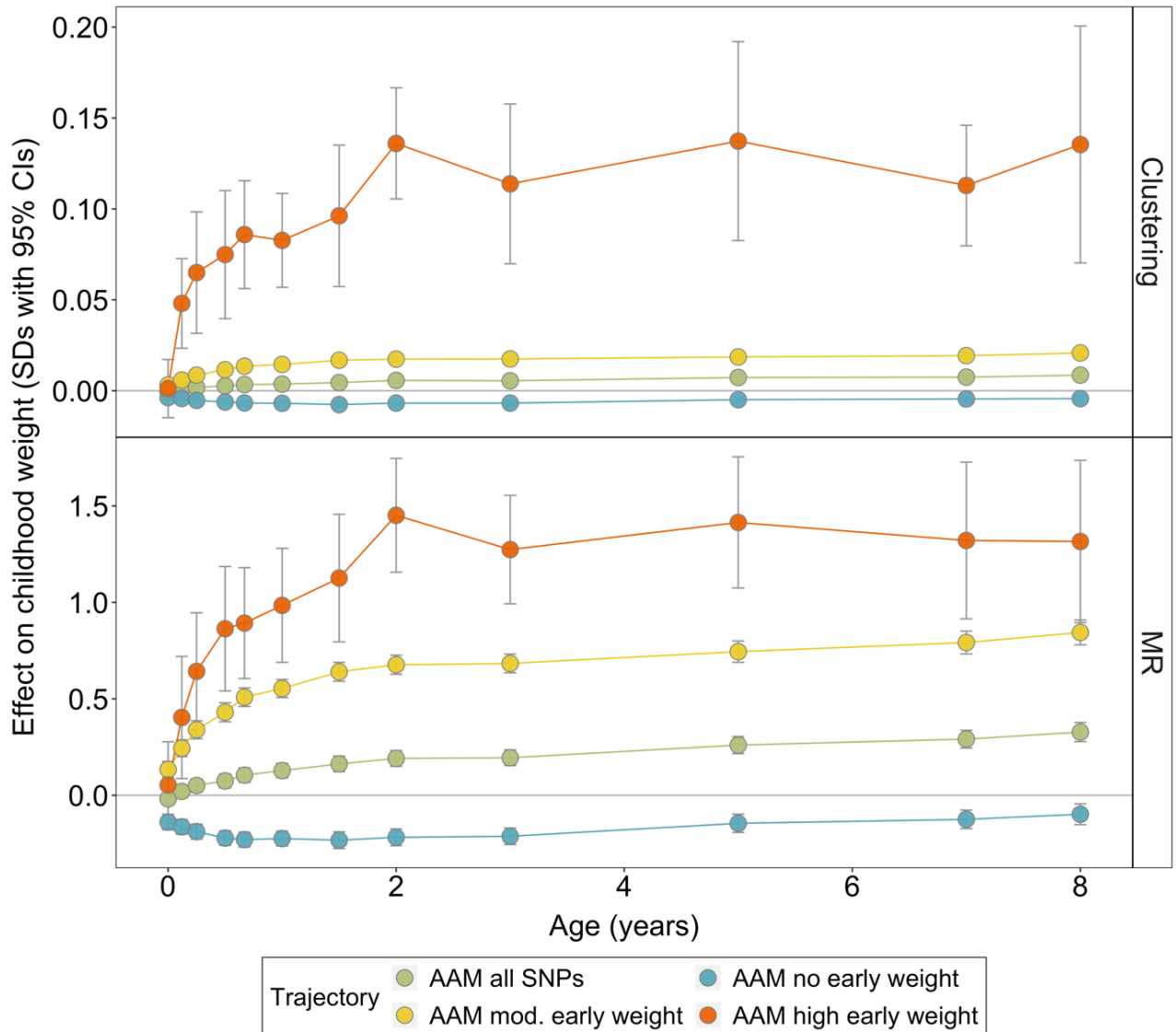
Supplementary Figure 9 | Tissue enrichment for age at menarche (AAM) GWAS associations. Linkage-disequilibrium score regression to specifically expressed genes (LDSC-SEG) was used to test for enrichment for AAM GWAS associations among European-only samples among genes specifically expressed across the different GTEx tissues. LDSC-SEG unadjusted P-values represent a one-sided test that the coefficient is greater than zero. Extended data are shown in Supplementary Table 17.



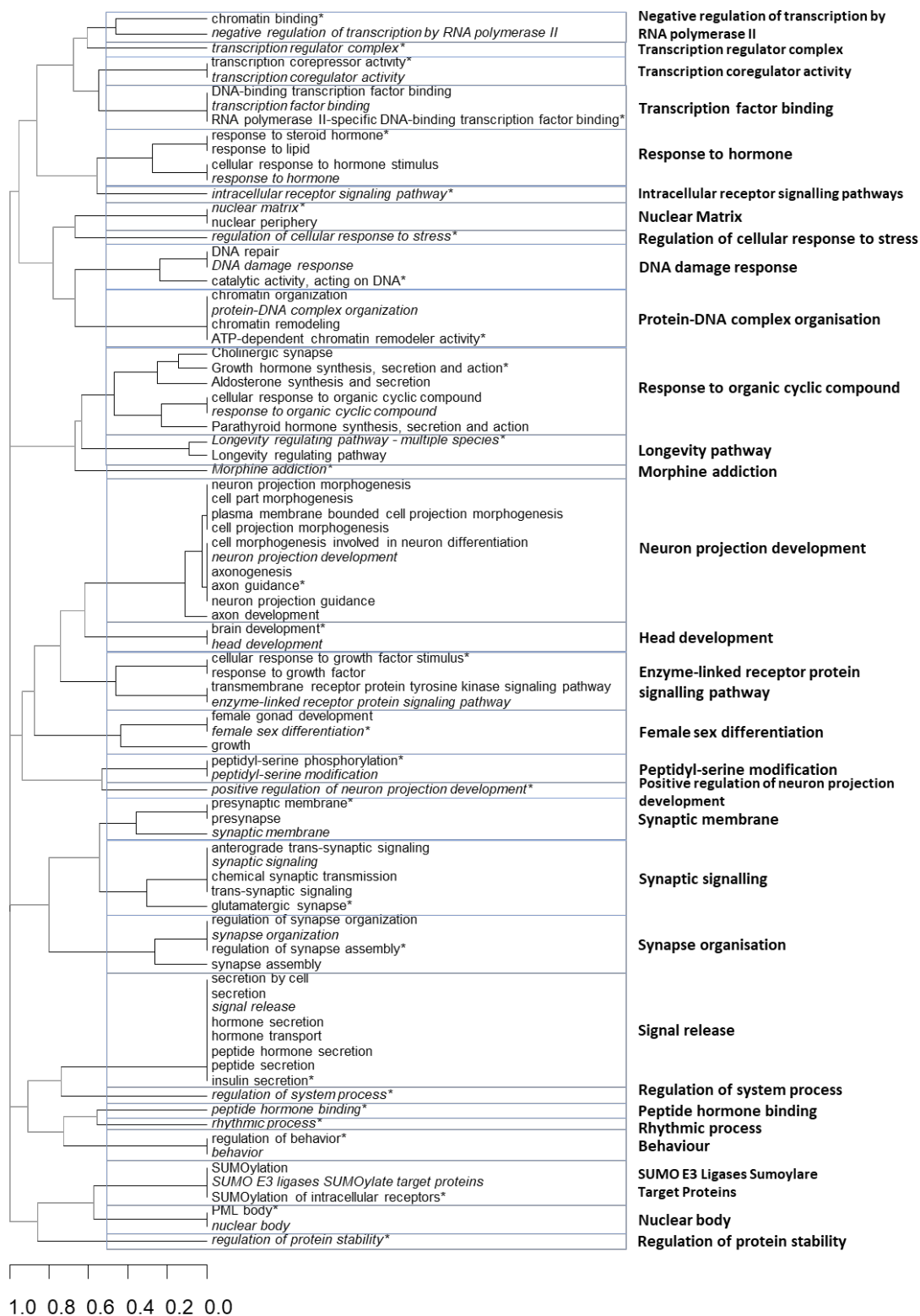
Supplementary Figure 10 | Distribution of age at menarche (AAM) GWAS signals and genes, as identified by GWAS to Genes (G2G). (a) Based on the European-only GWAS meta-analysis, 4,668 genes were implicated by G2G as potential regulators of AAM. Most were proximal to (within 500kb) only 1 GWAS signal (median signals per gene: 1; range: 1 to 7). (b) Conversely, each GWAS signal was annotated to be proximal (within 500kb) to (median) 4 genes (range: 0 to 48). (c) The 4,657 genes had a median G2G score -0.25 (range -3.57 to 5.88) derived from a maximum of 6 predictors (up to 5 observed). The 665 high confidence AAM genes were defined as the top-scoring gene at each independent AAM signal with at least 2 concordant predictors. Extended data are shown in Supplementary Table 13.



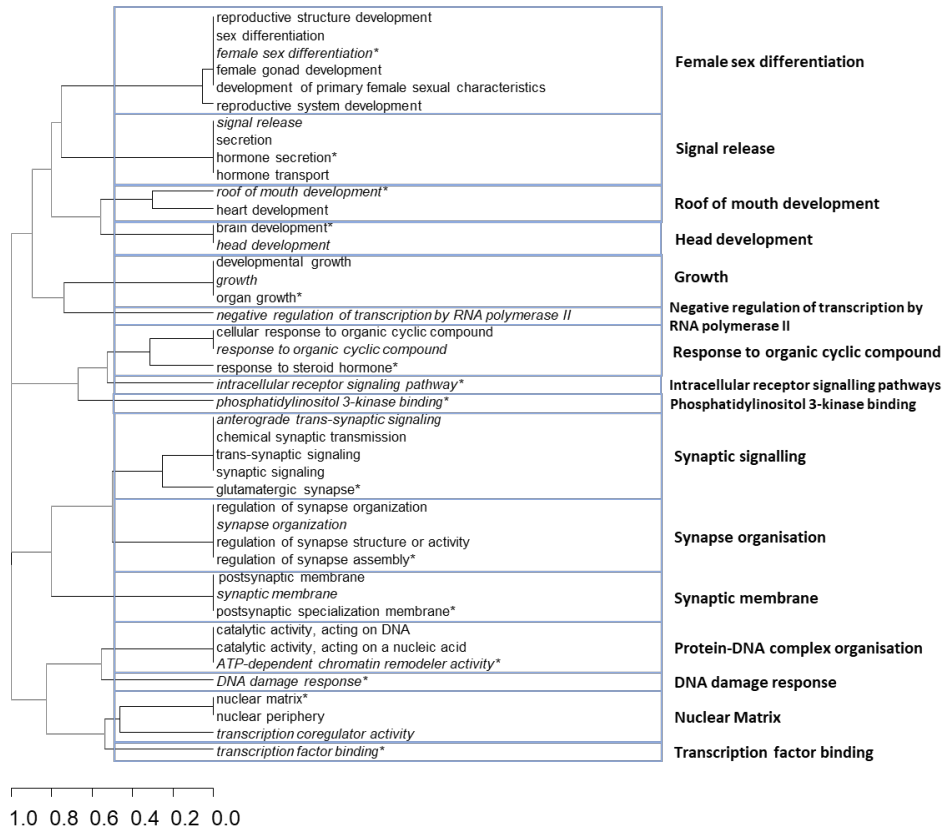
Supplementary Figure 11 | GWAS loci proximal to the six exome-wide significantly associated genes. Associations among European-only samples in regions surrounding the 6 genes ($\pm 500\text{kb}$) identified via exome-wide rare variant associations with AAM; (a) *TACR3*, (b) *MKRN3*, (c) *MC3R*, (d) *ZNF483*, (e) *PDE10A*, and (f) *KDM4C*.



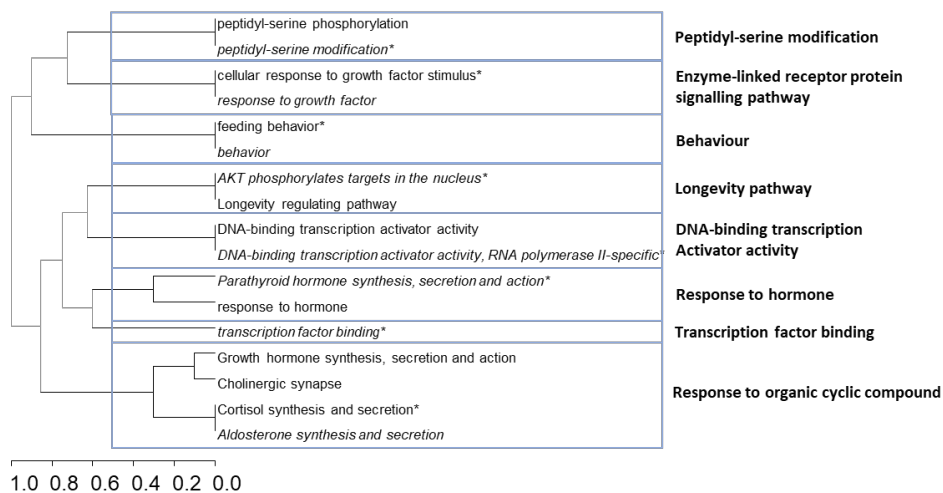
Supplementary Figure 12 | Mean childhood body weight trajectory for each of the three age at menarche (AAM) GWAS SNP clusters (aligned to AAM-decreasing alleles). Mendelian randomisation (MR) estimates with 95% confidence intervals (CIs) for all 1080 AAM signals, and the three AAM SNP clusters as separate exposures, on childhood body weight at 12 time-points (n=26,681 children). Extended data are shown in Supplementary Tables 21 and 22.



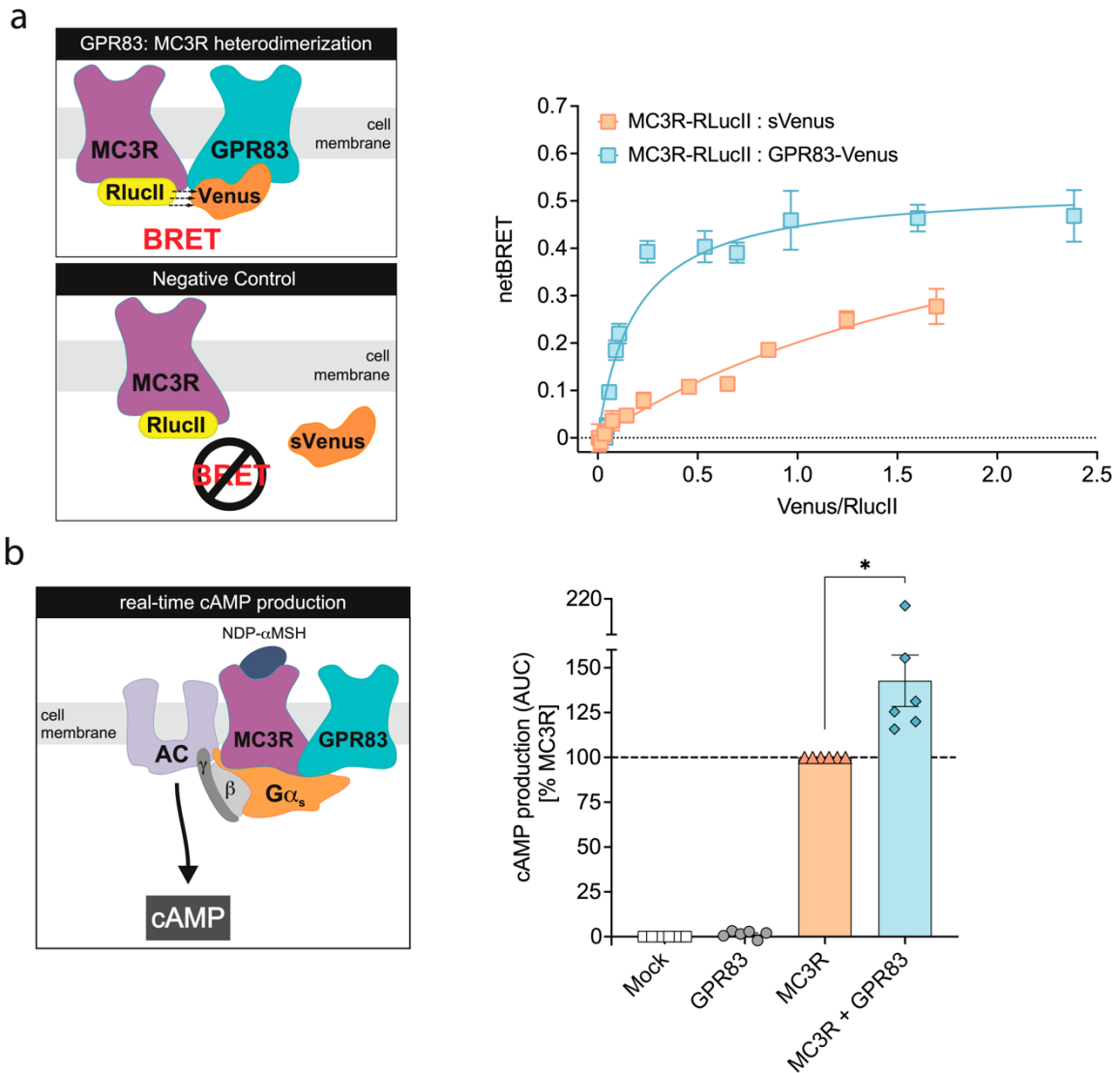
Supplementary Figure 13 | Dendrogram showing clustering of biological pathways enriched for the 665 high confidence AAM genes. In each cluster, the most significantly associated pathway is highlighted in italics and the most highly enriched pathway is marked by an asterisk. Extended data are shown in Supplementary Tables 23-24.



Supplementary Figure 14 | Clustering of biological pathways enriched for AAM genes in the “early weight gain” trajectory. In each cluster the most significant pathway is highlighted in italics and the most highly enriched pathway is marked with an asterisk. Extended data are shown in Supplementary Tables 25-26.



Supplementary Figure 15 | Clustering of biological pathways enriched for AAM genes in the “no early weight gain” trajectory. In each cluster the most significant pathway is highlighted in italics and the most highly enriched pathway is marked with an asterisk. Extended data are shown in Supplementary Tables 25-26.



Supplementary Figure 16 | GPR83-MC3R heterodimerization modulates canonical MC3R-cAMP signalling pathway. (a) BRET saturation curve from HEK293 cells co-transfected with a constant amount of MC3R-RlucII donor construct and increasing amounts of the GPR83-Venus acceptor construct, indicating a specific and saturable GPR83-MC3R interaction. The selectivity of the observed signal was further supported by the observation that co-expression of MC3R-RlucII with a soluble acceptor (sVenus) led to lower BRET signals that progressed linearly over the same range of acceptor/donor ratios. (b) NDP- α MSH-stimulated cAMP production area under the curve calculation from HEK293 cells transfected with *MC3R* or co-transfected with both *MC3R* and *GPR83*. The time-resolved data are shown in Figure 6. Data are expressed as a percentage of maximal control response [% MC3R] and plotted as mean \pm standard error from 6 independent experiments. Statistical significance was determined by unpaired t-test with Welch's correction; * $P < 0.05$. Extended data are shown in Supplementary Tables 29-31.

Consortia membership

The China Kadoorie Biobank Collaborative Group

International Steering Committee: Junshi Chen, Zhengming Chen (PI), Robert Clarke, Rory Collins, Yu Guo, Liming Li (PI), Chen Wang, Jun Lv, Richard Peto, Robin Walters.

International Co-ordinating Centre, Oxford: Daniel Avery, Derrick Bennett, Ruth Boxall, Sushila Burgess, Ka Hung Chan, Yumei Chang, Yiping Chen, Zhengming Chen, Johnathan Clarke; Robert Clarke, Huaidong Du, Ahmed Edris, Hannah Fry, Simon Gilbert, Mike Hill, Michael Holmes, Pek Kei Im, Andri Iona, Maria Kakkoura, Christiana Kartsonaki, Hubert Lam, Kuang Lin, Mohsen Mazidi, Iona Millwood, Sam Morris, Qunhua Nie, Alfred Pozarickij, Paul Ryder, Saredo Said, Dan Schmidt, Paul Sherliker, Becky Stevens, Iain Turnbull, Robin Walters, Lin Wang, Neil Wright, Ling Yang, Xiaoming Yang, Pang Yao.

National Co-ordinating Centre, Beijing: Yu Guo, Xiao Han, Can Hou, Qingmei Xia, Chao Liu, Jun Lv, Pei Pei, Canqing Yu.

Regional Co-ordinating Centres:

Gansu: Gansu Provincial CDC – Caixia Dong, Pengfei Ge, Xiaolan Ren. Maiji CDC – Zhongxiao Li, Enke Mao, Tao Wang, Hui Zhang, Xi Zhang.

Haikou: Hainan Provincial CDC – Jinyan Chen, Ximin Hu, Xiaohuan Wang. Meilan CDC – Zhendong Guo, Huimei Li, Yilei Li, Min Weng, Shukuan Wu.

Harbin: Heilongjiang Provincial CDC – Shichun Yan, Mingyuan Zou, Xue Zhou. Nangang CDC – Ziyang Guo, Quan Kang, Yanjie Li, Bo Yu, Qinai Xu.

Henan: Henan Provincial CDC – Liang Chang, Lei Fan, Shixian Feng, Ding Zhang, Gang Zhou. Huixian CDC – Yulian Gao, Tianyou He, Pan He, Chen Hu, Huarong Sun, Xukui Zhang.

Hunan: Hunan Provincial CDC – Biyun Chen, Zhongxi Fu, Yuelong Huang, Huilin Liu, Qiaohua Xu, Li Yin. Liuyang CDC – Huajun Long, Xin Xu, Hao Zhang, Libo Zhang.

Liuzhou: Guangxi Provincial CDC – Naying Chen, Duo Liu, Zhenzhu Tang. Liuzhou CDC – Ningyu Chen, Qilian Jiang, Jian Lan, Mingqiang Li, Yun Liu, Fanwen Meng, Jinhuai Meng, Rong Pan, Yulu Qin, Ping Wang, Sisi Wang, Liuping Wei, Liyuan Zhou.

Qingdao: Qingdao CDC – Liang Cheng, Ranran Du, Ruqin Gao, Feifei Li, Shanpeng Li, Yongmei Liu, Feng Ning, Zengchang Pang, Xiaohui Sun, Xiaocao Tian, Shaojie Wang, Yaoming Zhai, Hua Zhang. Licang CDC – Wei Hou, Silu Lv, Junzheng Wang.

Sichuan: Sichuan Provincial CDC – Xiaofang Chen, Xianping Wu, Ningmei Zhang, Weiwei Zhou. Pengzhou CDC – Xiaofang Chen, Jianguo Li, Jiaqiu Liu, Guojin Luo, Qiang Sun, Xunfu Zhong.

Suzhou: Jiangsu Provincial CDC – Jian Su, Ran Tao, Ming Wu, Jie Yang, Jinyi Zhou, Yonglin Zhou. Suzhou CDC – Yihe Hu, Yujie Hua, Jianrong Jin Fang Liu, Jingchao Liu, Yan Lu, Liangcai Ma, Aiyu Tang, Jun Zhang.

Zhejiang: Zhejiang Provincial CDC – Weiwei Gong, Ruying Hu, Hao Wang, Meng Wang, Min Yu. Tongxiang CDC – Lingli Chen, Qijun Gu, Dongxia Pan, Chunmei Wang, Kaixu Xie, Xiaoyi Zhang.

Lifelines Cohort Study

Behrooz Z Alizadeh¹, H Marike Boezen¹, Lude Franke², Pim van der Harst³, Gerjan Navis⁴, Marianne Rots⁵, Harold Snieder¹, Morris Swertz², Bruce HR Wolffenbuttel⁶, and Cisca Wijmenga²

1. Department of Epidemiology, University of Groningen, University Medical Center Groningen, The Netherlands
2. Department of Genetics, University of Groningen, University Medical Center Groningen, The Netherlands
3. Department of Cardiology, University of Groningen, University Medical Center Groningen, The Netherlands
4. Department of Internal Medicine, Division of Nephrology, University of Groningen, University Medical Center Groningen, The Netherlands
5. Department of Pathology and Medical Biology, University of Groningen, University Medical Center Groningen, The Netherlands
6. Department of Endocrinology, University of Groningen, University Medical Center Groningen, The Netherlands

Danish Blood Donor Study

Karina Banasik¹, Jakob Bay², Jens Kjærgaard Boldsen³, Thorsten Brodersen², Søren Brunak¹, Kristoffer Burgdorf¹, Mona Ameri Chalmer⁴, Maria Didriksen⁵, Khoa Manh Dinh³, Joseph Dowsett⁵, Christian Erikstrup^{3,6}, Bjarke Feenstra^{5,7}, Frank Geller^{5,7}, Daniel Gudbjartsson⁸, Thomas Folkmann Hansen⁴, Lotte Hindhede³, Henrik Hjalgrim^{9,7}, Rikke Louise Jacobsen⁵, Gregor Jemec¹⁰, Bitten Aagaard Jensen¹¹, Katrine Kaspersen³, Bertram Dalskov Kjerulff³, Lisette Kogelman⁴, Margit Anita Hørup Larsen⁵, Ioannis Louloudis¹, Agnete Lundgaard¹, Susan Mikkelsen³, Christina Mikkelsen⁵, Ioanna Nissen⁵, Mette Nyegaard¹², Sisse Rye Ostrowski^{5,13}, Ole Birger Pedersen^{2,13}, Alexander Pil Henriksen¹, Palle Duun Rohde¹², Klaus Rostgaard^{9,7}, Michael Schwinn⁵, Kari Stefansson⁸, Hreinn Stefánsson⁸, Erik Sørensen⁵, Unnur Þorsteinsdóttir⁸, Lise Wegner Thørner⁵, Mie Topholm Bruun¹⁴, Henrik Ullum¹⁵, Thomas Werge^{16,13}, and David Westergaard¹

1. Novo Nordisk Foundation Center for Protein Research, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark,
2. Department of Clinical Immunology, Zealand University Hospital, Køge, Denmark,
3. Department of Clinical Immunology, Aarhus University Hospital, Aarhus, Denmark,
4. Danish Headache Center, Department of Neurology, Copenhagen University Hospital, Rigshospitalet-Glostrup, Copenhagen, Denmark,
5. Department of Clinical Immunology, Copenhagen University Hospital,

- Rigshospitalet, Copenhagen, Denmark,
 6. Department of Clinical Medicine, Health, Aarhus University, Aarhus, Denmark,
 7. Department of Epidemiology Research, Statens Serum Institut, Copenhagen, Denmark,
 8. deCODE Genetics, Reykjavik, Iceland,
 9. Danish Cancer Society Research Center, Copenhagen, Denmark,
 10. Department of Dermatology, Zealand University hospital, Roskilde, Denmark,
 11. Department of Clinical Immunology, Aalborg University Hospital, Aalborg, Denmark,
 12. Department of Health Science and Technology, Faculty of Medicine, Aalborg University, Aalborg, Denmark,
 13. Department of Clinical Medicine, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark,
 14. Department of Clinical Immunology, Odense University Hospital, Odense, Denmark,
 15. Statens Serum Institut, Copenhagen, Denmark,
 16. Institute of Biological Psychiatry, Mental Health Centre, Sct. Hans, Copenhagen University Hospital, Roskilde, Denmark.

The Ovarian Cancer Association Consortium

CM Phelan¹, KB Kuchenbaecker^{2,3}, JP Tyrer⁴, SP Kar⁴, K Lawrenson⁵, SJ Winham⁶, J Dennis⁴, A Pirie⁴, M J Riggan⁷, G Chornokur⁸, MA Earp⁹, PC Lyra, Jr.⁸, JM. Lee¹⁰, S Coetzee¹⁰, J Beesley¹¹, L McGuffog¹², P Soucy¹³, E Dicks⁴, A Lee¹², D Barrowdale¹², J Lecarpentier¹², G Leslie¹², CM Aalfs¹⁴, KKH Aben^{15,16}, M Adams¹⁷, J Adlard¹⁸, IL Andrusis¹⁹, H Anton-Culver²⁰, N Antonenkova²¹, AOCS study group²², G Aravantinos²³, N Arnold²⁴, BK Arun²⁵, B Arver²⁶, J Azzollini²⁷, J Balmaña²⁸, SN Banerjee²⁹, L Barjhoux³⁰, RB Barkardottir^{31,32}, Y Bean³³, MW Beckmann³⁴, A BeeghlyFadiel³⁵, J Benitez³⁶, M Bermisheva³⁷, MQ Bernardini³⁸, MJ Birrer³⁹, L Bjorge^{40,41}, A Black⁴², K Blankstein⁴³, MJ Blok⁴⁴, C Bodelon⁴², N Bogdanova⁴⁵, A Bojesen⁴⁶, B Bonanni⁴⁷, Å Borg⁴⁸, AR Bradbury⁴⁹, JD Brenton⁵⁰, C Brewer⁵¹, L Brinton⁴², P Broberg⁵², A Brooks-Wilson⁵³, F Bruinsma⁵⁴, J Brunet⁵⁵, B Buecher⁵⁶, R Butzow⁵⁷, SS Buys⁵⁸, T Caldes⁵⁹, MA Caligo⁶⁰, I Campbell^{61,62}, R Cannioto⁶³, ME Carney⁶⁴, T Cescon⁴³, SB Chan⁶⁵, J Chang-Claude^{66,67}, S Chanock⁴², X Qing Chen¹¹, Y-E Chiew^{68,69}, J Chiquette⁷⁰, WK Chung⁷¹, KBM Claes⁷², T Conner⁵⁸, LS Cook⁷³, J Cook⁷⁴, DW Cramer⁷⁵, JM Cunningham⁷⁶, AA D’Aloisio⁷⁷, MB Daly⁷⁸, F Damiola³⁰, S Dina Damirovna⁷⁹, A Dansonka-Mieszkowska⁸⁰, F Dao⁸¹, R Davidson⁸², A DeFazio^{68,69}, C Delnatte⁸³, KF Doherty¹⁷, O Diez^{84,85}, Y Chun Ding⁸⁶, J Anne Doherty⁸⁷, SM Domchek⁴⁹, CM Dorfling⁸⁸, T Dörk⁸⁹, L Dossus⁹⁰, M Duran⁹¹, M Dürst⁹², B Dworniczak⁹³, D Eccles⁹⁴, T Edwards³⁵, R Eeles⁹⁵, U Eilber⁶⁶, B Ejlersen⁹⁶, AB Ekici⁹⁷, S Ellis¹², M Elvira⁷⁹, Study EMBRACE²², KH Eng⁹⁸, C Engel⁹⁹, DG Evans¹⁰⁰, PA Fasching^{101,34}, S Ferguson³⁸, S Fert Ferrer¹⁰², JM Flanagan¹⁰³, ZC Fogarty⁶, RT Fortner⁶⁶, F Fostira¹⁰⁴, WD Foulkes¹⁰⁵, G Fountzilas¹⁰⁶, BL Fridley¹⁰⁷, TM Friebel¹⁰⁸, E Friedman¹⁰⁹, D Frost¹², PA Ganz¹¹⁰, J Garber¹¹¹, MJ García³⁶, V GarciaBarberan⁵⁹, A Gehrig¹¹², GEMO Study Collaborators²², A GentryMaharaj¹¹³, A-M Gerdes¹¹⁴, GG Giles^{54,115,116}, R Glasspool¹¹⁷, G Glendon¹¹⁸, AK Godwin¹¹⁹, DE Goldgar¹²⁰, T Goranova⁵⁰, M Gore¹²¹, MH Greene¹²², J Gronwald¹²³, S Gruber¹²⁴, E Hahnen¹²⁵, CA Haiman¹²⁶, N Håkansson¹²⁷, U Hamann¹²⁸, TVO Hansen¹²⁹, PA Harrington⁴, HR Harris¹²⁷, J Hauke¹²⁵ HEBON Study²², A Hein³⁴, A Henderson¹³⁰, MAT Hildebrandt¹³¹, P Hillemanns¹³², S Hodgson¹³³, CK Høgdall¹³⁴, E Høgdall^{135,136}, FBL Hogervorst¹³⁷, H

Holland¹¹, MJ Hooning¹³⁸, K Hosking⁴, R-Y Huang¹³⁹, PJ Hulick¹⁴⁰, J Hung^{68,69}, DJ Hunter¹⁴¹, DG Huntsman¹⁴², T Huzarski¹²³, EN Imyanitov¹⁴³, C Isaacs¹⁴⁴, ES Iversen¹⁴⁵, L Izatt¹⁴⁶, A Izquierdo⁵⁵, A Jakubowska¹²³, P James¹⁴⁷, R Janavicius^{148,149}, M Jernetz¹⁵⁰, A Jensen¹³⁵, U Birk Jensen¹⁵¹, EM John¹⁵², S Johnatty¹¹, ME Jones¹⁵³, P Kannisto¹⁵⁰, BY Karlan⁵, A Karnezis¹⁴², K Kast¹⁵⁴, KconFab Investigators²², CJ Kennedy^{68,69}, E Khusnutdinova³⁷, LA Kiemeney¹⁵, JI Kiiski¹⁵⁵, S-W Kim¹⁵⁶, SK Kjaer^{134,135}, M Köbel¹⁵⁷, RK Kopperud^{40,41}, TA Kruse¹⁵⁸, J Kupryjanczyk⁸⁰, A Kwong^{159,160,161}, Y Laitman¹⁰⁹, D Lambrechts^{162,163}, N Larrañaga^{164,165}, MC Larson⁶, C Lazaro¹⁶⁶, ND Le¹⁶⁷, L Le Marchand¹⁶⁸, J Won Lee¹⁶⁹, SB Lele¹⁷⁰, A Leminen¹⁵⁵, D Leroux¹⁷¹, J Lester⁵, F Lesueur¹⁷², DA Levine⁸¹, D Liang¹⁷³, C Liebrich¹⁷⁴, J Lilyquist¹⁷⁵, L Lipworth¹⁷⁶, J Lissowska¹⁷⁷, KH Lu¹⁷⁸, J Lubiński¹²³, C Luccarini⁴, L Lundvall¹⁷⁹, PL Mai¹²², G Mendoza-Fandiño⁸, S Manoukian²⁷, L FAG Massuger¹⁵, T May³⁸, S Mazoyer¹⁸⁰, JN McAlpine¹⁸¹, V McGuire¹⁸², JR McLaughlin¹⁸³, I McNeish¹⁸⁴, H MeijersHeijboer¹⁸⁵, A Meindl¹⁸⁶, U Menon¹¹³, AR Mensenkamp¹⁸⁷, MA Merritt¹⁸⁸, RL Milne^{54,115}, G Mitchell^{147,189}, F Modugno^{190,191,192}, J Moes-Sosnowska⁸⁰, M Moffitt^{193,194}, M Montagna¹⁹⁵, KB Moysich⁶³, AM Mulligan^{196,197}, J Musinsky¹⁹⁸, KL Nathanson⁴⁹, L Nedergaard¹⁹⁹, RB Ness²⁰⁰, SL Neuhausen⁸⁶, H Nevanlinna¹⁵⁵, D Niederacher²⁰¹, RL Nussbaum²⁰², K Odunsi¹⁷⁰, E Olah²⁰³, OI Olopade²⁰⁴, H Jernström²⁰⁵, C Olswold¹⁷⁵, DM O'Malley²⁰⁶, K Ong²⁰⁷, NC Onland-Moret²⁰⁸, OPAL study group²², N Orr²⁰⁹, S Orsulic⁵, A Osorio³⁶, D Palli²¹⁰, L Papi²¹¹, T-W Park-Simon¹³², J Paul²¹², CL Pearce^{213,126}, I Søkilde Pedersen²¹⁴, PHM Peeters²⁰⁸, B Peissel²⁷, A Peixoto²¹⁵, T Pejovic^{193,194}, LM Pelttari¹⁵⁵, JB Permeth⁸, P Peterlongo²¹⁶, L Pezzani²⁷, G Pfeiler²¹⁷, K-A Phillips^{189,115,218}, M Piedmonte²¹⁹, MC Pike^{126,220}, AM Piskorz⁵⁰, SR Poblete¹⁷⁰, T Poczta²⁰³, EM Poole²²¹, B Poppe⁷², ME Porteous²²², F Prieur²²³, D Prokofyeva⁷⁹, E Pugh¹⁷, M Angel Pujana²²⁴, P Pujol²²⁵, P Radice²²⁶, J Rantala²²⁷, C Rappaport-Fuerhauser²¹⁷, G Rennert²²⁸, K Rhiem¹²⁵, P Rice⁴³, A Richardson²²⁹, M Robson²³⁰, GC Rodriguez²³¹, C Rodríguez-Antona²³², J Romm¹⁷, MA Rookus²³³, M Anne Rossing²³⁴, JH Rothstein¹⁸², A Rudolph⁶⁶, IB Runnebaum⁹², HB Salvesen^{40,41}, DP Sandler²³⁵, MJ Schoemaker¹⁵³, L Senter²³⁶, VW Setiawan¹²⁶, G Severi^{237,238,239,240}, P Sharma²⁴¹, T Shelford¹⁷, N Siddiqui²⁴², LE Side²⁴³, W Sieh¹⁸², CF Singer²¹⁷, H Sobol²⁴⁴, H Song⁴, MC Southey²⁴⁵, AB Spurdle¹¹, Z Stadler²³⁰, D Steinemann²⁴⁶, D Stoppa-Lyonnet⁵⁶, LE Sucheston-Campbell²⁴⁷, G Sukiennicki¹²³, R Sutphen²⁴⁸, C Sutter²⁴⁹, AJ Swerdlow^{153,250}, CI Szabo²⁵¹, L Szafron⁸⁰, YY Tan²¹⁷, JA Taylor²³⁵, M-K Tea²¹⁷, MR Teixeira²⁵², S-H Teo^{253,254}, KL Terry^{75,255}, PJ Thompson²⁵⁶, LCV Thomsen^{40,41}, DL Thull²⁵⁷, L Tihomirova²⁵⁸, AV Tinker²⁵⁹, M Tischkowitz^{105,260}, S Tognazzo¹⁹⁵, A Ewart Toland²⁶¹, A Tone³⁸, B Trabert⁴², RC Travis²⁶², A Trichopoulou^{263,264}, N Tung²⁶⁵, SS. Tworoger^{221,255}, AM van Altena²⁶⁶, D Van Den Berg¹²⁶, AH van der Hout²⁶⁷, RB van der Luijt²⁶⁸, M Van Heetvelde⁷², E Van Nieuwenhuysen²⁶⁹, EJ van Rensburg⁸⁸, A Vanderstichele²⁶⁹, R Varon-Mateeva²⁷⁰, V Ana^{271,272}, D Velez Edwards²⁷³, I Vergote²⁶⁹, RA Vierkant⁶, J Vijai¹⁹⁸, A Vratimos¹⁰⁴, L Walker²⁷⁴, C Walsh⁵, D Wand²⁷⁵, S Wang-Gohrke²⁷⁶, B Wappenschmidt¹²⁵, PM Webb²⁷⁷, CR Weinberg²⁷⁸, JN Weitzel²⁷⁹, N Wentzensen⁴², AS Whittemore^{182,280}, JT Wijnen²⁸¹, LR Wilkens¹⁶⁸, A Wolk¹²⁷, M Woo¹⁴², X Wu¹³¹, AH Wu¹²⁶, H Yang⁴², D Yannoukakos¹⁰⁴, A Ziogas²⁸², KK Zorn²⁵⁷, SA Narod²⁸³, DF Easton^{3,4}, CI Amos²⁸⁴, JM Schildkraut^{28,5}, SJ Ramus^{286,287}, L Ottini²⁸⁸, MT Goodman^{256,289}, SK Park^{290,291,292}, LE Kelemen²⁹³, HA Risch²⁹⁴, M Thomassen¹⁵⁸, K Offit¹⁹⁸, J Simard¹³, R Katharina Schmutzler¹²⁵, D Hazelett²⁹⁵, AN Monteiro⁸, FJ Couch⁷⁶, A Berchuck⁷, G Chenevix-Trench¹¹, EL Goode⁹, TA. Sellers⁸, SA Gayther¹⁰, AC Antoniou^{3,4}, and PDP Pharoah^{3,4}

1. Departments of Cancer Epidemiology and Gynecologic Oncology, Moffitt Cancer Center, Tampa, FL, USA
2. Wellcome Trust Sanger Institute, Wellcome Genome Campus, Hinxton, Cambridgeshire CB10 1SA, UK
3. Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Strangeways Research Laboratory, Worts Causeway, Cambridge, UK
4. Department of Oncology, Centre for Cancer Genetic Epidemiology, University of Cambridge, Strangeways Research Laboratory, Cambridge, UK
5. Women's Cancer Program at the Samuel Oschin Comprehensive Cancer Institute, CedarsSinai Medical Center, 8700 Beverly Boulevard, Suite 290W, Los Angeles, CA, USA
6. Department of Health Science Research, Division of Biomedical Statistics and Informatics, Mayo Clinic, Rochester, Minnesota, USA
7. Department of Obstetrics and Gynecology, Duke University Medical Center, Durham, North Carolina, USA
8. Department of Cancer Epidemiology, Moffitt Cancer Center, Tampa, FL, USA
9. Department of Health Science Research, Division of Epidemiology, Mayo Clinic, Rochester, Minnesota, USA
10. Center for Bioinformatics and Functional Genomics, Samuel Oschin Comprehensive Cancer Institute, Cedars-Sinai Medical Center, Los Angeles, California, USA
11. Department of Genetics and Computational Biology, QIMR Berghofer Medical Research Institute, Brisbane, QLD, Australia
12. Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK
13. Genomics Center, Centre Hospitalier Universitaire de Québec Research Center and Laval University, 2705 Laurier Boulevard, Quebec City (Quebec), Canada
14. Department of Clinical Genetics, Academic Medical Center, Amsterdam, The Netherlands
15. Radboud university medical center, Radboud Institute for Health Sciences, Department for Health Evidence, Nijmegen, The Netherlands
16. Netherlands Comprehensive Cancer Organisation, Utrecht, The Netherlands
17. Center for Inherited Disease Research, Institute of Genetic Medicine, Johns Hopkins University School of Medicine, Baltimore, MD 21224
18. Yorkshire Regional Genetics Service, Chapel Allerton Hospital, Leeds, UK
19. Lunenfeld-Tanenbaum Research Institute, Mount Sinai Hospital, Toronto, Ontario M5G 1X5, Departments of Molecular Genetics and Laboratory Medicine and Pathobiology, University of Toronto, Ontario, Canada
20. Department of Epidemiology, Director of Genetic Epidemiology Research Institute, UCI Center for Cancer Genetics Research & Prevention, School of Medicine, University of California Irvine, Irvine, California, USA
21. N.N. Alexandrov National Cancer Centre of Belarus, Minsk, Belarus
22. A list of members and affiliations appears in the Supplementary note
23. "Agi Anargiri" Cancer Hospital, Athens, Greece
24. Department of Gynaecology and Obstetrics, University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University Kiel, Germany
25. Department of Breast Medical Oncology and Clinical Cancer Genetics Program, University Of Texas MD Andersson Cancer Center, 1515 Pressler Street, CBP 5, Houston, TX, USA

26. Department of Oncology and Pathology, Karolinska University Hospital and Karolinska Institutet, Stockholm, Sweden
27. Unit of Medical Genetics, Department of Preventive and Predictive Medicine, Fondazione IRCCS (Istituto Di Ricovero e Cura a Carattere Scientifico) Istituto Nazionale Tumori (INT), Via Giacomo Venezian 1, 20133 Milan, Italy
28. Department of Medical Oncology, University Hospital, Vall d'Hebron, Barcelona, Spain
29. Gynaecology Unit, The Royal Marsden Hospital, London, UK
30. Bâtiment Cheney D, Centre Léon Bérard, 28 rue Laënnec, Lyon, France
31. Laboratory of Cell Biology, Department of Pathology, hus 9, Landspítali-LSH v/Hringbraut, 101 Reykjavik, Iceland
32. BMC (Biomedical Centre), Faculty of Medicine, University of Iceland, Vatnsmýrarvegi 16, 101 Reykjavik, Iceland
33. Department of Gynecologic Oncology, Oregon Health & Science University, Portland, OR, USA; Knight Cancer Institute, Portland, OR, USA
34. Universtiy Hospital Erlangen, Department of Gynecology and Obstetrics, FriedrichAlexander-University Erlangen-Nuremberg, Comprehensive Cancer Center Erlangen Nuremberg, Universitaetsstrasse 21-23, 91054 Erlangen, Germany
35. Division of Epidemiology, Department of Medicine, Vanderbilt Epidemiology Center, Institute for Medicine and Public Health, Vanderbilt University Medical Center, VanderbiltIngram Cancer Center, Nashville, TN, USA
36. Human Genetics Group, Spanish National Cancer Centre (CNIO), and Biomedical Network on Rare Diseases (CIBERER), Madrid, Spain
37. Institute of Biochemistry and Genetics, Ufa Science Center, Russian Academy of Sciences, Ufa, Bashkortostan, Russia
38. Division of Gynecologic Oncology, Princess Margaret Hospital, University Health Network, Toronto, Ontario, Canada
39. Department of Medicine, Massachusetts General Hospital, Boston, USA
40. Department of Gynecology and Obstetrics, Haukeland University Hospital, Bergen, Norway
41. Centre for Cancer Biomarkers, Department of Clinical Science, University of Bergen, Bergen, Norway
42. Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda, MD, USA
43. Clinical Cancer Genetics, for the City of Hope Clinical Cancer Genetics Community Research Network, Duarte, CA, USA
44. Department of Clinical Genetics, Maastricht University Medical Center, Maastricht, The Netherlands
45. Radiation Oncology Research Unit, Hannover Medical School, Hannover, Germany
46. Department of Clinical Genetics, Vejle Hospital, Vejle, Denmark
47. Division of Cancer Prevention and Genetics, Istituto Europeo di Oncologia (IEO), via Ripamonti 435, 20141 Milan, Italy
48. Department of Oncology, Clinical Sciences, Lund University and Skåne University Hospital, Lund, Sweden
49. Department of Medicine, Abramson Cancer Center, Perelman School of Medicine at The University of Pennsylvania, Philadelphia, PA, USA
50. Cancer Research UK (CRUK) Cambridge Institute, University of Cambridge
51. Department of Clinical Genetics, Royal Devon and Exeter Hospital, Exeter, UK
52. Department of Cancer Epidemiology, University Hospital, Lund, Lund University,

Lund, Sweden

53. Canada's Michael Smith Genome Sciences Centre, BC Cancer Agency, Vancouver, BC, Canada
54. Cancer Epidemiology Centre, Cancer Council Victoria, Melbourne, VIC, Australia
55. Genetic Counseling Unit, Hereditary Cancer Program, IDIBGI (Institut d'Investigació Biomèdica de Girona), Catalan Institute of Oncology. Av. França s/n. 1707 Girona, Spain
56. Service de Génétique Oncologique, Institut Curie, 26, rue d'Ulm, Paris Cedex 05, France
57. Department of Pathology, University of Helsinki and Helsinki University Hospital, Helsinki, Finland
58. Department of Medicine, Huntsman Cancer Institute, 2000 Circle of Hope, Salt Lake City, UT 84112, USA
59. Molecular Oncology Laboratory, Hospital Clinico San Carlos, IdISSC (Instituto de Investigación Sanitaria del Hospital Clínico San Carlos), Martin Lagos s/n, Madrid, Spain
60. Section of Genetic Oncology, Dept. of Laboratory Medicine, University and University Hospital of Pisa, Pisa Italy
61. Cancer Genetics Laboratory, Research Division, Peter MacCallum Cancer Centre, Melbourne, VIC, Australia
62. Department of Pathology, University of Melbourne, Parkville, VIC, Australia
63. Cancer Pathology & Prevention, Division of Cancer Prevention and Population Sciences, Roswell Park Cancer Institute, Buffalo, NY
64. Department of Obstetrics and Gynecology, John A. Burns School of Medicine, University of Hawaii, Honolulu, Hawaii, US
65. University of California, San Francisco, 1600 Divisadero Street, C415, San Francisco, CA 94143 – 1714, USA
66. Division of Cancer Epidemiology, German Cancer Research Center (DKFZ), Heidelberg, Germany
67. University Cancer Center Hamburg (UCCH), University Medical Center Hamburg Eppendorf, Hamburg, Germany
68. Centre for Cancer Research, The Westmead Institute for Medical Research, The University of Sydney, Sydney, New South Wales, Australia
69. Department of Gynaecological Oncology, Westmead Hospital, Sydney, New South Wales, Australia
70. Unité de Recherche en Santé des Populations, Centre des Maladies du Sein DeschênesFabia, Centre de Recherche FRSQ du Centre Hospitalier Affilié Universitaire de Québec, Québec, QC, Canada
71. Departments of Pediatrics and Medicine, Columbia University, New York, NY, USA
72. Center for Medical Genetics, Ghent University, Ghent, Belgium
73. Division of Epidemiology, Biostatistics and Preventative Medicine, Department of Internal Medicine, University of New Mexico, Albuquerque, New Mexico, USA
74. Sheffield Clinical Genetics Service, Sheffield Children's Hospital, Sheffield, UK
75. Obstetrics and Gynecology Epidemiology Center, Brigham and Women's Hospital and Harvard Medical School, Boston, MA, USA
76. Department of Laboratory Medicine and Pathology, Division of Experimental Pathology, Mayo Clinic, Rochester, MN, USA
77. Social & Scientific Systems, Inc. Durham, NC 27703, USA
78. Department of Clinical Genetics, Fox Chase Cancer Center, 333 Cottman

Avenue, Philadelphia, PA 19111, USA

79. Department of Genetics and Fundamental Medicine, Bashkir State University, Ufa, Russia

80. Department of Pathology and Laboratory Diagnostics, the Maria Sklodowska-Curie Memorial Cancer Center and Institute of Oncology, Warsaw, Poland

81. Gynecology Service, Department of Surgery, Memorial Sloan Kettering Cancer Center, New York, NY, USA

82. Department of Clinical Genetics, South Glasgow University Hospitals, Glasgow, UK

83. Unité d'oncogénétique, ICO-Centre René Gauducheau, Boulevard Jacques Monod, 44805 Nantes Saint Herblain Cedex, France

84. Oncogenetics Group, Vall d'Hebron Institute of Oncology (VHIO), Barcelona, Spain

85. Clinical and Molecular Genetics Area, Vall d'Hebron University Hospital. Barcelona, Spain

86. Department of Population Sciences, Beckman Research Institute of City of Hope, Duarte, CA, USA

87. Department of Epidemiology, The Geisel School of Medicine – at Dartmouth, Hanover, New Hampshire, USA

88. Cancer Genetics Laboratory, Department of Genetics, University of Pretoria, Pretoria, South Africa

89. Gynaecology Research Unit, Hannover Medical School, Hannover, Germany

90. Nutrition and Metabolism Section, International Agency for Research on Cancer (IARCWHO), Lyon, France

91. Institute of Biology and Molecular Genetics, Universidad de Valladolid (IBGM-UVA), Valladolid, Spain

92. Department of Gynecology, Jena University Hospital – Friedrich Schiller University, Jena, Germany

93. Institute of Human Genetics, Münster, Germany

94. University of Southampton Faculty of Medicine, Southampton University Hospitals NHS Trust, Southampton, UK

95. Oncogenetics Team, The Institute of Cancer Research and Royal Marsden NHS Foundation Trust, Sutton, UK

96. Department of Oncology, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark

97. Institute of Human Genetics, Friedrich-Alexander-University Erlangen-Nuremberg, Erlangen, Germany

98. Department of Biostatistics & Bioinformatics, Roswell park Institute, Buffalo, NY, USA

99. Institute for Medical Informatics, Statistics and Epidemiology, University of Leipzig, Leipzig, Germany

100. Genomic Medicine, Manchester Academic Health Sciences Centre, University of Manchester, Central Manchester University Hospitals NHS Foundation Trust, Manchester, UK

101. University of California at Los Angeles, David Geffen School of Medicine, Department of Medicine, Division of Hematology and Oncology, Los Angeles, CA, USA

102. Laboratoire de Génétique Chromosomique, Hôtel Dieu Centre Hospitalier, Chambéry, France

103. Department of Surgery and Cancer, Imperial College London, London, UK

104. Molecular Diagnostics Laboratory, INRASTES, National Centre for Scientific Research “Demokritos”, Aghia Paraskevi Attikis, Athens, Greece
105. Program in Cancer Genetics, Departments of Human Genetics and Oncology, McGill University, Montreal, Quebec, Canada
106. Department of Medical Oncology, Papageorgiou, Hospital, Aristotle University of Thessaloniki School of Medicine, Thessaloniki, Greece
107. Biostatistics and Informatics Shared Resource, University of Kansas Medical Center, Kansas City, KS, USA
108. Harvard TH Chan School of Public Health and Dana Farber Cancer Institute, 1101 Dana Building, 450 Brookline Ave, Boston, MA 02215, USA
109. The Susanne Levy Gertner Oncogenetics Unit, Institute of Human Genetics, Chaim Sheba Medical Center, Ramat Gan 52621, and Sackler Faculty of Medicine, Tel Aviv University, Ramat Aviv 69978, Israel
110. UCLA Schools of Medicine and Public Health, Division of Cancer Prevention and Control Research, Jonsson Comprehensive Cancer Center, Los Angeles, CA, USA
111. Cancer Risk and Prevention Clinic, Dana Farber Cancer Institute, Boston, MA, USA
112. Centre of Familial Breast and Ovarian Cancer, Department of Medical Genetics, Institute of Human Genetics, University Würzburg, Germany
113. Women’s Cancer, Institute for Women’s Health, University College London, London, United Kingdom
114. Department of Clinical Genetics, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark
115. Centre for Epidemiology and Biostatistics, School of Population and Global Health, University of Melbourne, Australia
116. Department of Epidemiology and Preventive Medicine, Monash University, Melbourne, VIC, Australia
117. The Beatson West of Scotland Cancer Centre, Glasgow, UK
118. Ontario Cancer Genetics Network: Samuel Lunenfeld Research Institute, Mount Sinai Hospital, Toronto, Ontario M5G 1X5
119. Department of Pathology and Laboratory Medicine, University of Kansas Medical Center, Kansas City, KS, USA
120. Department of Dermatology, University of Utah School of Medicine, Salt Lake City, Utah, USA
121. Department of Medicine, Royal Marsden Hospital, London, UK
122. Clinical Genetics Branch, DCEG, NCI, NIH, 9609 Medical Center Drive, Room 6E-454, Bethesda, MD, USA
123. Department of Genetics and Pathology, Pomeranian Medical University, Szczecin, Poland
124. Keck School of Medicine, and Norris Comprehensive Cancer Center, University of Southern California, Los Angeles, CA
125. Center for Familial Breast and Ovarian Cancer, Center for Integrated Oncology (CIO) and Center for Molecular Medicine Cologne (CMMC), University Hospital Cologne, Cologne, Germany
126. Department of Preventive Medicine, Keck School of Medicine, University of Southern California Norris Comprehensive Cancer Center, Los Angeles, CA, USA
127. Karolinska Institutet, Department of Environmental Medicine, Division of Nutritional Epidemiology, SE-171 77 STOCKHOLM, Sweden
128. Molecular Genetics of Breast Cancer, Deutsches Krebsforschungszentrum

- (DKFZ), Heidelberg, Germany
129. Center for Genomic Medicine, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark
 130. Institute of Genetic Medicine, Centre for Life, Newcastle Upon Tyne Hospitals NHS Trust, Newcastle upon Tyne, UK
 131. Department of Epidemiology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA
 132. Clinics of Obstetrics and Gynaecology, Hannover Medical School, Hannover, Germany
 133. Medical Genetics Unit, St George's, University of London, UK
 134. Department of Gynecology, Rigshospitalet, University of Copenhagen, Copenhagen, Denmark
 135. Department of Virus, Lifestyle and Genes, Danish Cancer Society Research Center, Copenhagen, Denmark
 136. Molecular Unit, Department of Pathology, Herlev Hospital, University of Copenhagen, Copenhagen, Denmark
 137. Family Cancer Clinic, Netherlands Cancer Institute, Amsterdam, The Netherlands
 138. Department of Medical Oncology, Family Cancer Clinic, Erasmus University Medical Center, Rotterdam, The Netherlands
 139. Center For Immunotherapy, Roswell Park Cancer Institute, Buffalo, NY, USA
 140. Center for Medical Genetics, NorthShore University HealthSystem, University of Chicago Pritzker School of Medicine, 1000 Central Street, Suite 620, Evanston, IL 60201, US
 141. Program in Genetic Epidemiology and Statistical Genetics, Department of Epidemiology, The Harvard T.H. Chan School of Public Health, 677 Huntington Avenue, Boston, MA, 02115, USA
 142. British Columbia's Ovarian Cancer Research (OVCARE) Program, Vancouver General Hospital, BC Cancer Agency and University of British Columbia; Departments of Pathology and Laboratory Medicine, Obstetrics and Gynaecology and Molecular Oncology, Vancouver, British Columbia, CANADA
 143. N.N. Petrov Institute of Oncology, St.Petersburg, Russia
 144. Lombardi Comprehensive Cancer Center, Georgetown University, 3800 Reservoir Road NW, Washington, DC, USA
 145. Department of Statistical Science, Duke University, Durham, North Carolina, USA
 146. Clinical Genetics, Guy's and St. Thomas' NHS Foundation Trust, London, UK
 147. Familial Cancer Centre, Peter MacCallum Cancer Centre, Locked Bag 1, A'Beckett Street, Melbourne, VIC 8006 AUSTRALIA; Sir Peter MacCallum Dept of Oncology, University of Melbourne, VIC 3010
 148. Vilnius University Hospital Santariskiu Clinics, Hematology, Oncology and Transfusion Medicine Center, Department of Molecular and Regenerative Medicine, Vilnius, Lithuania
 149. State Research Institute Centre for Innovative Medicine, Vilnius, Lithuania
 150. Department of Obstetrics and Gynecology Lund University Hospital, Lund Sweden
 151. Department of Clinical Genetics, Aarhus University Hospital, Brendstrupgaardsvej 21C, Aarhus N, Denmark
 152. Department of Epidemiology, Cancer Prevention Institute of California, Fremont, California, USA

153. Division of Genetics and Epidemiology, Institute of Cancer Research, London, UK
154. Department of Gynaecology and Obstetrics, University Hospital Carl Gustav Carus, Technical University Dresden, Germany
155. Department of Obstetrics and Gynecology, University of Helsinki and Helsinki University Central Hospital, Helsinki, HUS, Finland
156. Department of Surgery, Breast Care Center, Daerim St. Mary's Hospital, 657 Siheungdaero, Yeongdeungpo-gu, Seoul, 150-822, Korea
157. Department of Pathology, University of Calgary, Calgary, Alberta, Canada
158. Department of Clinical Genetics, Odense University Hospital, Odense C, Denmark
159. The Hong Kong Hereditary Breast Cancer Family Registry, Hong Kong
160. Department of Surgery, The University of Hong Kong, Hong Kong
161. Cancer Genetics Center and Department of Surgery, Hong Kong Sanatorium and Hospital, Hong Kong
162. Vesalius Research Center, VIB, Leuven, Belgium
163. Laboratory for Translational Genetics, Department of Oncology, KULeuven, Belgium
164. Public Health Division of Gipuzkoa, Regional Government of the Basque Country, Spain
165. CIBER of Epidemiology and Public Health (CIBERESP), Spain
166. Molecular Diagnostic Unit, Hereditary Cancer Program, IDIBELL-Catalan Institute of Oncology, Barcelona, Spain
167. Cancer Control Research, BC Cancer Agency, Vancouver, BC, Canada
168. Cancer Epidemiology Program, University of Hawaii Cancer Center, Hawaii, USA
169. Department of Surgery, Ulsan College of Medicine and Asan Medical Center, Seoul, Korea
170. Department of Gynecological Oncology, Roswell Park Cancer Institute, Buffalo, NY, USA
171. Département de Génétique, Centre Hospitalier Universitaire de Grenoble, BP 217, Grenoble Cedex 9, France
172. Institut Curie, PSL Research University and Inserm, U900, Paris, France; Mines Paris Tech, Fontainebleau, France
173. College of Pharmacy and Health Sciences, Texas Southern University, Houston, Texas, USA
174. Cancer Center Wolfsburg, Clinics of Gynaecology, Wolfsburg, Germany
175. Department of Health Sciences Research, Division of Epidemiology, Mayo Clinic, Rochester, MN, USA
176. Division of Epidemiology, Department of Medicine, Vanderbilt Epidemiology Center and Vanderbilt-Ingram Cancer Center, Vanderbilt University School of Medicine, Nashville, TN, USA
177. Department of Cancer Epidemiology and Prevention, Maria Sklodowska-Curie Memorial Cancer Center and Institute of Oncology, Warsaw, Poland
178. Department of Gynecologic Oncology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA
179. The Juliane Marie Centre, Department of Gynecology, Rigshospitalet, University of Copenhagen, Copenhagen, Denmark
180. INSERM U1052, CNRS UMR5286, Université Lyon 1, Centre de Recherche en Cancérologie de Lyon, Centre Léon Bérard, Lyon, France

181. Ovarian Cancer Research (OVCARE) Program – Gynecologic Tissue Bank, Vancouver General Hospital and BC Cancer Agency, Vancouver, British Columbia CANADA
182. Department of Health Research and Policy – Epidemiology, Stanford University School of Medicine, Stanford, CA, USA
183. Public Health Ontario, Toronto, ON, Canada
184. Institute of Cancer Sciences, University of Glasgow, Wolfson Wohl Cancer Research Centre, Beatson Institute for Cancer Research, Glasgow, UK
185. Department of Clinical Genetics, VU University Medical Centre, P.O. Box 7057, 1007 MB Amsterdam, the Netherlands
186. Department of Gynaecology and Obstetrics, Division of Tumor Genetics, Klinikum rechts der Isar, Technical University of Munich, Munich, Germany
187. Department of Human Genetics, Radboud University Medical Centre, Nijmegen, The Netherlands
188. Department of Epidemiology and Biostatistics, School of Public Health, Imperial College London, London, W2 1PG, UK
189. Sir Peter MacCallum Department of Oncology, University of Melbourne, Parkville, VIC, Australia
190. Division of Gynecologic Oncology, Department of Obstetrics, Gynecology and Reproductive Sciences, University of Pittsburgh School of Medicine, Pittsburgh, Pennsylvania, USA
191. Department of Epidemiology, University of Pittsburgh Graduate School of Public Health, Pittsburgh, Pennsylvania, USA
192. Ovarian Cancer Center of Excellence, Womens Cancer Research Program, MageeWomens Research Institute and University of Pittsburgh Cancer Institute, Pittsburgh, Pennsylvania, USA
193. Department of Obstetrics and Gynecology, Oregon Health and Science University, Portland, OR, USA
194. Knight Cancer Institute, Oregon Health & Science University, Portland, Oregon, USA
195. Immunology and Molecular Oncology Unit, Veneto Institute of Oncology IOV – IRCCS, Via Gattamelata 64, Padua, Italy
196. Department of Laboratory Medicine and Pathobiology, University of Toronto, Toronto, ON, Canada
197. Department of Laboratory Medicine, and the Keenan Research Centre of the Li Ka Shing Knowledge Institute, St Michael's Hospital, Toronto, ON, Canada
198. Clinical Genetics Research Laboratory, Department of Medicine, Memorial Sloan Kettering Cancer Center, 1275 York Avenue, New York, NY 10044, USA
199. Department of Pathology, Rigshospitalet, University of Copenhagen, Denmark
200. The University of Texas School of Public Health, Houston, TX, USA
201. Department of Gynaecology and Obstetrics, University Hospital Düsseldorf, HeinrichHeine University Düsseldorf, Germany
202. Invitae Corporation and University of Southern California, San Francisco, 513 Parnassus Ave., HSE 901E, San Francisco, CA. 94143 – 0794
203. Department of Molecular Genetics, National Institute of Oncology, Budapest, Hungary
204. Center for Clinical Cancer Genetics and Global Health, University of Chicago Medical Center, 5841 South Maryland Avenue, MC 2115 Chicago, IL, USA
205. Oncology, Clinical Sciences in Lund, Lund University, 221 85 Lund, Sweden
206. The Ohio State University and the James Cancer Center, Columbus, Ohio, USA

207. West Midlands Regional Genetics Service, Birmingham Women's Hospital Healthcare NHS Trust, Edgbaston, Birmingham, UK
208. Julius Center for Health Sciences and Primary Care, UMC Utrecht, Utrecht, the Netherlands
209. The Breast Cancer Now Toby Robins Research Centre, The Institute of Cancer Research, 237 Fulham Road, London SW3 6JB, UK
210. Molecular and Nutritional Epidemiology Unit, Cancer Research and Prevention Institute ISPO, Florence, Italy
211. Unit of Medical Genetics, Department of Biomedical, Experimental and Clinical Sciences, University of Florence, Florence, Italy
212. Cancer Research UK Clinical Trials Unit, Institute of Cancer Sciences, University of Glasgow, UK
213. Department of Epidemiology, University of Michigan School of Public Health, Ann Arbor, Michigan, USA
214. Section of Molecular Diagnostics, Clinical Biochemistry, Aalborg University Hospital, Aalborg, Denmark
215. Department of Genetics, Portuguese Oncology Institute, Rua Dr. António Bernardino de Almeida, 4200-072 Porto, Portugal
216. IFOM, The FIRC (Italian Foundation for Cancer Research) Institute of Molecular Oncology, c/o IFOM-IEO campus, via Adamello 16 20139 Milan, Italy
217. Dept of OB/GYN, Medical University of Vienna and Comprehensive Cancer Center, Vienna, Austria, Waehringer Guertel 18-20, A 1090 Vienna, Austria
218. Division of Cancer Medicine, Peter MacCallum Cancer Centre, Locked Bag 1, A'Beckett St, East Melbourne, victoria 8006, Australia
219. NRG Oncology, Statistics and Data Management Center, Roswell Park Cancer Institute, Elm St & Carlton St, Buffalo, NY 14263, USA
220. Department of Epidemiology and Biostatistics, Memorial Sloan Kettering Cancer Center, New York, NY, USA
221. Channing Division of Network Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, MA, USA
222. South East of Scotland Regional Genetics Service, Western General Hospital, Edinburgh, UK
223. Service de Génétique Clinique Chromosomique et Moléculaire, Centre Hospitalier Universitaire de St Etienne, St Etienne, France
224. Translational Research Laboratory, IDIBELL (Bellvitge Biomedical Research Institute), Catalan Institute of Oncology, Barcelona, Spain
225. Unité d'Oncogénétique, CHU Arnaud de Villeneuve, Montpellier, France
226. Unit of "Predictive Medicine: Molecular Bases of Genetic Risk", Department of Experimental Oncology, Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy
227. Department of Clinical Genetics, Karolinska University Hospital, Stockholm, Sweden
228. Clalit National Israeli Cancer Control Center and Department of Community Medicine and Epidemiology, Carmel Medical Center and B. Rappaport Faculty of Medicine, Haifa, Israel
229. Brigham and Women's Hospital, Dana-Farber Cancer Institute, Boston, MA USA
230. Clinical Genetics Service, Department of Medicine, Memorial Sloan Kettering Cancer Center, New York, NY, USA
231. Division of Gynecologic Oncology, NorthShore University HealthSystem,

University of Chicago, Evanston, IL, USA

232. Hereditary Endocrine Cancer group, Spanish National Cancer Research Center (CNIO), and Biomedical Network on Rare Diseases (CIBERER), Madrid, Spain

233. Department of Epidemiology, Netherlands Cancer Institute, Amsterdam, The Netherlands

234. Program in Epidemiology, Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, WA, USA

235. Epidemiology Branch, Division of Intramural Research, National Institute of Environmental Health Sciences, NIH, Research Triangle Park, NC, USA

236. Clinical Cancer Genetics Program, Division of Human Genetics, Department of Internal Medicine, The Comprehensive Cancer Center, The Ohio State University, Columbus, USA

237. Université Paris-Saclay, Université Paris-Sud, UVSQ, CESP, INSERM, Villejuif, France

238. Gustave Roussy, F-94805, Villejuif, France

239. Human Genetics Foundation (HuGeF), Torino, Italy

240. Cancer Council Victoria and University of Melbourne, Australia

241. Department of Hematology and Oncology, University of Kansas Medical Center, Kansas City, KS, USA

242. Department of Gynaecological Oncology, Glasgow Royal Infirmary, Glasgow, UK

243. North East Thames Regional Genetics Service, Great Ormond Street Hospital for Children NHS Trust, London, UK

244. Département Oncologie Génétique, Prévention et Dépistage, INSERM CIC-P9502, Institut Paoli-Calmettes/Université d'Aix-Marseille II, Marseille, France

245. Genetic Epidemiology Laboratory, Department of Pathology, University of Melbourne, Parkville, VIC, Australia

246. Institute of Human Genetics, Hannover Medical School, Hannover, Germany

247. Department of Cancer Prevention and Control, Roswell Park Cancer Institute, Buffalo, NY, USA

248. Epidemiology Center, College of Medicine, University of South Florida, Tampa, Florida, USA

249. Institute of Human Genetics, Department of Human Genetics, University Hospital Heidelberg, Germany

250. Division of Breast Cancer Research, The Institute of Cancer Research, London, UK

251. National Human Genome Research Institute, National Institutes of Health, Bethesda, MD, USA

252. Department of Genetics, Portuguese Oncology Institute, Rua Dr. António Bernardino de Almeida, 4200-072 Porto, Portugal and Biomedical Sciences Institute (ICBAS), University of Porto, Porto, Portugal

253. Cancer Research Initiatives Foundation, Sime Darby Medical Centre, Subang Jaya, Malaysia

254. University Malaya Cancer Research Institute, Faculty of Medicine, University Malaya Medical Centre, University Malaya, Kuala Lumpur, Malaysia

255. Department of Epidemiology, Harvard T. Chan School of Public Health, Boston, MA, USA

256. Cancer Prevention and Control, Samuel Oschin Comprehensive Cancer Institute, CedarsSinai Medical Center, Los Angeles, CA, USA

257. Magee-Womens Hospital of UPMC, University of Pittsburgh School of Medicine, Pittsburgh, PA, USA

258. Latvian Biomedical Research and Study Centre. Ratsupites str 1, Riga, Latvia
259. Ovarian Cancer Research (OVCARE) Program – Cheryl Brown Ovarian Cancer Outcomes Unit (CBOCOU), BC Cancer Agency, Vancouver, British Columbia
CANADA
260. Department of Medical Genetics, Box 134, Level 6 Addenbrooke’s Treatment Centre, Addenbrooke’s Hospital, Hills Road, Cambridge CB2 0QQ, UK
261. Division of Human Cancer Genetics, Departments of Internal Medicine and Molecular Virology, Immunology and Medical Genetics, Comprehensive Cancer Center, The Ohio State University, Columbus, OH, USA
262. Cancer Epidemiology Unit, Nuffield Department of Population Health, University of Oxford, Oxford, UK
263. Hellenic Health Foundation, Athens, Greece
264. WHO Collaborating Center for Nutrition and Health, Unit of Nutritional Epidemiology and Nutrition in Public Health, Dept. of Hygiene, Epidemiology and Medical Statistics, University of Athens Medical School, Greece
265. Department of Medical Oncology, Beth Israel Deaconess Medical Center, Boston, MA, USA
266. Division of Gynecologic Oncology, Department of Obstetrics and Gynaecology, Radboud University Medical Centre, Nijmegen, The Netherlands
267. Department of Genetics, University Medical Center, Groningen University, Groningen, The Netherlands
268. Department of Medical Genetics, University Medical Center Utrecht, Utrecht, The Netherlands
269. Division of Gynecologic Oncology, Department of Obstetrics and Gynaecology and Leuven Cancer Institute, University Hospitals Leuven, Leuven, Belgium
270. Institute of Human Genetics, Campus Virchow Klinikum, Charite Berlin, Germany
271. Fundación Pública Galega de Medicina Xenómica, Servizo Galego de Saúde (SERGAS), Instituto de Investigaciones Sanitarias (IDIS), Santiago de Compostela, Spain
272. Grupo de Medicina Xenómica, Centro de Investigación Biomédica en Red de Enfermedades Raras (CIBERER), Universidade de Santiago de Compostela (USC), Santiago de Compostela, Spain
273. Vanderbilt Epidemiology Center, Vanderbilt Genetics Institute, Department of Obstetrics and Gynecology, Vanderbilt University Medical Center, Nashville, TN, USA
274. Oxford Regional Genetics Service, Churchill Hospital, Oxford, UK
275. Institute of Human Genetics, University Hospital, Leipzig, Germany
276. Department of Obstetrics and Gynecology, University of Ulm, Ulm, Germany
277. Population Health Department, QIMR Berghofer Medical Research Institute, 300 Herston Road, Herston, QLD 4006, Australia
278. Biostatistics and Computational Biology Branch, Division of Intramural Research, National Institute of Environmental Health Sciences, NIH, Research Triangle Park, NC, USA
279. Clinical Cancer Genetics, City of Hope, Duarte, CA, USA
280. Department of Data Management Science- Stanford University School of Medicine, Stanford, CA, USA
281. Department of Human Genetics and Department of Clinical Genetics, Leiden University Medical Center, Leiden, The Netherlands
282. Department of Epidemiology, University of California Irvine, Irvine, CA, USA

283. Women's College Research Institute, University of Toronto, Toronto, ON, Canada
284. Center for Genomic Medicine, Department of Biomedical Data Science, Geisel School of Medicine at Dartmouth, Williamson Translational Research Building, Room HB 7261, Lebanon, NH 03756, USA
285. Department of Public Health Sciences, The University of Virginia, Charlottesville, VA, USA
286. School of Women's and Children's Health, Lowy Cancer Research Centre, The University of New South Wales UNSW Sydney NSW 2052 AUSTRALIA
287. The Kinghorn Cancer Centre, Garvan Institute of Medical Research, 384 Victoria Street, Darlinghurst NSW 2010, Australia
288. Department of Molecular Medicine, University La Sapienza, c/o Policlinico Umberto I, viale Regina Elena 324, 00161 Rome, Italy
289. Community and Population Health Research Institute, Department of Biomedical Sciences, Cedars-Sinai Medical Center, Los Angeles, California, USA
290. Department of Preventive Medicine, College of Medicine, Seoul National University, 103 Daehak-ro, Jongno-gu, Seoul 110-799, Korea
291. Seoul National University Cancer Research Institute, 103 Daehak-ro, Jongno-gu, Seoul 110-799, Korea
292. Department of Biomedical Science, Graduate School, Seoul National University, 103 Daehak-ro, Jongno-gu, Seoul 110-799, Korea
293. Department of Public Health Sciences, College of Medicine, Medical University of South Carolina, Charleston, SC 29425
294. Department of Chronic Disease Epidemiology, Yale School of Public Health, New Haven, CT, USA
295. Department of Biomedical Sciences, Cedars-Sinai Medical Center, Los Angeles, California, USA

The Breast Cancer Association Consortium

K Michailidou^{1,2}, S Lindström³, J Dennis¹, MK Bolla¹, Q Wang¹, R Keeman⁴, S Behrens⁵, H Anton-Culver⁶, KJ Aronson⁷, PL Auer^{8,9}, J Benitez^{10,11}, SE Bojesen^{12,13,14}, H Brenner^{15,16,17}, B Burwinkel^{18,19}, F Canzian²⁰, JE Castelao²¹, JY Choi^{22,23}, CL Clarke²⁴, NBCS Collaborators, A Cox²⁵, K Czene²⁶, MB Daly²⁷, P Devilee^{28,29}, T Dörk³⁰, M Dwek³¹, DM Eccles³², PA Fasching^{33,34}, L Fritschi³⁵, M Gago-Dominguez^{36,37}, JA García-Sáenz³⁸, GG Giles^{39,40}, MS Goldberg^{41,42}, DE Goldgar⁴³, P Guénel⁴⁴, CA Haiman⁴⁵, U Hamann⁴⁶, M Hartman^{47,48}, A Hollestelle⁴⁹, MJ Hoening⁴⁹, R Hoppe^{50,51}, JL Hopper⁴⁰, M Iwasaki⁵², A Jakubowska⁵³, EM John⁵⁴, R Kaaks⁵, D Kang^{22,23,55}, VN Kristensen^{56,57}, A Kwong^{58,59,60}, D Lambrechts^{61,62}, A Lindblom⁶³, J Lubinski⁵³, C Luccarini⁶⁴, A Mannermaa^{65,66,67}, S Margolin⁶⁸, K Matsuo^{69,70}, U Menon⁷¹, K Muir^{72,73}, SL Neuhausen⁷⁴, H Nevanlinna⁷⁵, OI Olopade⁷⁶, H Jernström⁷⁷, SK Park^{22,23,55}, P Peterlongo⁷⁸, J Peto⁷⁹, KA Phillips^{40,80,81,82}, D Plaseska-Karanfilska⁸³, R Prentice⁸, P Radice⁸⁴, G Rennert⁸⁵, A Romero⁸⁶, E Saloustros⁸⁷, DP Sandler⁸⁸, EJ Sawyer⁸⁹, RK Schmutzler^{90,91,92}, CY Shen^{93,94}, XO Shu⁹⁵, MC Southey^{39,96,97}, JJ Spinelli^{98,99}, J Stone^{40,100}, A Swerdlow^{101,102}, R Tamimi^{103,104,105}, JA Taylor^{88,106}, LR Teras¹⁰⁷, MB Terry¹⁰⁸, C Vachon¹⁰⁹, CR Weinberg¹¹⁰, C Wendt¹¹¹, R Winqvist^{112,113}, A Wolk¹¹⁴, AH Wu⁴⁵, W Zheng⁹⁵, E Ziv¹¹⁵, ABCTB Investigators*; kConFab/AOCS Investigators, AC Antoniou¹, IL

Andrulis^{116,117}, FJ Couch¹¹⁸, PDP Pharoah^{1,64}, J Chang-Claude^{5,119}, P Hall^{26,68}, DJ Hunter^{104,105}, RL Milne^{39,40}, M García-Closas¹²⁰, MK Schmidt^{4,121}, SJ Chanock¹²⁰, AM Dunning⁶⁴, G Chenevix-Trench¹²², J Simard¹²³, P Kraft^{104,105}, DF Easton^{1,64}

1 Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK

2 Department of Electron Microscopy/Molecular Pathology, The Cyprus Institute of Neurology and Genetics, Nicosia, Cyprus

3 Department of Epidemiology, University of Washington School of Public Health, Seattle, WA, USA

4 Division of Molecular Pathology, The Netherlands Cancer Institute - Antoni van Leeuwenhoek Hospital, Amsterdam, The Netherlands

5 Division of Cancer Epidemiology, German Cancer Research Center (DKFZ), Heidelberg, Germany

6 Department of Epidemiology, University of California Irvine, Irvine, CA, USA

7 Department of Public Health Sciences, and Cancer Research Institute, Queen's University, Kingston, ON, Canada

8 Cancer Prevention Program, Fred Hutchinson Cancer Research Center, Seattle, WA, USA

9 Zilber School of Public Health, University of Wisconsin-Milwaukee, Milwaukee, WI, USA

10 Human Cancer Genetics Program, Spanish National Cancer Research Centre, Madrid, Spain

11 Centro de Investigación en Red de Enfermedades Raras (CIBERER), Valencia, Spain

12 Copenhagen General Population Study, Herlev and Gentofte Hospital, Copenhagen University Hospital, Herlev, Denmark

13 Department of Clinical Biochemistry, Herlev and Gentofte Hospital, Copenhagen University Hospital, Herlev, Denmark

14 Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark

15 German Cancer Consortium (DKTK), German Cancer Research Center (DKFZ), Heidelberg, Germany

16 Division of Clinical Epidemiology and Aging Research, German Cancer Research Center (DKFZ), Heidelberg, Germany

17 Division of Preventive Oncology, German Cancer Research Center (DKFZ) and National Center for Tumor Diseases (NCT), Heidelberg, Germany

18 Molecular Epidemiology Group, C080, German Cancer Research Center (DKFZ), Heidelberg, Germany

19 Molecular Biology of Breast Cancer, University Womens Clinic Heidelberg, University of Heidelberg, Heidelberg, Germany

20 Genomic Epidemiology Group, German Cancer Research Center (DKFZ), Heidelberg, Germany

21 Oncology and Genetics Unit, Instituto de Investigacion Biomedica (IBI) Orense-PontevedraVigo, Xerencia de Xestion Integrada de Vigo-SERGAS, Vigo, Spain

22 Department of Biomedical Sciences, Seoul National University Graduate School, Seoul, Korea

- 23 Cancer Research Institute, Seoul National University, Seoul, Korea
- 24 Westmead Institute for Medical Research, University of Sydney, Sydney, Australia
- 25 Sheffield Institute for Nucleic Acids (SInFoNiA), Department of Oncology and Metabolism, University of Sheffield, Sheffield, UK
- 26 Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden
- 27 Department of Clinical Genetics, Fox Chase Cancer Center, Philadelphia, PA, USA
- 28 Department of Pathology, Leiden University Medical Center, Leiden, The Netherlands
- 29 Department of Human Genetics, Leiden University Medical Center, Leiden, The Netherlands
- 30 Gynaecology Research Unit, Hannover Medical School, Hannover, Germany
- 31 School of Life Sciences, University of Westminster, London, UK
- 32 Faculty of Medicine, University of Southampton, Southampton, UK
- 33 Institute of Human Genetics, University Hospital Erlangen, Friedrich-Alexander University Erlangen-Nuremberg, Comprehensive Cancer Center Erlangen-EMN, Erlangen, Germany
- 34 David Geffen School of Medicine, Department of Medicine Division of Hematology and Oncology, University of California at Los Angeles, Los Angeles, CA, USA
- 35 School of Public Health, Curtin University, Perth, Australia
- 36 Genomic Medicine Group, Galician Foundation of Genomic Medicine, Instituto de Investigación Sanitaria de Santiago de Compostela (IDIS), Complejo Hospitalario Universitario de Santiago, SERGAS, Santiago de Compostela, Spain
- 37 Moores Cancer Center, University of California San Diego, La Jolla, CA, USA
- 38 Medical Oncology Department, Hospital Clínico San Carlos, IdISSC (Centro Investigación Biomedica en Red), CIBERONC (Instituto de Investigación Sanitaria San Carlos), Madrid, Spain
- 39 Cancer Epidemiology Division, Cancer Council Victoria, Melbourne, Victoria, Australia
- 40 Centre for Epidemiology and Biostatistics, Melbourne School of Population and Global Health, The University of Melbourne, Melbourne, Australia
- 41 Department of Medicine, McGill University, Montréal, QC, Canada
- 42 Division of Clinical Epidemiology, Royal Victoria Hospital, McGill University, Montréal, QC, Canada
- 43 Department of Dermatology, Huntsman Cancer Institute, University of Utah School of Medicine, Salt Lake City, UT, USA
- 44 Cancer & Environment Group, Center for Research in Epidemiology and Population Health (CESP), INSERM, University Paris-Sud, University Paris-Saclay, Villejuif, France
- 45 Department of Preventive Medicine, Keck School of Medicine, University of Southern California, Los Angeles, CA, USA
- 46 Molecular Genetics of Breast Cancer, German Cancer Research Center (DKFZ), Heidelberg, Germany

47 Saw Swee Hock School of Public Health, National University of Singapore, Singapore, Singapore

48 Department of Surgery, National University Health System, Singapore, Singapore

49 Department of Medical Oncology, Family Cancer Clinic, Erasmus MC Cancer Institute, Rotterdam, The Netherlands

50 Dr. Margarete Fischer-Bosch-Institute of Clinical Pharmacology, Stuttgart, Germany

51 University of Tübingen, Tübingen, Germany

52 Division of Epidemiology, Center for Public Health Sciences, National Cancer Center, Tokyo, Japan

53 Department of Genetics and Pathology, Pomeranian Medical University, Szczecin, Poland

54 Department of Medicine, Division of Oncology, Stanford Cancer Institute, Stanford University School of Medicine, Stanford, CA, USA

55 Department of Preventive Medicine, Seoul National University College of Medicine, Seoul, Korea

56 Department of Cancer Genetics, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, Oslo, Norway

57 Institute of Clinical Medicine, Faculty of Medicine, University of Oslo, Oslo, Norway

58 Hong Kong Hereditary Breast Cancer Family Registry, Happy Valley, Hong Kong

59 Department of Surgery, The University of Hong Kong, Pok Fu Lam, Hong Kong

60 Department of Surgery, Hong Kong Sanatorium and Hospital, Happy Valley, Hong Kong

61 VIB Center for Cancer Biology, Leuven, Belgium

62 Laboratory for Translational Genetics, Department of Human Genetics, University of Leuven, Leuven, Belgium

63 Department of Molecular Medicine and Surgery, Karolinska Institutet, Stockholm, Sweden

64 Centre for Cancer Genetic Epidemiology, Department of Oncology, University of Cambridge, Cambridge, UK

65 Translational Cancer Research Area, University of Eastern Finland, Kuopio, Finland

66 Institute of Clinical Medicine, Pathology and Forensic Medicine, University of Eastern Finland, Kuopio, Finland

67 Imaging Center, Department of Clinical Pathology, Kuopio University Hospital, Kuopio, Finland

68 Department of Oncology, Södersjukhuset, Stockholm, Sweden

69 Division of Cancer Epidemiology and Prevention, Aichi Cancer Center Research Institute, Nagoya, Japan

70 Division of Cancer Epidemiology, Nagoya University Graduate School of Medicine, Nagoya, Japan

71 MRC Clinical Trials Unit at UCL, Institute of Clinical Trials & Methodology, University College London, London, UK

72 Division of Population Health, Health Services Research and Primary Care, School of Health Sciences, Faculty of Biology, Medicine and Health, The University

of Manchester, Manchester, UK
73 Division of Health Sciences, Warwick Medical School, University of Warwick, Coventry, UK
74 Department of Population Sciences, Beckman Research Institute of City of Hope, Duarte, CA, USA
75 Department of Obstetrics and Gynecology, Helsinki University Hospital, University of Helsinki, Helsinki, Finland
76 Center for Clinical Cancer Genetics, The University of Chicago, Chicago, IL, USA
78. Oncology, Clinical Sciences in Lund, Lund University, 221 85 Lund, Sweden
77 Oncology, Clinical Sciences in Lund, Lund University, 221 85 Lund, Sweden
78 Genome Diagnostics Program, IFOM - the FIRC (Italian Foundation for Cancer Research) Institute of Molecular Oncology, Milan, Italy
79 Department of Non-Communicable Disease Epidemiology, London School of Hygiene and Tropical Medicine, London, UK
80 Peter MacCallum Cancer Center, Melbourne, Australia
81 Sir Peter MacCallum Department of Oncology, The University of Melbourne, Melbourne, Australia
82 Department of Medicine, St Vincent's Hospital, The University of Melbourne, Fitzroy, Australia
83 Research Centre for Genetic Engineering and Biotechnology "Georgi D. Efremov", MASA, Skopje, Republic of North Macedonia
84 Unit of "Predictive Medicine: Molecular Bases of Genetic Risk", Department of Experimental Oncology, Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy
85 Clalit National Cancer Control Center, Carmel Medical Center and Technion Faculty of Medicine, Haifa, Israel
86 Medical Oncology Department, Hospital Universitario Puerta de Hierro, Madrid, Spain
87 Department of Oncology, University Hospital of Larissa, Larissa, Greece
88 Epidemiology Branch, National Institute of Environmental Health Sciences, NIH, Research Triangle Park, NC, USA
89 Research Oncology, Guy's Hospital, King's College London, London, UK
90 Center for Familial Breast and Ovarian Cancer, Faculty of Medicine and University Hospital Cologne, University of Cologne, Cologne, Germany
91 Center for Integrated Oncology (CIO), Faculty of Medicine and University Hospital Cologne, University of Cologne, Cologne, Germany
92 Center for Molecular Medicine Cologne (CMMC), Faculty of Medicine and University Hospital Cologne, University of Cologne, Cologne, Germany
93 Taiwan Biobank, Institute of Biomedical Sciences, Academia Sinica, Taipei, Taiwan
94 School of Public Health, China Medical University, Taichung, Taiwan
95 Division of Epidemiology, Department of Medicine, Vanderbilt Epidemiology Center, Vanderbilt-Ingram Cancer Center, Vanderbilt University School of Medicine, Nashville, TN, USA
96 Precision Medicine, School of Clinical Sciences at Monash Health, Monash University, Clayton, Victoria, Australia

97 Department of Clinical Pathology, The University of Melbourne, Melbourne, Victoria, Australia

98 Population Oncology, BC Cancer, Vancouver, BC, Canada

99 School of Population and Public Health, University of British Columbia, Vancouver, BC, Canada

100 The Curtin UWA Centre for Genetic Origins of Health and Disease, Curtin University and University of Western Australia, Perth, Australia

101 Division of Genetics and Epidemiology, The Institute of Cancer Research, London, UK

102 Division of Breast Cancer Research, The Institute of Cancer Research, London, UK

103 Channing Division of Network Medicine, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, MA, USA

104 Department of Epidemiology, Harvard T.H. Chan School of Public Health, Boston, MA, USA

105 Program in Genetic Epidemiology and Statistical Genetics, Harvard T.H. Chan School of Public Health, Boston, MA, USA

106 Epigenetic and Stem Cell Biology Laboratory, National Institute of Environmental Health Sciences, NIH, Research Triangle Park, NC, USA

107 Department of Population Science American Cancer Society Atlanta, GA, USA

108 Department of Epidemiology, Mailman School of Public Health, Columbia University, New York, NY, USA

109 Department of Health Sciences Research, Division of Epidemiology, Mayo Clinic, Rochester, MN, USA

110 Biostatistics and Computational Biology Branch, National Institute of Environmental Health Sciences, NIH, Research Triangle Park, NC, USA

111 Department of Clinical Science and Education, Södersjukhuset, Karolinska Institutet, Stockholm, Sweden

112 Laboratory of Cancer Genetics and Tumor Biology, Cancer and Translational Medicine Research Unit, Biocenter Oulu, University of Oulu, Oulu, Finland

113 Laboratory of Cancer Genetics and Tumor Biology, Northern Finland Laboratory Centre Oulu, Oulu, Finland

114 Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden

115 Department of Medicine, Institute for Human Genetics, UCSF Helen Diller Family Comprehensive Cancer Center, University of California San Francisco, San Francisco, CA, USA

116 Fred A. Litwin Center for Cancer Genetics, Lunenfeld-Tanenbaum Research Institute of Mount Sinai Hospital, Toronto, ON, Canada

117 Department of Molecular Genetics, University of Toronto, Toronto, ON, Canada

118 Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN, USA

119 Cancer Epidemiology Group, University Cancer Center Hamburg (UCCH), University Medical Center Hamburg-Eppendorf, Hamburg, Germany

120 Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institutes of Health, Department of Health and Human Services, Bethesda, MD, USA

121 Division of Psychosocial Research and Epidemiology, The Netherlands Cancer Institute - Antoni van Leeuwenhoek hospital, Amsterdam, The Netherlands

122 Department of Genetics and Computational Biology, QIMR Berghofer Medical Research Institute, Brisbane, Australia

123 Genomics Center, Centre Hospitalier Universitaire de Québec – Université Laval Research Center, Québec City, QC, Canada

***ABCTB Investigators:**

Christine Clarke (Westmead Institute for Medical Research, University of Sydney, NSW, Australia); Rosemary Balleine (Pathology West ICPMR, Westmead, NSW, Australia); Robert Baxter (Kolling Institute of Medical Research, University of Sydney, Royal North Shore Hospital, NSW, Australia); Stephen Braye (Pathology North, John Hunter Hospital, Newcastle, NSW, 2305, Australia); Jane Carpenter (Westmead Institute for Medical Research, University of Sydney); Jane Dahlstrom (Department of Anatomical Pathology, ACT Pathology, Canberra Hospital, ACT, Australia; ANU Medical School, Australian National University, ACT, Australia); John Forbes (Department of Surgical Oncology, Calvary Mater Newcastle Hospital, Australian New Zealand Breast Cancer Trials Group, and School of Medicine and Public Health, University of Newcastle, NSW, Australia); C Soon Lee (School of Science and Health, The University of Western Sydney, Sydney, Australia); Deborah Marsh (Hormones and Cancer Group, Kolling Institute of Medical Research, Royal North Shore Hospital, University of Sydney, NSW, Australia); Adrienne Morey (SydPath St Vincent's Hospital, Sydney, NSW, Australia); Nirmala Pathmanathan (Department of Tissue Pathology and Diagnostic Oncology, Pathology West; Westmead Breast Cancer Institute, Westmead Hospital, NSW, Australia); Mythily Sachchithananthan (Australian Breast Cancer Tissue Bank, Westmead Institute for Medical Research, University of Sydney, Sydney, New South Wales, Australia); Rodney Scott (Centre for Information Based Medicine, Hunter Medical Research Institute, NSW, 2305, Australia; Priority Research Centre for Cancer, School of Biomedical Sciences and Pharmacy, Faculty of Health, University of Newcastle, NSW, Australia); Peter Simpson (The University of Queensland: UQ Centre for Clinical Research and School of Medicine, QLD, Australia); Allan Spigelman (Hereditary Cancer Clinic, St Vincent's Hospital, The Kinghorn Cancer Centre, Sydney, New South Wales, 2010, Australia); Nicholas Wilcken (Crown Princess Mary Cancer Centre, Westmead Hospital, Westmead, Australia; Sydney Medical School - Westmead, University of Sydney, NSW, Australia); Desmond Yip (Department of Medical Oncology, The Canberra Hospital, ACT, Australia; ANU Medical School, Australian National University, ACT, Australia); Nikolajs Zeps (St John of God Perth Northern Hospitals, Perth, WA, Australia).

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OCAC

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23andMe

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The Biobank Japan

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China Kadoorie Biobank

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MoBa

The Norwegian Mother, Father and Child Cohort Study (MoBa) is a population-based pregnancy cohort study conducted by the Norwegian Institute of Public Health. Participants were recruited from all over Norway from 1999-2008. The women consented to participation in 41% of the pregnancies. The cohort includes approximately 114.500 children, 95.200 mothers and 75.200 fathers. The current study is based on version 10 of the quality-assured data files released for research in Novel Tools for Early Childhood Predisposition to Obesity. The establishment of MoBa and initial data collection was based on a license from the Norwegian Data Protection Agency and approval from The Regional Committees for Medical and Health Research Ethics. The MoBa cohort is currently regulated by the Norwegian Health Registry Act. The current study was approved by The Regional Committees for Medical and Health Research Ethics (no. 2012/67). The Medical Birth Registry (MBRN) is a national health registry containing information about all births in Norway.

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Data from the Norwegian Mother, Father and Child Cohort Study and the Medical Birth Registry of Norway used in this study are managed by the national health register holders in Norway (Norwegian Institute of public health) and can be made available to researchers, provided approval from the Regional Committees for Medical and Health Research Ethics (REC), compliance with the EU General Data Protection Regulation (GDPR) and approval from the data owners. The consent given by the participants does not open for storage of data on an individual level in repositories or journals. Researchers who want access to data sets for replication should apply through helsedata.no. Access to data sets requires approval from The Regional Committee for Medical and Health Research Ethics in Norway and an agreement with MoBa.

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